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- (54) **H3 INFLUENZA A VIRUS**
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- (58) **Field of Classification Search**
None
See application file for complete search history.

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(57) **ABSTRACT**

The invention provides an isolated H3 equine influenza A virus, as well as methods of preparing and using the virus, and genes or proteins thereof.

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HAamino

MKTTIILILLTHWAYSQNPISGNNTATLCLGHHAVANGTLVKTISDDQIEVTNATE
LVQSISMGKICNNSYRILDGRNCTLIDAMLGDPHCDAFQYENWDLFIERSSAFSN
CYPYDIPDYASLRSIVASSGTLEFTAEGFTWTGVTQNGRSGACKRGSADSFPSRL
NWLTKSGSSYPTLNVTMPNKNFDKLYIWGIIHPSSNQEQTCLYIQESGRVTVST
KRSQQTIIPNIGSRPWVRGQSGRISYWTIVKPGDILMINSNGNLVAPRGYFKLKT
GKSSVMRSDVPIDICVSECTPNNGSISNDKPFQNVNKVTYGKCPKYIRQNTLKLAT
GMRNVPEKQIRGIFGALAGFIENGWEGMVDGWYGFYQNSEGTGQAADLKSTQ
AAIDQINGKLN RVIERTNEKFHQIEKEFSEVERRIQDLEKYVEDTKIDLWSYNAEL
LVALENQHTIDLTAEMNKLFEKTRRQLRENAEDMGGGCFKIYHKCDNACIGSI
RNGTYDHYIRDEALNNRFQIKGVELKSGYKDWILWISFAISCFLICVLLGFIM
WACQKGNIRCNICI

SEQ ID NO:1

FIG. 1A

NAamino

MNPNQKIIAIGFASLGILIIINVILHVVSIIIVTVLVLNNNRDLNCKGTIIREYNETVR
VEKITQWYNTSTIKYIERPSNEYMNNTEPLCEAQGFAPFSKDNGIRIGSRGHVVFV
IREPFVSCSPSECRFFLTQGSLLNDKHSNGTVKDRSPYRTLMSVKIGQSPNVYQA
RFESVAWSATAACHDGKKWMTVGVTGPDNQALAVVNYGGVPVDIINSWAGDILR
TQESSCTCIKGDCYWVMTDGFANRQAKYRIFKAKDGRVIGQTDISFNGGHIIECS
CYPNEGKVEICRDNWTGTNRPILVISSDLSYTVGYLCAGIPTDTPRGEDSQFTGS
CTSPLGNKCYGVKGFGFRQGTDVWAGRITSRTRSRSQFEIKIRNGWTQNSKDQIR
RQVIIDDPNWSGYSGSFTLPVELTKKGCLVPCFWVEMIRGKPEETTIWTSSSSIVM
CGVDHKIASWSWHDGAILPFDIDKM

SEQ ID NO:2

FIG. 1B

PBIamino

MDVNPTLLFLKVPQAISTTFPYTGDPPYSHGTGTGYTMDTVNRTHQYSEK GK
WTTNTEIGAPQLNPIDGPLPEDNEPSGYAQTDCVLEAMAFLEESHPIFENSCL
MEVIQQTRVDKLTQGRQTYDWTLNRNQPAAATALANTIEVFRSNGLTSNESGR
LMDFLKDVME SMNKEEMEITTHFQRKRVRDNMTKRMVTQRTIGKKKQRLNRKS
YLIRTLTLNTMTKDAERGKLRRAIATPGMQIRGFVYFVETLARRICEKLEQSG
L PVGGNEKKAKLANVVRKMMTNSQDTELSFTITGDNTKWNENQNPRIFLAMIT
YI TRNQPEWFRNVLSIAPIMFSNKMARLGKGYMFESKSMKLR TQIPAGMLASID
LK YFNDPTKKKIEKIRPLLVDGTASLSPGMMMGMFNMLSTVLCVLSILNLGQR
KYTK TTYWWDGLQSSDDFALIVNAPNHEGIQAGVDRFYRTCKLVGINMSKKK
SYINRT GTFEFTSFFYRYGFVANFSMELPSFGVSGINESADMSIGVTVIKNN
MINNDLGPAT AQMALQLFIKDYRYTYRCHRGDTQIQTRRSFELKKLWEQTR
SKTGLLVSDGGPN LYNIRNLHIPEVCLKWELMDEDYKGRLCNPLNPFVSHKEI
ESVNSAVVMPAHGP AKSMEYDAVATTHSWIPKRNR SILNTSQRGILEDEQMYQ
KCCNLFKFFPSSSYR RPVGISSMVEAMVSRARIDARIDFESGRICKDEFAEIM
KICSTIEELRRQK

SEQ ID NO:3

FIG. 1C

PB2amino

MERIKELRDLMLQSR TREILTKTTVDHMAIHKKYTSGRQEKNPALRMKWM
MAM KYPITADKRIMEMIPERNEQGQTLWSKTNDAGSDRVMVSPLAVTWWNR
NGPTT STIHYPKVYKTYFEKVERLKHGTFGPVHFRNQVKIRRRVDVNP
GHADLSAKEAQ DVIMEVVEFPNEVGARILTSESQLTITKEKKEELQDCKI
APLMVAAYMLERELVRKT RFLPVAGGTSSVYIEVLHLTQGTCWEQMYT
PGGEVRNDDIDQSLIIAARNIVRRA TVSADPLASLLEMCHSTQIGGIRM
VDILKQNPTEEQAVDICKAAMGLRISSSFSFG GFTFKRTSGSSVKREEE
MLTGNLQTLKIRVHEGYEEFTMVGRRATAILRKATRRL IQLIVSGRDEQ
SIAEAIIVAMVFSQEDCMIKAVRGDLN FVN RANQRLNPMHQLLR HFQK
DAKVL FQNWGIEPIDNVMGMIGILPDMPSTEMSLRGVRVSKMGVDEYSS
TERVVV SIDRFLRVRDQRGNILLSPEEVSETQGTEKLTIIYSSSMMWEI
NGPESVL VNTYQWII RNWEIVKIQWSQDPTMLYNKIEFEPFQSLVPRAT
RSQYSGFVRTLFQ QMRDVLGTFDTAQIHKLLPFAAAPPEQSRMQFSSLT
VNVV RGS GM RILVRGN SPVF NYNKATKRLTVL GK DAGALTEDPDEGT
AGVESAVLRGFLILGKENKRYGPALSI NELS KLAKGEKANVLIGQGDV
VLMKRKR DSSILTDSQTATKRIRMAIN

SEQ ID NO:4

FIG. 1D

PAmino

MEDFVRQCFNPMIVELAEKAMKEYGEDPKIETNKFAAICTHLEVCFMYSDFHFIN
 ELSSEVVIESGDPNALLKHRFEIIEGRDRTMAWTVVNSICNTTRAEEKPKFLPDLYD
 YKENRFVEIGVTRREVHIYYLEKANKIKSEKTHIHIFSFTGEEMATKADYTLDEES
 RARIKTRLFTIRQEMASRGLWDSFRQSERGEETIEERFEITGTMRKLANYSLPPNF
 SSLENFRVYVDGFEPNGCIESKLSQMSKEVNARIEPFSKTTPRPLKMPGGPPCHQR
 SKFLLMDALKLSIEDPSHEGEGIPLYDAIKCMKTFFGWKEPSIVKPHEKGINPNYL
 QTWKQVLAELQDLENEEKDPKTKNMKKTSQLKWALSENMAPEKVPDFEDCKDIS
 DLKQYDSDEPETRSLASWIQSEFNKACELTDSSWIELDEIGEDVAPIEYIASMRRN
 YFTAEVSHCRATEYIMKGVYINTALLNASCAAMDEFQLIPMISKCRTKEGRRKTN
 LYGFIVKGRSHLRNDTDVVNFVSMFSLTDPRFEPHKWEKYCVLEIGDMLLRTA
 VGQVSRPMFLYVRTNGTSKIKMKWGMEMRRCLLQSLQQIESMIEAESSVKEKD
 MTKEFFENKSETWPIGESPKGVVEEGSIGKVCRTLLAKSVFNSLYASPQLEGFSAES
 RKLLLIVQALRDNLEPGTFDIGGLYESIEECLINDPWVLLNASWFNSFLTHALK

SEQ ID NO:5

FIG. 1E

NPamino

MASQGTKRSYEQMETDGERQNAATEIRASVGRMVGGIGRFYVQMCTELKLNDE
 GRLIQNSITIERMVLSAFDERRNKYLEEHPSAGKDPKKTGGPIYRRKDQKWMREL
 ILHDKEEIMRIWRQANNGEDATAGLTHMMIWHSNLNDTTYQRTRALVRTGMDP
 RMCSLMQGSTLPRRSGAAGAAVKGVGTMVMELIRMIKRGINDRNFWROENGR
 RTRIAYERMCNILKGFQTAQAQRAMMDQVREGRNPGNAEIEDLIFLARSALILRG
 SVAHKSLPACVYGLAVTSGYDFEKEGYSLVGIDPFKLLQNSQIFSLIRPKENPAH
 KSQLVWMACHSAAFEDLRVLNFIRGTVIPRQQLTTRGVQIASNENMETIDSSTL
 ELRSKYWAIRTRSGGNTSQQRASAGQISVQPTFSVQRNLPFERATIMAAFTGNT
 GRTSDMRTEIIRMMENAKSEDVVSFQGRGVFELSDEKATNPVPSFDMSNEGSYFF
 GDNAEEFDS

SEQ ID NO:6

FIG. 1F

Mlamino

MSLLTEVETYVLSIVPSGPLKAEIAQRLEDVFAGKNTDLEALMEWLKTRPILSPLT
KGILGFVFTLTPSERGLQRRRFVQNALSGNGDPNNMDRAVKLYRKLKREITFH
GAKEVALSYSTGALASCMGLIYNRMGTVTTEVAFGLVCATCEQIADSQHRSHRQ
MVTTFNPLIRHENRMVLASTTAKAMEQMAGSSEQAAEAMEVASRARQMVQAM
RTIGTHPSSAGLKDDLENLQAYQKRMGVQMQRFK

SEQ ID NO:7

FIG. 1G

NSlamino

MDSNTVSSFQVDCFLWHVRKRFADQELGDAPFLDRLRRDQKSLRGRGSTLGLDI
ETATHAGKQIVEQILEKESDEALKMTIASVPTSRYLTDMTLDEMSRDWFMLMPK
QKVTGSLCIRMDQAIMDKNIILKANFSVIFERLETLILLRAFTEEGAVVGEISPLPSL
PGHTNEDVKNAIGVLIIGLKWNDNTVRISETLQRFAWRSSHENGRPSFPSKQKR
KMERTIKPKI

SEQ ID NO:8

FIG. 1H

HA

TCATGAAGACAACCATTATTTTGATACTACTGACCCATTGGGCTTACAGTCAA
AACCCAATCAGTGGCAACAACACAGCCACATTGTGTCTGGGACACCATGCAG
TAGCAAATGGAAACATTGGTAAAAACAATAAGTGATGATCAAATTGAGGTGAC
AAATGCTACAGAATTAGTTCAAAGCATTTC AATGGGGAAAATATGCAACAAC
TCATATAGAATTCTAGATGGAAGAAATTGCACATTAATAGATGCAATGCTAG
GAGACCCCACTGTGACGCCTTTCAGTATGAGAATTGGGACCTCTTTATAGAA
AGAAGCAGCGCTTTCAGCAATTGCTACCCATATGACATCCCTGACTATGCATC
GCTCCGATCCATTGTAGCATCCTCAGGAACATTGGAATTCACAGCAGAGGGA
TTCACATGGACAGGTGTCACTCAAAACGGAAGAAGTGGAGCCTGCAAAAAGG
GGATCAGCCGATAGTTTCTTTAGCCGACTGAATTGGCTAACAAAATCTGGAA
GCTCTTACCCACATTGAATGTGACAATGCCTAACAAATAAAAATTTGACAA
GCTATACATCTGGGGGATTCAACACCCGAGCTCAAATCAAGAGCAGACAAAA
TTGTACATCCAAGAATCAGGACGAGTAACAGTCTCAACAAAAAGAAGTCAAC
AAACAATAATCCCTAACATCGGATCTAGACCGTGGGTCAGAGGTCAATCAGG
TAGGATAAGCATATACTGGACCATTGTAAAACCTGGAGATATCCTAATGATA
AACAGTAATGGCAACTTAGTTGCACCCGCGGGGATATTTTAAATTGAAAACAG
GGAAAAGCTCTGTAATGAGATCAGATGTACCCATAGACATTTGTGTGTCTGA
ATGTATTACACCAAATGGAAGCATCTCCAACGACAAGCCATTCCAAAATGTG
AACAAAGTTACATATGGAAAATGCCCAAGTATATCAGGCAAAACACTTTAA
AGCTGGCCACTGGGATGAGGAATGTACCAGAAAAGCAAATCAGAGGAATCT
TTGGAGCAATAGCGGGATTCAACGAAAACGGCTGGGAAGGAATGGTTGATGG
GTGGTATGGGTTCCGATATCAAAACTCTGAAGGAACAGGGCAAGCTGCAGAT
CTAAAGAGCACTCAAGCAGCCATCGACCAGATTAATGGAAAAGTTAAACAGA
GTGATTGAAAGAACCAATGAGAAATTCATCAAAATAGAGAAGGAATTTCTCAG
AAGTAGAAAAGAAGAATTCAGGACTTGGAGAAATATGTAGAAGACACCAAAA
TAGACCTATGGTCCTACAATGCAGAATTGCTGGTGGCTCTAGAAAATCAACA
TACAATTGACTTAACAGATGCAGAAATGAATAAATLATTTGAGAAGACTAGA
CGCCAGTTAAGAGAAAACGCAGAAAGACATGGGAGGTGGATGTTTCAAGATTT
ACCACAAATGTGATAATGCATGCATTGGATCAATAAGAAATGGGACATATGA
CCATTACATATACAGAGATGAAGCATTAAACAACCGATTTTCAGATCAAAGGT
GTAGAGTTGAAATCAGGCTACAAAGATTGGATACTGTGGATTTTCATTGCGCA
TATCATGCTTCTTAATTTGCGTTGTICTATTGGGTTTCATTATGTGGGCTTGCC
AAAAAGGCAACATCAGATGCAACATTTCATTGAG

SEQ ID NO:9

FIG. 11

NA

ATGAATCCAAATCAAAGATAATAGCAATTGGATTTGCATCATTGGGGATAT
TAATCATTAAATGTCATTCTCCATGTAGTCAGCATTATAGTAACAGTACTGGTC
CTCAATAACAATAGAACAGATCTGAACTGCAAAGGGACGATCATAAGAGAG
TACAATGAAACAGTAAGAGTAGAAAAAATTACTCAATGGTATAATACCAGTA
CAATTAAGTACATAGAGAGACCTTCAAATGAATACTACATGAACAACACTGA
ACCACTTTGTGAGGCCCAAGGCTTTGCACCATTTTCAAAGATAATGGAATAC
GAATTGGGTCGAGAGGCCATGTTTTTGTGATAAGAGAACCTTTTGTATCATGT
TCGCCCTCAGAATGTAGAACCCTTTTCTCACACAGGGCTCATTACTCAATGA
CAAACATTCTAACGGCACAGTAAAGGACCGAAGTCCGTATAGGACTTTGATG
AGTGTCAAAATAGGGCAATCACCTAATGTATATCAAGCTAGGTTTGAATCGG
TGGCATGGTCAGCAACAGCATGCCATGATGGAAAAAATGGATGACAGTTGG
AGTCACAGGGCCCGACAATCAAGCAATTGCAGTAGTGAACCTATGGAGGTGT
CCGGTTGATATTATTAATTCATGGGCAGGGGATATTTTAAGAAACCAAGAAT
CATCATGCACCTGCATTAAGGAGACTGTTATTGGGTAATGACTGATGGACC
GCCAAATAGGCAAGCTAAATATAGGATATTCAAAGCAAAGATGGAAGAGT
AATTGGACAGACTGATATAAGTTTTCAATGGGGGACACATAGAGGAGTGTCT
TGTTACCCCAATGAAGGGAAGGTGGAATGCATATGCAGGGACAATTGGACTG
GAACAAATAGACCAATTCTGGTAATATCTTCTGATCTATCGTACACAGTTGGA
TATTTGTGTGCTGGCATTCCCCTGACACTCCTAGGGGAGAGGATAGTCAATT
CACAGGCTCATGTACAAGTCCTTTGGGAAATAAAAGGATACGGTGLAAAAGGT
TTCGGGTTTCGACAAGGAACTGACGTATGGGCCGGAAGGACAATTAGTAGGA
CTTCAAGATCAGGATTCGAAATAATAAAAAATCAGGAATGGTTGGACACAGAA
CAGTAAAGACCAAATCAGGAGGCAAGTGATTATCGATGACCCAAATTGGTCA
GGATATAGCGGTTCTTTCACATTGCCGTTGAACTAACAAAAAAGGGATGTT
TGGTCCCTGTTTCTGGGTTGAAATGATTAGAGGTAAACCTGAAGAAACAAC
AATATGGACCTCTAGCAGCTCCATTGTGATGTGTGGAGTAGATCATAAAATT
GCCAGTTGGTCATGGCACGATGGAGCTATTCTTCCCTTTGACATCGATAAGAT
GTAA

SEQ ID NO:10

FIG. 1J

PBI

ATGGATGTCAATCCGACTCTACTTTTTCTTAAAGGTGCCAGCGCAAATGCTAT
AAGCACAAACATTTCCCTTATACTGGAGATCCTCCCTACAGTCATGGAACAGGG
ACAGGATACACCATGGATACTGTCAACAGAACACACCAATATTCAGAAAAAG
GGAAATGGACAACAAACACTGAGATTGGAGCACCACAACCTTAATCCAATCGA
TGGACCACTTCCCTGAAGACAATGAACCAAGTGGGTACGCCCAAACAGATTGT
GTATTGGAAGCAATGGCTTTCCCTTGAAGAATCCCATCCCGGAATCTTTGAAAA
TTCGTGTCTTGAAACGATGGAGGTGATTCAGCAGACAAGAGTGGACAAACTA
ACACAAGGCCGACAAACTTATGATTGGACCTTGAATAGGAATCAACCTGCCG
CAACAGCACTTGCTAATACGATTGAAGTATTCAGATCAAATGGTCTGACTTCC
AATGAATCGGGGAGATTGATGGACTTCCCTCAAAGATGTCATGGAGTCCATGA
ACAAGGAAGAAATGGAAATAACAACACACTTCCAACGGAAGAGAAGAGTAA
GAGACAACATGACAAAGAGAATGGTAACACAGAGAACCATAGGGAAGAAAA
AACAAACGATTAACAGAAAAGAGCTATCTAATCAGAACATTAACCCTAAACAC
AATGACCAAGGACGCTGAGAGAGGGAAATTGAAACGACGAGCAATCGCTAC
CCCAGGOATGCAGATAAGAGGGTTTTGTATATTTTGTGAAACACTAGCCCGA
AGAATATGTGAAAAGCTTGAACAATCAGGATTGCCAGTTGGCGGTAATGAGA
AAAAGGCCAAACTGGCTAATGTCGTGAGAAAAATGATGACTAATTCCCAAGA
CACTGAACTCTCCTTACCATCACTGGGGACAATACCAAATGGAATGAAAAT
CAGAACCCACGCATATTCCTGGCAATGATCACATACATAACTAGAAACCAGC
CAGAATGGTTCAGAAATGTTCTAAGCATTGCACCGATTATGTTCTCAAATAAA
ATGGCAAGACTGGGGAAAGGATATATGTTTGAAGCAAAGTATGAAATTG
AGAACTCAAATACCAGCAGGAATGCTTGCAAGCATTGACCTGAAATATTTCA
ATGATCCAACAAAAAAGAAAATTGAAAAGATACGACCCTTCTGGTTGACGG
GACTGCTTCACTGAGTCCTGGCATGATGATGGGAATGTTCAACATGTTGAGC
ACTGTGCTAGGTGTATCCATATTAACCTGGGCCAGAGGAAATACACAAAGA
CCACATACTGGTGGGATGGTCTGCAATCATCCGATGACTTTGCTTTGATAGTG
AATGCGCCTAATCATGAAGGAATACAAGCTGGAGTAGACAGATTCTATAGGA
CTTGCAAACCTGGTCGGGATCAACATGAGCAAAAAGAAGTCTACATAAATAG
AACTGGAACATTGGAATTCACAAGCTTTTTCTACCGGTATGGTTTTGTAGCCA
ATTTGAGCATGGAACCTACCAGTTTTGGGGTTTTCCGGAATAAATGAATCTGCA
GACATGAGCATTGGAGTGACAGTCATCAAAAACAACATGATAAATAATGATC
TCGGTCTGCCACGGCACAAATGGCACTCCAACCTTTCATTAAGGATTATCGG
TACACATACCGGTGCCATAGAGGTGATACCCAGATACAAACCAGAAGATCTT
TTGAGTTGAAGAACTGTGGGAACAGACTCGATCAAAGACTGGTCTACTGGT
ATCAGATGGGGGTCCAAACCTATATAACATCAGAAACCTACACATCCCGGAA
GTCTGTTTAAAATGGGAGCTAATGGATGAAGATTATAAGGGGAGGCTATGCA
ATCCATTGAATCCTTTTCGTTAGTCACAAAGAAATTGAATCAGTCAACAGTGCA
GTAGTAATGCCTGCGCATGGCCCTGCCAAAAGCATGGAGTATGATGCTGTTG
CAACAACACATTCTTGGATCCCCAAGAGGAACCGGTCCATATTGAACACAAG
CCAAAGGGGAATACTCGAAGATGAGCAGATGTATCAGAAATGCTGCAACCTG
TTTGAAAAATTTTCCCAGCAGCTCATACAGAAGACCAGTCGGGATTTCTAG
TATGGTTGAGGCCATGGTGTCCAGGGCCCCGCATTGATGCACGAATTGACTTC
GAATCTGGACGGATAAAGAAGGATGAGTTCGCTGAGATCATGAAGATCTGTT
CCACCATTGAAGAGCTCAGACGGCAAAAATAGTGA

SEQ ID NO:11

FIG. 1K

PB2

ATGGAGAGAATAAAAAGAAGCTGAGAGATCTGATGTTACAATCCCGCACCCGCG
AGATACTAACAAAACTACTGTGGACCACATGGCCATAATCAAGAAATACAC
ATCAGGAAGACAAGAGAAGAACCCTGCACTTAGGATGAAATGGATGATGGC
AATGAAATACCCAATTACAGCAGATAAAGAGGATAATGGAGATGATTCCTGAG
AGAAATGAACAGGGACAAACCCTTTGGAGCAAACGAACGATGCTGGCTCA
GACCGCGTAATGGTATCACCTCTGGCAGTGACATGGTGGAAATAGGAATGGAC
CAACAACAAGCACAATTCATTATCCAAAAGTCTACAAAACCTTATTTTGAAAA
GGTTGAAAGATTGAAACACGGAACCTTTGGCCCCGTTTATTTTAGGAATCAA
GTCAAGATAAGACGAAGAGTTGATGTAAACCCTGGTCACGCGGACCTCAGTG
CCAAAGAAGCACAAGATGTGATCATGGAAGTTGTTTTCCCAAATGAAGTGGG
AGCCAGAATTCTAACATCGGAATCACAACCTAACAAATAACCAAAGAGAAAA
GGAAGAAGCTTCAGGACTGCAAAATTGCTCCCTTGATGGTAGCATAACATGCTA
GAAAGAGAGTTGGTCCGAAAAACAAGGTTCTCCAGTAGCAGGCGGAACA
AGCAGTGTATACATTGAAGTGTTCATCTGACTCAGGGAACATGCTGGGAGC
AAATGTACACCCCAGGAGGAGAAGTTAGAAACGATGATATTGATCAAAGTTT
AATTATTGCAGCCCGGAACATAGTGAGAAGAGCAACAGTATCAGCAGATCCA
CTAGCATCCCTACTGGAAATGTGCCACAGTACACAGATTGGTGGAAATAAGGA
TGGTAGACATCCTTAAGCAGAATCCAACAGAGGAACAAGCTGTGGATATATG
CAAAGCAGCAATGGGATTGAGAATTAGCTCATCATTGAGCTTTGGTGGATT
ACCTTCAAGAGAACAAAGTGGATCATCAGTCAAGAGAGAAGAAGAAATGCTT
ACGGGCAACCTTCAAACATTGAAAATAAGAGTGCATGAGGGCTATGAAGAAT
TCACAATGGTTCGGAAGAAGAGCAACAGCCATTCTCAGAAAGGCAACCAGAA
GATTGATTCAATTGATAGTAAGTGGGAGAGATGAACAGTCAATTGCTGAAGC
AATAATTGTAGCCATGGTGTTCGCAAGAAGATTGCATGATAAAAGCAGTT
CGAGGCGATTTGAACTTTGTTAATAGAGCAAATCAGCGCTTGAACCCCATGC
ATCAACTCTTGAGGCATTTCCAAAAGGATGCAAAAAGTGTTCCTTCCAAAATTG
GGGGATTGAACCCATCGACAATGTAATGGGAATGATTGGAATATTGCCTGAC
ATGACCCCAAGCACCAGAGATGTCATTGAGAGGAGTGAGAGTCAGCAAAATG
GGAGTGGATGAGTACTCCAGCACTGAGAGAGTGGTGGTGGAGCATTGACCGTT
TTTTAAGAGTTCGGGATCAAAGGGGAAACATACTACTGTCCCCTGAAGAAGT
CAGTGAAACACAAGGAACGGAAAAGCTGACAATAATTTATTCGTCAATCAATG
ATGTGGGAGATTAATGGTCCCGAATCAGTGTGGTCAATACTTATCAATGGAT
CATCAGGAAGTGGAAATTTGTAATAATTCAGTGGTACAGGACCCCAATG
TTATAACAATAAGATAGAATTTGAGCCATTCCAATCCCTGGTCCCTAGGGCTAC
CAGAAGCCAATACAGCGGTTTCGTAAGAACCCTGTTTCAGCAAATGCGAGAT
GTACTTGGAACATTTGATACTGCTCAAATAATAAACTCCTCCCTTTTGCCGC
TGCTCCCTCCGGAACAGAGTAGGATGCAGTTCCTCTTTGACTGTTAATGTAA
GAGGTTCCGGAATGAGGATACTTGTAAAGAGGCAATTCCCCAGTGTTCAACTA
CAATAAAGCCACTAAAAGGCTCACAGTCCCTCGGAAAGGATGCAGGTGCGCTT
ACTGAGGACCCAGATGAAGGTACGGCTGGAGTAGAATCTGCTGTTCTAAGAG
GGTTTCTCATTTTAGGTAAAGAAAATAAGAGATATGGCCCAGCACTAAGCAT
CAATGAACTAAGCAAACCTTGCAAAAGGGGAGAAAGCCAATGTACTAATTGG
GCAAGGGGACGTAGTGTGGTAATGAAACGGAAACGTGACTCTAGCATACTT
ACTGACAGCCAGACAGCGACCAAAAGGATTTCGGATGGCCATCAATTAGT

SEQ ID NO:12

FIG. 1L

PA

ATGGAAGACTTTGTGCGACAATGCTTCAATCCAATGATCGTCGAGCTTGCGG
AAAAGGCAATGAAAGAATATGGAGAGGACCCGAAAATCGAAACAAAACAAAT
TTGCAGCAATATGCACTCACTTGGAAAGTCTGCTTCATGTACTCGGATTTCCAC
TTTATTAATGAACTGAGTGAGTCAGTGGTTCATAGAGTCTGGTGACCCAAATG
CTCTTTTGAAACACAGATTTGAAATCATTGAGGGGAGAGATCGAACAATGGC
ATGGACAGTAGTAAACAGCATCTGCAACACCACAAGAGCTGAAAAACCTAA
ATTTCTTCCAGATTTATACGACTATAAGGAGAACAGATTTGTTGAAATTGGTG
TGACAAGGAGAGAAGTTCACATATACTACCTGGAGAAGGCCAACAAAATAA
AGTCTGAGAAAACACATATCCACATTTTCTCATTTACAGGAGAGGAAATGGC
TACAAAAGCGGACTATACTCTTGATGAAGAGAGTAGAGCCAGGATCAAGACC
AGACTATTCACTATAAGACAAGAAATGGCCAGTAGAGGCCTCTGGGATTCTT
TTCGTCAGTCCGAGAGAGGGCGAAGAGACAATTGAAGAAAGATTTGAAATCAC
AGGGACGATGCGCAAGCTTGCCAATTACAGTCTCCCACCGAACTTCTCCAGC
CTTGAAAATTTTAGAGTCTATGTGGATGGATTCGAACCGAACGGCTGCATTG
AGAGTAAGCTTTCTCAAATGTCCAAGAAGTAAATGCCAGAATCGAACCCATT
TTCAAAGACAACACCCCGACCACTCAAATGCCAGGTGGTCCACCCTGCCAT
CAGCGATCTAAATTCCTGCTAATGGATGCTCTGAAACTGAGCATTGAGGACC
CAAGTCACGAGGGAGAGGGAATACCACTATATGATGCCATCAAATGCATGAA
AACTTTCTTTGGATGGAAAAGAGCCCAGTATTGTTAAACCACATGAAAAGGGT
ATAAACCCGAACATCTCCAAACTTGGAAAGCAAGTATTAGCAGAATTACAAG
ACCTTGAGAACGAAGAAAAGGACCCCAAGACCAAGAATATGAAAAAAACAA
GCCAATTGAAATGGGCACTTAGTGAAAATATGGCACCAGAGAAAGTGGATTT
TGAGGATTGTAAGACATCAGTGATTTAAAACAGTATGACAGTGATGAGCCA
GAAACAAGGTCTCTTGCAAGTTGGATTCAAAGTGAGTTCAACAAAGCTTGTG
AACTGACAGATTCAAGCTGGATAGAGCTCGATGAAATTGGGGAGGATGTTGC
CCCAATAGAATACATTGCGAGCATGAGGAGAAATTATTTTACTGCTGAGGTT
TCCCATTGTAGAGCAACAGAATATATAATGAAGGGAGTGTACATCAACACTG
CTCTACTCAATGCATCCTGTGCTGCGATGGATGAATTCCAATTAATTCCGATG
ATAAGTAAATGCAGGACCAAAGAAGGGAGAAGGAAGACAAATTTATATGGA
TTCATAGTAAAGGGAAGGTCCCATTAAAGAAATGATACTGACGTGGTGAAC
TTGLAAGTATGGAATTTTCTCTCACTGATCCAAGATTTGAGCCACACAAATGG
GAAAAATACTGCGTTCTAGAAATTGGAGACATGCTTCTAAGAACTGCTGTAG
GTCAAGTGTCAAGACCCATGTTTTTGTATGTAAGGACAAATGGAACCTCTAA
AATTAATGAAATGGGGAATGGAAATGAGGCGCTGCCTCCTTCAGTCTCTG
CAACAGATTGAAAGCATGATCGAAGCTGAGTCCCTCAGTCAAAGAAAAGGAC
ATGACCAAAGAATTTTTTGAGAACAAATCAGAGACATGGCCTATAGGAGAGT
CCCCAAAGGAGTGGAAAGAGGGCTCAATCGGGAAGGTTTGCAGGACCTTATT
AGCAAAATCTGTGTTAACAGTTTGTATGCATCTCCACAACCTGGAAGGGTTTT
CAGCTGAATCTAGGAAATTACTTCTCATTGTTCAAGGCTCTTAGGGATAACCTG
GAACCTGGAACCTTTGATATTGGGGGGTTATATGAATCAATTGAGGAGTGCC
TGATTAATGATCCCTGGGTTTTGCTTAATGCATCTTGGTTCAACTCCTTCCTTA
CACATGCACTGAAGTAGTTGTGGCAATGCTACTATTTGCTATCCATACTGTCC
AAAAAAGTACCTTGTCTACT

SEQ ID NO:13

FIG. 1M

NP

ATGGCGTCTCAAGGCACCAAACGATCCTATGAACAGATGGAAACTGATGGGG
AACGCCAGAATGCAACTGAAATCAGAGCATCTGTCCGAAGGATGGTGGGAG
GAATCGGCCGGTTTTATGTTTCAGATGTGTACTGAGCTTAAACTAAACGACCAT
GAAGGGCGGCTGATTCAGAACAGCATAACAATAGAAAGGATGGTACTTTTCGG
CATTCCGACGAAAGAAGAAAACAAGTATCTCGAGGAGCATCCCAGTGCTGGGA
AAGACCCTAAGAAAACAGGAGGCCCGATATACAGAAGGAAAGATGGGAAAT
GGATGAGGGAACTCATCCTCCATGATAAAGAAGAAATCATGAGAATCTGGCG
TCAGGCCAACAAATGGTGAAGACGCTACTGCTGGTCTTACTCATATGATGATCT
GGCACTCCAATCTCAATGACACCACATACCAAAGAACAAGGGCTCTTGTTTCG
GACTGGGATGGATCCCAGAATGTGCTCTCTGATGCAAGGCTCAACCCTCCCA
CGGAGATCTGGAGCCGCTGGTGCTGCAGTAAAAGGTGTTGGAACAATGGTAA
TGGAACTCATCAGAATGATCAAACGCGGAATAAATGATCGGAATTTCTGGAG
AGGTGAAAATGGTCGAAGAACCAGAATTGCTTATGAAAGAATGTGCAATATC
CTCAAAGGGAAATTTTCAGACAGCAGCACAACGGGCTATGATGGACCAGGTG
AGGGAAAGGCCGCAATCCTGGAAACGCTGAGATTGAGGATCTCATTTTCTTGG
CACGATCAGCACTTATTTTGAGAGGATCAGTAGCCATAAATCATGCCTACCT
GCCTGTGTTTATGGCCTTGCAGTAACCAGTGGGTATGACTTTGAGAAGGAAG
GATACTCTCTGGTTGGAATTGATCCTTTCAAACACTCCAGAACAGTCAAATT
TTCAGTCTAATCAGACCAAAGAAAACCCAGCACACAAGAGCCAGTTGGTGT
GGATGGCATGCCATTCTGCAGCATTGAGGACCTGAGAGTTTTAAATTTCAAT
AGAGGAACCAAAGTAATCCCAAGAGGACAGTTAACAACCAGAGGAGTTCAA
ATAGCTTCAAATGAAAACATGGAGACAATAGATTCTAGCACACTTGAACCTGA
GAAGCAAATATTGGGCAATAAGGACCAGAAGCGGAGGAAACACCAGTCAAC
AGAGAGCATCTGCAGGACAGATAAGTGTGCAACCTACTTTCTCAGTACAGAG
AAATCTTCCCTTTGAGAGAGCAACCATTATGGCTGCATTCAGTGGTAACACTG
AAGGGAGGACTTCCGACATGAGAACGGAAATCATAAGGATGATGGAAAATG
CCAAATCAGAAGATGTGTCTTTCCAGGGGCGGGGAGTCTTCGAGCTCTCGGA
CGAAAAGGCAACGAACCCGATCGTGCCTTCTTTGACATGAGCAATGAAGGG
TCTTATTTCTTCGGAGACAATGCTGAGGAGTTTGACAGTTAAA

SEQ ID NO:14

FIG. 1N

M

ATGAGTCTTCTAACCGAGGTCGAAACGTACGTTCTCTCTATCGTACCATCAGG
CCCCCTCAAAGCCGAGATCGCGCAGAGACTTGAAGATGTCTTTGCAGGGAAG
AACACCGATCTTGAGGCACTCATGGAATGGCTAAAGACAAGACCAATCCTGT
CACCTCTGACTAAAGGGATTTTAGGATTTGTATTCACGCTCACCGTGCCCAGT
GAGCGAGGACTGCAGCGTAGACGCTTTGTCCAAATGCCCTFAGTGGAACG
GAGATCCAAACAACATGGACAGAGCAGTAAAACGTACAGGAAGCTTAAAA
GAGAAATAACATTCCATGGGGCAAAAAGAGGTGGCACTCAGCTATTCCACTGG
TGCCTAGCCAGCTGCATGGGACTCATATACAACAGAATGGGAACCTGTTACA
ACCGAAGTGGCATTGGCCTGGTATGCGCCACATGTGAACAGATTGCTGATT
CCCAGCATCGGTCTCACAGGCAGATGGTGACAACAACCAACCCATTAATCAG
ACATGAAAACAGAATGGTATTAGCCAGTACCACGGCTAAAGCCATGGAACA
GATGGCAGGATCGAGTGAGCAGGCAGCAGAGGCCATGGAGGTTGCTAGTAG
GGCTAGGCAGATGGTACAGGCAATGAGAACCATTGGGACCCACCCTAGCTCC
AGTGCCGGTTTGAAAGATGATCTCCTTGAAAATTTACAGGCCTACCAGAAAC
GGATGGGAGTGCAAATGCAGCGATTCAAGTGATCCTCTCGTTATTGCAGCAA
GTATCATTGGGATCTTGCACTTGATATTGTGGATTCTTGATCGICTTTTCTTCA
AATTCATTTATCGTCGCCTTAAATACGGGTTGAAAAGAGGGCCTTCTACGGA
AGGAGTACCTGAGTCTATGAGGGAAGAATATCGGCAGGAACAGCAGAATGC
TGTGGATGTTGACGATGGTCATTTGTCAACATAGAGCTGGAGTAA

SEQ ID NO:15

FIG. 10

NS

ATGGATTCCAACACTGTGTCAAGCTTTCAGGTAGACTGTTTTCTTTGGCATGT
CCGCAAACGATTTCGCAGACCAAGAAGTGGGTGATGCCCCATTCCCTTGACCGG
CTTCGCCGAGACCAGAAGTCCCTAAGGGGAAGAGGTAGCACTCTTGGICTGG
ACATCGAAACAGCCACTCATGCAGGAAAGCAGATAGTGGAGCAGATTCTGG
AAAAGGAATCAGATGAGGCACTTAAAATGACCATTGCCTCTGTTCCCTACTTC
ACGCTACTTAACTGACATGACTCTTGATGAGATGTCAAGAGACTGGTTCATGC
TCATGCCCAAGCAAAAAGTAAACAGGCTCCCTATGTATAAGAATGGACCAGGC
AATCATGGATAAGAACATCATACTTAAAGCAAACCTTTAGTGTGATTTTCGAA
AGGCTGGAAACACTAATACTACTTAGAGCCTTCACCGAAGAAGGAGCAGTCG
TTGGCGAAATTCACCATTACCTTCTCTTCCAGGACATACTAATGAGGATGTC
AAAAATGCAATTGGGGTCCCTCATCGGAGGACTTAAATGGAATGATAATACGG
TTAGAATCTCTGAAACTCTACAGAGATTCGCTTGGAGAAGCAGTCATGAGAA
TGGGAGACCTTCATTCCCTTCAAAGCAGAAAACGAAAAATGGAGAGAACAATT
AAGCCAAAAATTTGAAGAAATAAGATGGTTGATTGAAGAAGTGGCAGATAG
ATTGAAAAATACAGAAAATAGTTTTGAACAAAATAACATTTATGCAAGCCTTA
CAACTATTGCTTGAAGTAGAACAAGAGATAAGAACTTCTCGTTTCAGCTTAT
TTAA

SEQ ID NO:16

FIG. 1P

M2amino

MSLLTEVETPTRNGWECKCSDSSDPLVIAASHIGILHLILWILDRLFFKFIYRRLKY
GLKRGPSTEGVPESMREEYRQEQQNAVDVDDGHFVNIELE

SEQ ID NO:17

FIG. 1Q

NS2amino

MDSNTVSSFQLMRMSKMQLGSSSEDLNGMIIRLESCLKLYRDSLGEAVMRMGDL
HSLQSRNEKWREQLSQKFEEIRWLIIEVVRHRLKNTENSFEQITFMQALQLLLEVE
QEIRTFSFQLI

SEQ ID NO:18

FIG. 1R

MKTTIILLILLTHWAYSQNPISGNNTATLCL A/Equine/WI/1/03
MKTTIILLILLTHWAYSQNPISGNNTATLCL A/Equine/New York/99

GHHAVANGTLVKTISDDQIEVTNATELVQS A/Equine/WI/1/03
GHHAVANGTLVKTISDDQIEVTNATELVQS A/Equine/New York/99

ISMGKICNNSYRILDGRNCTLIDAMLGDPH A/Equine/WI/1/03
ISMGKICNNSYRILDGRNCTLIDAMLGDPH A/Equine/New York/99

CDAPQYENWDLPIERSSSAFSNCYPYDIPDY A/Equine/WI/1/03
CD[V]FQYENWDLPIERSSSAFSNCYPYDIPDY A/Equine/New York/99

ASLR SIVASSGTLEFTAEGFTWTGVTQNGR A/Equine/WI/1/03
ASLR SIVASSGTLEFTAEGFTWTGVTQNGR A/Equine/New York/99

SGACKRGSADSFFSRLNWLTKSSGSSYPTLN A/Equine/WI/1/03
SGACKRGSADSFFSRLNWLTKSSG[N]SYPTLN A/Equine/New York/99

VTMPNNKNFDKLYIWIHHPSSNQEQTKLY A/Equine/WI/1/03
VTMPNNKNFDKLYIWIHHPSSNQEQTKLY A/Equine/New York/99

IQESGRVTVSTKRSSQQTIIIPNIGSRPWVVG A/Equine/WI/1/03
IQESGRVTVSTKRSSQQTIIIPNIGSRPWVVG A/Equine/New York/99

QSGRISIIYWTIVKPGDILMINSNGNLVAPR A/Equine/WI/1/03
QSGRISIIYWTIVKPGDILMINSNGNLVAPR A/Equine/New York/99

GYFKLKTGKSSVMRSDVPIDICVSECITPN A/Equine/WI/1/03
GYFKLKTGKSSVMRSD[A]PIDICVSECITPN A/Equine/New York/99

GSISNDKPFQNVNKVTYGKCPKYIRQNTLK A/Equine/WI/1/03
GSISNDKPFQNVNKVTYGKCPKYIRQNTLK A/Equine/New York/99

LATGMRRNVPEKQIE A/Equine/WI/1/03
LATGMRRNVPEKQIE A/Equine/New York/99

FIG. 2

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H3 INFLUENZA A VIRUS**CROSS-REFERENCE TO RELATED APPLICATION**

This application is a continuation of Ser. No. 14/840,759, filed Aug. 31, 2015, which is a continuation of U.S. patent application Ser. No. 14/255,719, filed Apr. 17, 2014, which is a continuation of U.S. patent application Ser. No. 13/839,111, filed Mar. 15, 2013, which is a continuation of U.S. patent application Ser. No. 12/503,712, filed Jul. 15, 2009, which is a divisional of U.S. patent application Ser. No. 11/033,248, filed Jan. 11, 2005, which applications are incorporated herein by reference.

STATEMENT OF GOVERNMENT RIGHTS

This invention was made with government support under 2001-35204-10184 awarded by the USDA/NIFA. The government has certain rights in the invention.

BACKGROUND

Influenza is a major respiratory disease in some mammals including horses and is responsible for substantial morbidity and economic losses each year. In addition, influenza virus infections can cause severe systemic disease in some avian species, leading to death. The segmented nature of the influenza virus genome allows for reassortment of segments during virus replication in cells infected with two or more influenza viruses. The reassortment of segments, combined with genetic mutation and drift, can give rise to a myriad of divergent strains of influenza virus over time. The new strains exhibit antigenic variation in their hemagglutinin (HA) and/or neuraminidase (NA) proteins, and in particular the gene coding for the HA protein has a high rate of variability. The predominant current practice for the prevention of flu is vaccination. Most commonly, whole virus vaccines are used. As the influenza HA protein is the major target antigen for the protective immune responses of a host to the virus and is highly variable, the isolation of influenza virus and the identification and characterization of the HA antigen in viruses associated with recent outbreaks is important for vaccine production. Based on prevalence and prediction, a vaccine is designed to stimulate a protective immune response against the predominant and expected influenza virus strains (Park et al., 2004).

There are three general types of influenza viruses, Type A, Type B and Type C, which are defined by the absence of serological crossreactivity between their internal proteins. Influenza Type A viruses are further classified into subtypes based on antigenic and genetic differences of their glycoproteins, the HA and NA proteins. All the known HA and NA subtypes (H1 to H15 and N1 to N9) have been isolated from aquatic birds, which are thought to act as a natural reservoir for influenza. H7N7 and H3N8 Type A viruses are the most common causes of equine influenza, and those subtypes are generally incorporated into equine influenza vaccines.

Thus, there is a continuing need to isolate new influenza virus isolates, e.g., for vaccine production.

SUMMARY OF THE INVENTION

The invention provides isolated H3 equine derived influenza type A virus that was isolated from a foal that succumbed to a fatal pneumonia, which virus has characteristic

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substitutions at residues 78 and 159 of HA (numbering of positions is that in the mature protein which lacks a 15 amino acid signal peptide), i.e., the residue at position 78 of HA is not valine and the residue at position 159 is not asparagine. In one embodiment, the isolated H3 influenza A virus of the invention has a conservative substitution at residue 78, e.g., a valine to an alanine substitution, and a nonconservative substitution at residue 159, e.g., an asparagine to a serine substitution. In one embodiment, the isolated H3 influenza A virus of the invention has a residue other than methionine at position 29, e.g., a nonconservative substitution, a residue other than lysine at position 54, e.g., a nonconservative substitution, a residue other than serine at position 83, e.g., a nonconservative substitution, a residue other than asparagine at position 92, e.g., a nonconservative substitution, a residue other than leucine at position 222, e.g., a nonconservative substitution, a residue other than alanine at position 272, e.g., a conservative substitution, and/or a residue other than threonine at position 328, e.g., a conservative substitution. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine and tryptophan; a group of amino acids having basic side chains is lysine, arginine and histidine; and a group of amino acids having sulfur-containing side chain is cysteine and methionine. In one embodiment, conservative amino acid substitution groups are: threonine-valine-leucine-isoleucine-alanine; phenylalanine-tyrosine; lysine-arginine; alanine-valine; glutamic-aspartic; and asparagine-glutamine.

In one embodiment, the influenza virus of the invention includes one or more viral proteins (polypeptides) having substantially the same amino acid sequence as one of SEQ ID NOs:1-8, 17 and/or 18, so long as the HA has the characteristic substitutions at residues 78 and 159. An amino acid sequence which is substantially the same as a reference sequence has at least 95%, e.g., 96%, 97%, 98% or 99%, amino acid sequence identity to that reference sequence, and may include sequences with deletions, e.g., those that result in a deleted viral protein having substantially the same activity or capable of being expressed at substantially the same level as the corresponding full-length, mature viral protein, insertions, e.g., those that result in a modified viral protein having substantially the same activity or capable of being expressed at substantially the same level as the corresponding full-length, mature viral protein, and/or substitutions, e.g., those that result in a viral protein having substantially the same activity or capable of being expressed at substantially the same level as the reference protein. In one embodiment, the one or more residues which are not identical to those in the reference sequence may be conservative or nonconservative substitutions which one or more substitutions do not substantially alter the expressed level or activity of the protein with the substitution(s), and/or the level of virus obtained from a cell infected with a virus having that protein. As used herein, "substantially the same expressed level or activity" includes a detectable protein level that is about 80%, 90% or more, the protein level, or a measurable activity that is about 30%, 50%, 90%, e.g., up to 100% or more, the activity, of a full-length mature polypeptide corresponding to one of SEQ ID NOs:1-8, 17 or 18. In one embodiment, the virus comprises a polypeptide

with one or more, for instance, 2, 5, 10, 15, 20 or more, amino acid substitutions, e.g., conservative substitutions of up to 5% of the residues of the full-length, mature form of a polypeptide having SEQ ID NOs:1-8, 17 or 18. The isolated virus of the invention may be employed alone or

with one or more other virus isolates, e.g., other influenza virus isolates, in a vaccine, to raise virus-specific antisera, in gene therapy, and/or in diagnostics. Accordingly, the invention provides host cells infected with the virus of the invention, and isolated antibody specific for the virus.

The invention also provides an isolated nucleic acid molecule (polynucleotide) comprising a nucleic acid segment corresponding to at least one of the proteins of the virus of the invention, a portion of the nucleic acid segment for a viral protein having substantially the same level or activity as a corresponding polypeptide encoded by one of SEQ ID NOs:1-8, 17 or 18, or the complement of the nucleic acid molecule. In one embodiment, the isolated nucleic acid molecule encodes a polypeptide which has substantially the same amino acid sequence, e.g., has at least 95%, e.g., 96%, 97%, 98% or 99%, contiguous amino acid sequence identity to a polypeptide having one of SEQ ID NOs:1-8, 17 or 18. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence which is substantially the same as, e.g., has at least 50%, e.g., 60%, 70%, 80% or 90% or more, contiguous nucleic acid sequence identity to, one of SEQ ID NOs:9-16, or the complement thereof, and encodes a polypeptide having at least 95%, e.g., 96%, 97%, 98% or 99%, contiguous amino acid sequence identity to a polypeptide having one of SEQ ID NOs:1-8, 17 or 18.

The isolated nucleic acid molecule of the invention may be employed in a vector to express influenza proteins, e.g., for recombinant protein vaccine production or to raise antisera, as a nucleic acid vaccine, for use in diagnostics or, for vRNA production, to prepare chimeric genes, e.g., with other viral genes including other influenza virus genes, and/or to prepare recombinant virus, e.g., see Neumann et al. (1999) which is incorporated by reference herein. Thus, the invention also provides isolated viral polypeptides, recombinant virus, and host cells contacted with the nucleic acid molecule(s) and/or recombinant virus of the invention, as well as isolated virus-specific antibodies, for instance, obtained from mammals infected with the virus or immunized with an isolated viral polypeptide or polynucleotide encoding one or more viral polypeptides.

The invention further provides at least one of the following isolated vectors, for instance, one or more isolated influenza virus vectors, or a composition comprising the one or more vectors: a vector comprising a promoter operably linked to an influenza virus PA DNA for a PA having substantially the same amino acid sequence as SEQ ID NO:5 linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus PB1 DNA for a PB1 having substantially the same amino acid sequence as SEQ ID NO:3 linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus PB2 DNA for a PB2 having substantially the same amino acid sequence as SEQ ID NO:4 linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus HA DNA for a HA having substantially the same amino acid sequence as SEQ ID NO:1 linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus NP DNA for a NP having substantially the same amino acid sequence as SEQ ID NO:6

NA DNA for a NA having substantially the same amino acid sequence as SEQ ID NO:2 linked to a transcription termination sequence, a vector comprising a promoter operably linked to an influenza virus M DNA for a M having substantially the same amino acid sequence as SEQ ID NO:7 (M1) and/or SEQ ID NO:17 (M2), linked to a transcription termination sequence, and/or a vector comprising a promoter operably linked to an influenza virus NS DNA for a NS having substantially the same amino acid sequence as SEQ ID NO:8 (NS1) and/or SEQ ID NO:18 (NS2), linked to a transcription termination sequence. Optionally, two vectors may be employed in place of the vector comprising a promoter operably linked to an influenza virus M DNA linked to a transcription termination sequence, e.g., a vector comprising a promoter operably linked to an influenza virus M1 DNA linked to a transcription termination sequence and a vector comprising a promoter operably linked to an influenza virus M2 DNA linked to a transcription termination sequence. Optionally, two vectors may be employed in place of the vector comprising a promoter operably linked to an influenza virus NS DNA linked to a transcription termination sequence, e.g., a vector comprising a promoter operably linked to an influenza virus NS1 DNA linked to a transcription termination sequence and a vector comprising a promoter operably linked to an influenza virus NS2 DNA linked to a transcription termination sequence. An influenza virus vector is one which includes at least 5' and 3' noncoding influenza virus sequences.

Hence, the invention provides vectors, e.g., plasmids, which encode influenza virus proteins, and/or encode influenza vRNA, both native and recombinant vRNA. Thus, a vector of the invention may encode an influenza virus protein (sense) or vRNA (antisense). Any suitable promoter or transcription termination sequence may be employed to express a protein or peptide, e.g., a viral protein or peptide, a protein or peptide of a nonviral pathogen, or a therapeutic protein or peptide. In one embodiment, to express vRNA, the promoter is a RNA polymerase I promoter, a RNA polymerase II promoter, a RNA polymerase III promoter, a T3 promoter or a T7 promoter. Optionally the vector comprises a transcription termination sequence such as a RNA polymerase I transcription termination sequence, a RNA polymerase II transcription termination sequence, a RNA polymerase III transcription termination sequence, or a ribozyme.

A composition of the invention may also comprise a gene or open reading frame of interest, e.g., a foreign gene encoding an immunogenic peptide or protein useful as a vaccine. Thus, another embodiment of the invention comprises a composition of the invention as described above in which one of the influenza virus genes in the vectors is replaced with a foreign gene, or the composition further comprises, in addition to all the influenza virus genes, a vector comprising a promoter linked to 5' influenza virus sequences linked to a desired nucleic acid sequence, e.g., a cDNA of interest, linked to 3' influenza virus sequences linked to a transcription termination sequence, which, when contacted with a host cell permissive for influenza virus replication optionally results in recombinant virus. In one embodiment, the DNA of interest is in an antisense orientation. The DNA of interest, whether in a vector for vRNA or protein production, may encode an immunogenic epitope, such as an epitope useful in a cancer therapy or vaccine, or a peptide or polypeptide useful in gene therapy.

A plurality of the vectors of the invention may be physically linked or each vector may be present on an individual plasmid or other, e.g., linear, nucleic acid delivery vehicle.

The invention also provides a method to prepare influenza virus. The method comprises contacting a cell, e.g., an avian or a mammalian cell, with the isolated virus of the invention or a plurality of the vectors of the invention, e.g., sequentially or simultaneously, for example, employing a composition comprising a plurality of the vectors, in an amount effective to yield infectious influenza virus. The invention also includes isolating virus from a cell infected with the virus or contacted with the vectors and/or composition. The invention further provides a host cell infected with the virus of the invention or contacted with the composition or vectors of the invention. In one embodiment, a host cell is infected with an attenuated (e.g., cold adapted) donor virus and a virus of the invention to prepare a cold-adapted reassortant virus useful as a cold-adapted live virus vaccine.

The invention also provides a method to induce an immune response in a mammal, e.g., to immunize a mammal, against one more pathogens, e.g., against a virus of the invention and optionally a bacteria, a different virus, or a parasite or other antigen. An immunological response to a composition or vaccine is the development in the host organism of a cellular and/or antibody-mediated immune response to a viral polypeptide, e.g., an administered viral preparation, polypeptide or one encoded by an administered nucleic acid molecule, which can prevent or inhibit infection to that virus or a closely (structurally) related virus. Usually, such a response consists of the subject producing antibodies, B cell, helper T cells, suppressor T cells, and/or cytotoxic T cells directed specifically to an antigen or antigens included in the composition or vaccine of interest. The method includes administering to the host organism, e.g., a mammal, an effective amount of the influenza virus of the invention, e.g., an attenuated, live virus, optionally in combination with an adjuvant and/or a carrier, e.g., in an amount effective to prevent or ameliorate infection of an animal such as a mammal by that virus or an antigenically closely related virus. In one embodiment, the virus is administered intramuscularly while in another embodiment, the virus is administered intranasally. In some dosing protocols, all doses may be administered intramuscularly or intranasally, while in others a combination of intramuscular and intranasal administration is employed. The vaccine may further contain other isolates of influenza virus including recombinant influenza virus, other pathogen(s), additional biological agents or microbial components, e.g., to form a multivalent vaccine. In one embodiment, intranasal vaccination with inactivated equine influenza virus and a mucosal adjuvant, e.g., the non-toxic B chain of cholera toxin, may induce virus-specific IgA and neutralizing antibody in the nasopharynx as well as serum IgG.

The equine influenza vaccine may employed with other anti-virals, e.g., amantadine, rimantadine, and/or neuraminidase inhibitors, e.g., may be administered separately in conjunction with those anti-virals, for instance, administered before, during and/or after.

Further provided is a diagnostic method which employs a virus of the invention, an isolated viral protein encoded thereby, or antisera specific for the virus or protein, to detect viral specific antibodies or viral specific proteins.

BRIEF DESCRIPTION OF THE FIGURES

FIGS. 1A-1R. Sequences of A/Equine/Wisconsin/1/03. SEQ ID NOs:1-8, 17 and 18 represent the deduced amino acid sequence for HA, NA, PB1, PB2, PA, NP, M1, NS1, M2, and NS2, respectively, of A/Equine/Wisconsin/1/03. SEQ ID NOs:9-16 represent the mRNA sense nucleotide

sequence for HA, NA, PB1, PB2, PA, NP, M (M1 and M2) and NS (NS1 and NS2), respectively, of A/Equine/Wisconsin/1/03.

FIG. 2. Sequence alignment of HA-1 of A/Equine/NewYork/99 (SEQ ID NO:19) and A/Equine/Wisconsin/1/03 (SEQ ID NO:20).

DETAILED DESCRIPTION OF THE INVENTION

Definitions

As used herein, the term "isolated" refers to in vitro preparation and/or isolation of a nucleic acid molecule, e.g., vector or plasmid, peptide or polypeptide (protein), or virus of the invention, so that it is not associated with in vivo substances, or is substantially purified from in vitro substances. An isolated virus preparation is generally obtained by in vitro culture and propagation, and is substantially free from other infectious agents.

As used herein, "substantially purified" means the object species is the predominant species, e.g., on a molar basis it is more abundant than any other individual species in a composition, and preferably is at least about 80% of the species present, and optionally 90% or greater, e.g., 95%, 98%, 99% or more, of the species present in the composition.

As used herein, "substantially free" means below the level of detection for a particular infectious agent using standard detection methods for that agent.

A "recombinant" virus is one which has been manipulated in vitro, e.g., using recombinant DNA techniques, to introduce changes to the viral genome.

As used herein, the term "recombinant nucleic acid" or "recombinant DNA sequence or segment" refers to a nucleic acid, e.g., to DNA, that has been derived or isolated from a source, that may be subsequently chemically altered in vitro, so that its sequence is not naturally occurring, or corresponds to naturally occurring sequences that are not positioned as they would be positioned in the native genome. An example of DNA "derived" from a source, would be a DNA sequence that is identified as a useful fragment, and which is then chemically synthesized in essentially pure form. An example of such DNA "isolated" from a source would be a useful DNA sequence that is excised or removed from said source by chemical means, e.g., by the use of restriction endonucleases, so that it can be further manipulated, e.g., amplified, for use in the invention, by the methodology of genetic engineering.

Influenza Virus Type A Structure and Propagation

Influenza A viruses possess a genome of eight single-stranded negative-sense viral RNAs (vRNAs) that encode at least ten proteins. The influenza virus life cycle begins with binding of the hemagglutinin (HA) to sialic acid-containing receptors on the surface of the host cell, followed by receptor-mediated endocytosis. The low pH in late endosomes triggers a conformational shift in the HA, thereby exposing the N-terminus of the HA2 subunit (the so-called fusion peptide). The fusion peptide initiates the fusion of the viral and endosomal membrane, and the matrix protein (M1) and RNP complexes are released into the cytoplasm. RNPs consist of the nucleoprotein (NP), which encapsidates vRNA, and the viral polymerase complex, which is formed by the PA, PB1, and PB2 proteins. RNPs are transported into the nucleus, where transcription and replication take place. The RNA polymerase complex catalyzes three different reactions: synthesis of an mRNA with a 5' cap and 3' polyA

structure, of a full-length complementary RNA (cRNA), and of genomic vRNA using the cRNA as a template. Newly synthesized vRNAs, NP, and polymerase proteins are then assembled into RNPs, exported from the nucleus, and transported to the plasma membrane, where budding of progeny virus particles occurs. The neuraminidase (NA) protein plays a crucial role late in infection by removing sialic acid from sialyloligosaccharides, thus releasing newly assembled virions from the cell surface and preventing the self aggregation of virus particles. Although virus assembly involves protein-protein and protein-vRNA interactions, the nature of these interactions is largely unknown.

Any cell, e.g., any avian or mammalian cell, such as a human, canine, bovine, equine, feline, swine, ovine, mink, e.g., MvLu1 cells, or non-human primate cell, including mutant cells, which supports efficient replication of influenza virus can be employed to isolate and/or propagate influenza viruses. Isolated viruses can be used to prepare a reassortant virus, e.g., an attenuated virus. In one embodiment, host cells for vaccine production are those found in avian eggs. In another embodiment, host cells for vaccine production are continuous mammalian or avian cell lines or cell strains. It is preferred to establish a complete characterization of the cells to be used, so that appropriate tests for purity of the final product can be included. Data that can be used for the characterization of a cell includes (a) information on its origin, derivation, and passage history; (b) information on its growth and morphological characteristics; (c) results of tests of adventitious agents; (d) distinguishing features, such as biochemical, immunological, and cytogenetic patterns which allow the cells to be clearly recognized among other cell lines; and (e) results of tests for tumorigenicity. Preferably, the passage level, or population doubling, of the host cell used is as low as possible.

It is preferred that the virus produced by the host cell is highly purified prior to vaccine or gene therapy formulation. Generally, the purification procedures result in the extensive removal of cellular DNA, other cellular components, and adventitious agents. Procedures that extensively degrade or denature DNA can also be used.

Equine Influenza Virus Detection

Disease causing equine influenza viruses are generally Type A influenza viruses of the H7N7 (equi-1) and H3N8 (equi-2) subtypes. These generally differ from the subtypes that cause infection in man (H1N1, H2N2 and H3N2). Equine influenza is contracted by either inhalation or contact with secretions (e.g., physiological fluid) containing live virus. The virus infects the epithelial cells of the upper and lower airways and can cause deciliation of large areas of the respiratory tract within 4 to 6 days. As a result, the mucociliary clearance mechanism is compromised and tracheal clearance rates may be reduced for up to 32 days following infection. Bronchitis and bronchiolitis develop followed by interstitial pneumonia accompanied by congestion, edema and leukocyte infiltration. In general, H3N8 viruses cause more severe disease than H7N7 viruses, viruses of the H3N8 subtype are more pneumotropic and have also been associated with myocarditis.

Clinical signs in previously influenza-naïve animals are easily recognizable. Influenza is characterized by its sudden onset with an incubation period of 1 to 3 days. The first sign is an elevation of body temperature (up to 41° C.), which is usually biphasic. This is followed by a deep dry cough that releases large quantities of virus into the atmosphere often accompanied by a serous nasal discharge, which may become mucopurulent due to secondary bacterial infection. The other most commonly observed clinical signs are myal-

gia, inappetance, and enlarged submandibular lymph nodes. Edema of the legs and scrotum is observed very rarely. The severity of the disease varies with the dose and strain of virus and the immune status of the horse.

Previously healthy, immunocompetent adult horses usually recover from uncomplicated influenza within 10 days, although coughing may persist for longer. If secondary bacterial infection occurs, it can prolong the recovery period. However, relatively high mortality rates have been recorded in foals, animals in poor condition and donkeys. If maternal antibody is absent at the time of exposure, young foals may develop a viral pneumonia leading to death. Deaths among adult animals are usually a consequence of secondary bacterial infection leading to pleuritis, suppurative pneumonia or rarely, purpura haemorrhagica. Sequelae of equine influenza can include chronic pharyngitis, chronic bronchiolitis, myocarditis, and alveolar emphysema, which can contribute to heaves, and secondary sinus and guttural pouch infections.

Clinical signs in animals partially immune as a result of vaccination or previous infection are more difficult to recognize as there may be little or no coughing or pyrexia. Whereas spread of infection throughout a group of naïve animals is always rapid, there have been outbreaks in which the infection circulated subclinically in vaccinated horses for 18 days before inducing recognizable clinical signs.

Outbreaks of infectious respiratory disease may be caused by various agents, including equine herpes viruses, rhinoviruses, adenoviruses, and arteritis viruses, *Streptococcus equi*, or *S. zooepidemicus*. A presumptive diagnosis of influenza based on clinical signs should be confirmed by virus isolation or detection, or by serological testing. Laboratory confirmation of a clinical diagnosis may be by traditional isolation of virus from nasopharyngeal swabs or serology to demonstrate seroconversion, or by rapid diagnostic tests which detect the presence of viral antigens, viral nucleic acid, or virally infected cells in respiratory secretions. Rapid diagnostic tests, despite their convenience and ease of use, provide little or no information about genetic or antigenic characteristics of the infecting strain of virus and do not allow isolation of the virus.

Nasopharyngeal swabs for virus isolation or detection should be taken as promptly as possible. Results of experimental challenge studies suggest that peak viral titers are obtained during the initial 24 to 48 hours of fever, on the second or third day after infection, and the duration of viral shedding is usually not more than 4 or 5 days. Nasal swab samples are taken by passing a swab as far as possible into the horse's nasopharynx via the ventral meatus to absorb respiratory secretions. Swabs should be transferred immediately to a container with virus transport medium and transported on ice to maintain viability of the virus. Virus is unlikely to survive if dry swabs are taken and there is an increased chance of contamination if bacterial transport medium is used. Nasal swab samples may be inoculated into the allantoic (or amniotic) cavity of 9- to 11-day-old embryonated hens' eggs. After incubation at 33-35° C. for 3 days, the allantoic fluid is harvested and tested for haemagglutinating activity. Alternatively, cell culture may be used to isolate viruses. Influenza infection can also be diagnosed by comparison of the results of serological testing of an acute serum sample taken as soon as possible after the onset of clinical signs and a convalescent serum sample taken 2 to 4 weeks later.

The haemagglutination inhibition (HI) test measures the capacity of influenza-specific antibody present in serum samples to inhibit the agglutination of red blood cells by

virus. Sera are heat-inactivated and pre-treated to reduce non-specific reactions and serially diluted prior to incubation with a standard dose of virus in a U-bottomed microtiter plate. A suspension of red blood cells is added and, after a further incubation period, examined for agglutination. A four-fold rise in virus-specific antibodies indicates infection. Whole virus antigen may be used for H7N7 viruses, but Tween 80-ether disrupted antigen is usually required to enhance the sensitivity of the assay for H3N8 viruses. In repeatedly vaccinated horses, infection may fail to stimulate a 4-fold increase in HI titer.

The single-radial haemolysis (SRH) test, although less strain-specific, is more reproducible and less error prone than the HI test and, as it is a linear test, is more sensitive, enabling detection of smaller increases in antibody induced by infection in heavily vaccinated horses. The SRH test is based on the ability of influenza-specific antibodies to lyse virus-coated red blood cells in the presence of complement. Test sera are added to wells punched in agarose containing coated red blood cells and complement and allowed to diffuse through the agarose for 20 hours. The areas of clear zones of haemolysis around the wells are proportional to the level of influenza antibody present in the serum samples.

If horses are vaccinated in the face of infection, it may not be possible, using the HI and SRH assays, to determine whether any increase in antibody levels is due to vaccination or infection.

Influenza Vaccines

A vaccine of the invention includes an isolated influenza virus of the invention, and optionally one or more other isolated viruses including other isolated influenza viruses, West Nile virus, equine herpes virus, equine arteritis virus, equine infectious anemia lentivirus, rabies virus, Eastern and/or Western and/or Venezuelan equine encephalitis virus, one or more immunogenic proteins or glycoproteins of one or more isolated influenza viruses or one or more other pathogens, e.g., an immunogenic protein from one or more bacteria, non-influenza viruses, yeast or fungi, or isolated nucleic acid encoding one or more viral proteins (e.g., DNA vaccines) including one or more immunogenic proteins of the isolated influenza virus of the invention. In one embodiment, the influenza viruses of the invention may be vaccine vectors for influenza virus or other pathogens.

A complete virion vaccine may be concentrated by ultrafiltration and then purified by zonal centrifugation or by chromatography. It is inactivated before or after purification using formalin or beta-propiolactone, for instance.

A subunit vaccine comprises purified glycoproteins. Such a vaccine may be prepared as follows: using viral suspensions fragmented by treatment with detergent, the surface antigens are purified, by ultracentrifugation for example. The subunit vaccines thus contain mainly HA protein, and also NA. The detergent used may be cationic detergent for example, such as hexadecyl trimethyl ammonium bromide (Bachmeyer, 1975), an anionic detergent such as ammonium deoxycholate (Laver & Webster, 1976); or a nonionic detergent such as that commercialized under the name TRITON X100. The hemagglutinin may also be isolated after treatment of the virions with a protease such as bromelain, then purified by a method such as that described by Grand and Skehel (1972).

A split vaccine comprises virions which have been subjected to treatment with agents that dissolve lipids. A split vaccine can be prepared as follows: an aqueous suspension of the purified virus obtained as above, inactivated or not, is treated, under stirring, by lipid solvents such as ethyl ether or chloroform, associated with detergents. The dissolution of

the viral envelope lipids results in fragmentation of the viral particles. The aqueous phase is recuperated containing the split vaccine, constituted mainly of hemagglutinin and neuraminidase with their original lipid environment removed, and the core or its degradation products. Then the residual infectious particles are inactivated if this has not already been done.

Inactivated Vaccines.

Inactivated influenza virus vaccines are provided by inactivating replicated virus using known methods, such as, but not limited to, formalin or β -propiolactone treatment. Inactivated vaccine types that can be used in the invention can include whole-virus (WV) vaccines or subvirion (SV) (split) vaccines. The WV vaccine contains intact, inactivated virus, while the SV vaccine contains purified virus disrupted with detergents that solubilize the lipid-containing viral envelope, followed by chemical inactivation of residual virus.

In addition, vaccines that can be used include those containing the isolated HA and NA surface proteins, which are referred to as surface antigen or subunit vaccines.

Live Attenuated Virus Vaccines.

Live, attenuated influenza virus vaccines can be used for preventing or treating influenza virus infection. Attenuation may be achieved in a single step by transfer of attenuated genes from an attenuated donor virus to a replicated isolate or reassorted virus according to known methods (see, e.g., Murphy, 1993). Since resistance to influenza A virus is mediated primarily by the development of an immune response to the HA and/or NA glycoproteins, the genes coding for these surface antigens must come from the reassorted viruses or clinical isolates. The attenuated genes are derived from the attenuated parent. In this approach, genes that confer attenuation preferably do not code for the HA and NA glycoproteins.

Viruses (donor influenza viruses) are available that are capable of reproducibly attenuating influenza viruses, e.g., a cold adapted (ca) donor virus can be used for attenuated vaccine production. Live, attenuated reassortant virus vaccines can be generated by mating the ca donor virus with a virulent replicated virus. Reassortant progeny are then selected at 25° C., (restrictive for replication of virulent virus), in the presence of an appropriate antiserum, which inhibits replication of the viruses bearing the surface antigens of the attenuated ca donor virus. Useful reassortants are: (a) infectious, (b) attenuated for seronegative non-adult mammals and immunologically primed adult mammals, (c) immunogenic and (d) genetically stable. The immunogenicity of the ca reassortants parallels their level of replication. Thus, the acquisition of the six transferable genes of the ca donor virus by new wild-type viruses has reproducibly attenuated these viruses for use in vaccinating susceptible mammals both adults and non-adult.

Other attenuating mutations can be introduced into influenza virus genes by site-directed mutagenesis to rescue infectious viruses bearing these mutant genes. Attenuating mutations can be introduced into non-coding regions of the genome, as well as into coding regions. Such attenuating mutations can also be introduced into genes other than the HA or NA, e.g., the PB2 polymerase gene (Subbarao et al., 1993). Thus, new donor viruses can also be generated bearing attenuating mutations introduced by site-directed mutagenesis, and such new donor viruses can be used in the production of live attenuated reassortants vaccine candidates in a manner analogous to that described above for the ca donor virus. Similarly, other known and suitable attenuated donor strains can be reassorted with influenza virus to obtain

attenuated vaccines suitable for use in the vaccination of mammals (Enami et al., 1990; Muster et al., 1991; Subbarao et al., 1993).

It is preferred that such attenuated viruses maintain the genes from the virus that encode antigenic determinants substantially similar to those of the original clinical isolates. This is because the purpose of the attenuated vaccine is to provide substantially the same antigenicity as the original clinical isolate of the virus, while at the same time lacking pathogenicity to the degree that the vaccine causes minimal chance of inducing a serious disease condition in the vaccinated mammal.

The virus can thus be attenuated or inactivated, formulated and administered, according to known methods, as a vaccine to induce an immune response in an animal, e.g., a mammal. Methods are well-known in the art for determining whether such attenuated or inactivated vaccines have maintained similar antigenicity to that of the clinical isolate or high growth strain derived therefrom. Such known methods include the use of antisera or antibodies to eliminate viruses expressing antigenic determinants of the donor virus; chemical selection (e.g., amantadine or rimantidine); HA and NA activity and inhibition; and nucleic acid screening (such as probe hybridization or PCR) to confirm that donor genes encoding the antigenic determinants (e.g., HA or NA genes) are not present in the attenuated viruses. See, e.g., Robertson et al., 1988; Kilbourne, 1969; Aymard-Henry et al., 1985; Robertson et al., 1992.

Pharmaceutical Compositions

Pharmaceutical compositions of the present invention, suitable for inoculation, e.g., nasal, parenteral or oral administration, comprise one or more influenza virus isolates, e.g., one or more attenuated or inactivated influenza viruses, a subunit thereof, isolated protein(s) thereof, and/or isolated nucleic acid encoding one or more proteins thereof, optionally further comprising sterile aqueous or non-aqueous solutions, suspensions, and emulsions. The compositions can further comprise auxiliary agents or excipients, as known in the art. See, e.g., Berkow et al., 1987; *Avery's Drug Treatment*, 1987; Osol, 1980. The composition of the invention is generally presented in the form of individual doses (unit doses).

Conventional vaccines generally contain about 0.1 to 200 µg, e.g., 30 to 100 µg, of HA from each of the strains entering into their composition. The vaccine forming the main constituent of the vaccine composition of the invention may comprise a single influenza virus, or a combination of influenza viruses, for example, at least two or three influenza viruses, including one or more reassortant(s).

Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and/or emulsions, which may contain auxiliary agents or excipients known in the art. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Carriers or occlusive dressings can be used to increase skin permeability and enhance antigen absorption. Liquid dosage forms for oral administration may generally comprise a liposome solution containing the liquid dosage form. Suitable forms for suspending liposomes include emulsions, suspensions, solutions, syrups, and elixirs containing inert diluents commonly used in the art, such as purified water. Besides the inert diluents, such compositions can also include adjuvants, wetting agents, emulsifying and suspending agents, or sweetening, flavoring, or perfuming agents. See, e.g., Berkow et al., 1992; *Avery's*, 1987; and Osol, 1980.

When a composition of the present invention is used for administration to an individual, it can further comprise salts, buffers, adjuvants, or other substances which are desirable for improving the efficacy of the composition. For vaccines, adjuvants, substances which can augment a specific immune response, can be used. Normally, the adjuvant and the composition are mixed prior to presentation to the immune system, or presented separately, but into the same site of the organism being immunized. Examples of materials suitable for use in vaccine compositions are provided in Osol (1980).

Heterogeneity in a vaccine may be provided by mixing replicated influenza viruses for at least two influenza virus strains, such as 2-20 strains or any range or value therein. Influenza A virus strains having a modern antigenic composition are preferred. Vaccines can be provided for variations in a single strain of an influenza virus, using techniques known in the art.

A pharmaceutical composition according to the present invention may further or additionally comprise at least one chemotherapeutic compound, for example, for gene therapy, immunosuppressants, anti-inflammatory agents or immune enhancers, and for vaccines, chemotherapeutics including, but not limited to, gamma globulin, amantadine, guanidine, hydroxybenzimidazole, interferon- α , interferon- β , interferon- γ , tumor necrosis factor-alpha, thiosemicarbazones, methisazone, rifampin, ribavirin, a pyrimidine analog, a purine analog, foscarnet, phosphonoacetic acid, acyclovir, dideoxynucleosides, a protease inhibitor, or ganciclovir.

The composition can also contain variable but small quantities of endotoxin-free formaldehyde, and preservatives, which have been found safe and not contributing to undesirable effects in the organism to which the composition is administered.

Pharmaceutical Purposes

The administration of the composition (or the antisera that it elicits) may be for either a "prophylactic" or "therapeutic" purpose. When provided prophylactically, the compositions of the invention which are vaccines are provided before any symptom or clinical sign of a pathogen infection becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate any subsequent infection. When provided prophylactically, the gene therapy compositions of the invention, are provided before any symptom or clinical sign of a disease becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate one or more symptoms or clinical signs associated with the disease.

When provided therapeutically, an attenuated or inactivated viral vaccine is provided upon the detection of a symptom or clinical sign of actual infection. The therapeutic administration of the compound(s) serves to attenuate any actual infection. See, e.g., Berkow et al., 1992; and Avery, 1987. When provided therapeutically, a gene therapy composition is provided upon the detection of a symptom or clinical sign of the disease. The therapeutic administration of the compound(s) serves to attenuate a symptom or clinical sign of that disease.

Thus, an attenuated or inactivated vaccine composition of the present invention may be provided either before the onset of infection (so as to prevent or attenuate an anticipated infection) or after the initiation of an actual infection. Similarly, for gene therapy, the composition may be provided before any symptom or clinical sign of a disorder or disease is manifested or after one or more symptoms are detected.

A composition is said to be "pharmacologically acceptable" if its administration can be tolerated by a recipient

mammal. Such an agent is said to be administered in a “therapeutically effective amount” if the amount administered is physiologically significant. A composition of the present invention is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient, e.g., enhances at least one primary or secondary humoral or cellular immune response against at least one strain of an infectious influenza virus.

The “protection” provided need not be absolute, i.e., the influenza infection need not be totally prevented or eradicated, if there is a statistically significant improvement compared with a control population or set of mammals. Protection may be limited to mitigating the severity or rapidity of onset of symptoms or clinical signs of the influenza virus infection.

Pharmaceutical Administration

A composition of the present invention may confer resistance to one or more pathogens, e.g., one or more influenza virus strains, by either passive immunization or active immunization. In active immunization, an inactivated or attenuated live vaccine composition is administered prophylactically to a host (e.g., a mammal), and the host’s immune response to the administration protects against infection and/or disease. For passive immunization, the elicited antisera can be recovered and administered to a recipient suspected of having an infection caused by at least one influenza virus strain. A gene therapy composition of the present invention may yield prophylactic or therapeutic levels of the desired gene product by active immunization.

In one embodiment, the vaccine is provided to a mammalian female (at or prior to pregnancy or parturition), under conditions of time and amount sufficient to cause the production of an immune response which serves to protect both the female and the fetus or newborn (via passive incorporation of the antibodies across the placenta or in the mother’s milk).

The present invention thus includes methods for preventing or attenuating a disorder or disease, e.g., an infection by at least one strain of pathogen. As used herein, a vaccine is said to prevent or attenuate a disease if its administration results either in the total or partial attenuation (i.e., suppression) of a clinical sign or condition of the disease, or in the total or partial immunity of the individual to the disease. As used herein, a gene therapy composition is said to prevent or attenuate a disease if its administration results either in the total or partial attenuation (i.e., suppression) of a clinical sign or condition of the disease, or in the total or partial immunity of the individual to the disease.

At least one influenza virus isolate of the present invention, including one which is inactivated or attenuated, one or more isolated viral proteins thereof, one or more isolated nucleic acid molecules encoding one or more viral proteins thereof, or a combination thereof, may be administered by any means that achieve the intended purposes.

For example, administration of such a composition may be by various parenteral routes such as subcutaneous, intravenous, intradermal, intramuscular, intraperitoneal, intranasal, oral or transdermal routes. Parenteral administration can be accomplished by bolus injection or by gradual perfusion over time.

A typical regimen for preventing, suppressing, or treating an influenza virus related pathology, comprises administration of an effective amount of a vaccine composition as described herein, administered as a single treatment, or repeated as enhancing or booster dosages, over a period up to and including between one week and about 24 months, or any range or value therein.

According to the present invention, an “effective amount” of a composition is one that is sufficient to achieve a desired effect. It is understood that the effective dosage may be dependent upon the species, age, sex, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect wanted. The ranges of effective doses provided below are not intended to limit the invention and represent dose ranges.

The dosage of a live, attenuated or killed virus vaccine for an animal such as a mammalian adult organism can be from about 10^2 - 10^{15} , e.g., 10^3 - 10^{12} , plaque forming units (PFU)/kg, or any range or value therein. The dose of inactivated vaccine can range from about 0.1 to 1000, e.g., 30 to 100 μ g, of HA protein. However, the dosage should be a safe and effective amount as determined by conventional methods, using existing vaccines as a starting point.

The dosage of immunoreactive HA in each dose of replicated virus vaccine can be standardized to contain a suitable amount, e.g., 30 to 100 μ g or any range or value therein, or the amount recommended by government agencies or recognized professional organizations. The quantity of NA can also be standardized, however, this glycoprotein may be labile during purification and storage.

Compositions and Dosing for Equine Influenza Vaccines

Equine influenza vaccines generally include representative strains of H7N7 and H3N8 subtypes either as inactivated whole virus or their subunits. They provide protection against influenza by inducing antibody to the surface glycoproteins, in particular to HA, which is essential for viral attachment and entry into cells, and/or potentially important cell-mediated immune responses to other viral proteins. Vaccination is helpful in preventing influenza but the protection is short-lived (3-4 months using conventional inactivated virus vaccines), so the frequency of vaccination varies according to how often the horse will likely come in contact with the virus (see Table 1). The usual procedure for the primary course is vaccination with a single dose followed 3 to 6 weeks later with a second dose. Vaccine manufacturers recommend that booster vaccinations be given at 6- to 12-month intervals thereafter. Alternatively, a horse is administered one 1 to 2 ml dose, e.g., via intramuscular (IM) injection, a second 1 to 2 ml dose 3 to 4 weeks later at a different injection site, e.g., via IM injection, and optionally a third 1 to 2 ml dose, e.g., IM or intranasal (IN) administration. Each 1 to 2 ml dose of vaccine may contain approximately 1-500 billion virus particles, and preferably 100 billion particles. Horses in contact with a large number of horses, for example, at a boarding stable, training centers, racetracks, shows, and other such events, are often vaccinated every 2-3 months. A three-dose primary series has been shown to induce a higher and more persistent immunity than the recommended two-dose series regardless of the age.

Using conventional vaccines, it is advisable to vaccinate young horses, particularly racehorses and other competition horses, at 4 to 6 month intervals for several years after their primary course of vaccinations. It has been demonstrated that inclusion of an additional booster vaccination between the second and third vaccination recommended by the vaccine manufacturers is of benefit to young horses. An annual booster will usually suffice for older horses such as show jumpers and brood mares that have been vaccinated regularly since they were foals. Vaccination in the face of an ongoing outbreak is sometimes practiced, but is not likely to be effective without an interval of at least 7 to 10 days before the freshly vaccinated horses are exposed to infection. Equine influenza outbreaks are not seasonal as in man but are frequently associated with sales or race meets where

horses from different regions congregate and mix. It may therefore be advantageous to time additional booster vaccinations to be given prior to such events.

Brood mares should be vaccinated in the later stages of pregnancy, but not later than 2 weeks prior to foaling, to ensure a good supply of colostral antibodies for the foal. Foal vaccinations should begin at 3-6 months of age, with a booster at 4-7 months, again at 5-8 months, and repeated every three months if the foal is at high risk of exposure.

TABLE 1

	Foals & Weanlings from Vaccinated Mares	Foal & Weanlings from non-Vaccinated Mares	Yearlings	Performance Horse	Pleasure Horses	Brood-mares
Influenza inactivated injectable	1st Dose: 9 months 2nd Dose: 10 months 3rd Dose: 11-12 months Then at 3 month intervals	1st Dose: 6 months 2nd Dose: 7 months 3rd Dose: 8 months Then at 3 month intervals	Every 3-4 months	Every 3-5 months	Annual with Boosters prior to likely exposure	At least semi-annual, with 1 Booster 4-6 weeks prepartum
Influenza intranasal adapted live virus	1st Dose: 12 months; has been safely administered to foals less than 11 months	1st Dose: 12 months; has been safely administered to foals less than 11 months	Every 4-6 months	Every 4-6 months	Every 4-6 months	Annual before breeding

Influenza vaccines may be combined with tetanus or herpesvirus antigens as well as other pathogens, e.g., equine pathogens. The immune response elicited by tetanus toxoid is much more durable than that induced by influenza antigen. In an intensive influenza vaccination program, vaccines containing influenza only are thus preferred.

Levels of antibody (measured by the SRH assay) required for protection of horses have been identified through vaccination and challenge studies and from field data. Because the vaccine-induced antibody response to HA in horses is remarkably short-lived, adjuvants such as aluminum hydroxide or carbomer are normally included to enhance the amplitude and duration of the immune response to whole virus vaccines. Subunit equine influenza vaccines containing immune stimulating complexes (ISCOMs) are also immunogenic.

Historically, antigenic content in inactivated vaccines has been expressed in terms of chick cell agglutinating (CCA) units of HA and potency in terms of HI antibody responses induced in guinea pigs and horses, neither of which yields reproducible results. The single radial diffusion (SRD) assay is an improved in vitro potency test that measures the concentration of immunologically active HA (expressed in terms of micrograms of HA) and can be used for in-process testing before the addition of adjuvant.

The invention will be further described by the following non-limiting example.

Example

An approximately 36-hour-old Morgan/Friesian colt was referred to the large animal hospital at the University of Wisconsin for an evaluation of altered mentation (mental status), first noticed shortly after birth. Parturition had been unobserved, but the foal had been found separated from the

mare by a fence at a few hours of age. The foal was ambulatory and able to nurse when first discovered but showed progressive disorientation, apparent blindness, and aimless wandering during the following 36-hour period. A SNAP immunoglobulin G (IgG) assay (Idexx Laboratories, Westbrook, Me.) at 24 hours of age had shown an IgG concentration >800 mg/dL, and a CBC performed at that time was normal. The foal was treated twice with dimethyl sulfoxide 1 g/kg IV, diluted in 5% dextrose before referral.

At presentation, the colt wandered aimlessly, bumped into objects, and appeared blind with sluggish but intact pupillary light responses. When positioned under the mare, the foal nursed successfully. Physical examination was unremarkable. A CBC and serum biochemistry were normal, including a serum IgG concentration of 937 mg/dL measured by radioimmunodiffusion.

Initial treatment for presumptive hypoxemic, ischemic encephalopathy included a 250 mL loading dose of 20% magnesium sulfate for 1 hour, followed by a constant rate infusion at 42 mL/h and thiamine hydrochloride 2.2 mg/kg IV q24 h. Antimicrobial therapy consisted of amikacin 20 mg/kg IV q24 h and procaine penicillin G 22,000 U/kg IM q12 h. Omeprazole 1 mg/kg PO q24 h also was administered to the foal to help prevent the development of gastric ulcers.

The foal's mental status remained static during the next 24 hours, and additional treatment with mannitol 1 g/kg IV q24 h and dexamethasone sodium phosphate 0.1 mg/kg IV q24 h on days 2 and 3 of hospitalization was not associated with improvement. On day 3, the foal underwent general anesthesia for a computerized tomographic scan of the skull and proximal spine, which was normal. A cerebrospinal fluid sample was obtained from the lumbosacral space and was normal on cytologic evaluation and had a normal protein concentration.

On day 4 of hospitalization, the foal developed a right-sided head tilt but otherwise remained static through day 5 of hospitalization. Magnesium sulfate therapy was discontinued on day 5, but the remainder of the therapeutic regimen was unchanged. On day 6, the foal had 2 brief, generalized seizures that were controlled with midazolam 0.05 mg/kg IV. Between seizures, the foal was still bright, afebrile, and nursing.

On day 7 of hospitalization, the foal became febrile (40°C.) and developed a mucopurulent nasal discharge and

progressive tachypnea with diffuse adventitious crackles and wheezes on auscultation. Fever, mucopurulent nasal discharge, and coughing had been noted in several other mares and foals in the neonatal care unit during the previous 7 days. Antimicrobial therapy was changed to ticarcillin/clavulanic acid 50 mg/kg IV q8 h had gentamicin 6.6 mg/kg IV q24 h, and the foal was treated with polyionic fluids, although it was still nursing. During days 8-10, the foal's neurologic status continued to improve, with a resolution of the head tilt and a return to normal mentation, but the tachypnea, dyspnea, and adventitious lung sounds worsened. Thoracic radiography at this time showed a severe, diffuse bronchointerstitial pattern. Aminophylline 0.5 mg/kg IV q12 h by slow infusion and nasal insufflation of oxygen were instituted on days 9 and 10 of hospitalization. Serial arterial blood gas analysis identified severe hypoxemia (PaO₂, 52 mm Hg), hypercapnia (PaCO₂, 68.4 mm Hg), and reduced oxygen saturation (76%) by the end of day 10. Consequently, the foal was placed on a mechanical ventilator. Ventilatory support and total parenteral nutrition were continued for 48 hours, during which time arterial blood gas values normalized on 100% oxygen. Antimicrobial therapy was continued as before. When challenged on day 13 by the removal of ventilatory support, the foal developed severe dyspnea and cyanosis and was euthanized at the owner's request. An aerobic culture of a transtracheal aspirate obtained on day 13 grew *Klebsiella pneumoniae* and *Escherichia coli* resistant to ticarcillin/clavulanic acid and gentamicin.

A complete gross and histopathologic postmortem examination was performed, as well as a real-time quantitative polymerase chain reaction (PCR) evaluation for the presence of equine herpes virus (EHV)-1 and EHV-4 in samples of nasal secretions; serologic tests to determine if there was exposure to equine viral arteritis virus; and a Directigen Flu A assay (Bectin Dickinson and Co., Franklin, N.J.) and virus isolation from samples of nasal secretions to test for the presence of influenza virus. Samples of nasal secretions were collected with Dacron swabs that were subsequently placed in 2 mL of viral transport media containing phosphate-buffered saline, 0.5% bovine serum albumin, and penicillin G, streptomycin, nystatin, and gentamicin. The nasal swab samples were collected on day 8 of hospitalization. Follow-up evaluations for the influenza virus included immunohistochemistry on snap-frozen and formalin-fixed lung, abdominal viscera, and central nervous system tissues for the presence of influenza nucleoprotein (NP) expression, virus isolation from frozen lung tissue, and viral sequence analyses. Gross post-mortem examination identified severe diffuse interstitial pneumonia and subdural hemorrhage on the caudal ventral surface of the brain around the pituitary gland but no evidence of sepsis or pathology in other organs. Histopathologic examination of the lung identified necrotizing bronchitis and bronchiolitis, diffuse squamous metaplasia, and multifocal interstitial pneumonia. A mild mononuclear infiltrate lined the lower airways and, occasionally, areas of alveolar collapse associated with congestion and exudate. Evaluation of the brain tissue revealed a mild dilatation of the ventricular system with diffuse white matter vacuolation, particularly in the cerebellum. Cresyl violet staining for the presence of myelin was performed on multiple sections and showed diminished but present myelin throughout the brain and spinal cord when compared to tissues from an age-matched control stained in parallel. Additional histopathologic abnormalities in the central nervous system included an apparent absence of the molecular

layer within the cerebellum. Serologic tests for equine viral arteritis and a real-time PCR assay for EHV-1 and EHV-4 DNA were negative.

The presence of influenza virus in nasal secretions initially was confirmed by a positive Directigen assay. Previous studies have documented the sensitivity and specificity of this assay when applied to equine nasal secretion samples (Morely et al., 1995 and Chambers et al., 1994). Samples of the nasal swab transport media also were inoculated into the allantoic cavity of embryonated chicken eggs and onto Madin-Darby canine kidney (MDCK) cells growing in 24-well cell culture plates. Cytopathologic effects consistent with influenza virus growth were observed in the inoculated MDCK cells, and an agent that caused the hemagglutination of chicken red blood cells was isolated from the inoculated eggs (Palmar et al., 1975). The presence of influenza virus in the MDCK cell cultures was confirmed by the immunocytochemical staining (Landolt et al., 2003) of the inoculated cells with an anti-NP monoclonal antibody (Mab 68D2 (kindly provided by Dr. Yoshihiro Kawaoka, University of Wisconsin-Madison School of Veterinary Medicine) with positive (swine influenza virus inoculated) and negative (mock inoculated) control cells included on the same plate. The identity of the virus as an H3-subtype equine influenza virus was confirmed by reverse transcription-PCR amplification of the hemagglutinin (HA) gene from the isolate, with primers described in Olsen et al. (1997), followed by cycle sequencing of the full-length protein coding region of the HA gene and pairwise comparisons to viral sequences available in GenBank (DNASTAR software, version 4.0 for Win32, Bestfit, Madison, Wis.). The virus was shown to be derived from the North American lineage of H3 equine influenza viruses by a phylogenetic analysis that used a maximum parsimony bootstrap analysis (PAUP software, version 4.0b6; David Swofford, Smithsonian Institution, Washington, D.C.) of the HA sequence compared to reference virus strains with a fast-heuristic search of 1,000 bootstrap replicates. Similar analyses of portions of the nucleotide sequences of the nonstructural protein gene (544 nucleotides sequenced) and the NP gene (885 nucleotides sequenced) further confirmed the identity of the virus as a North American-lineage equine influenza virus. This virus is now defined as A/Equine/Wisconsin/1/03. FIG. 1 provides sequences for the coding region of each gene of that virus.

The presence of influenza virus also was assessed in the lungs and other tissues of the foal. Specifically, immunohistochemistry with Mab 68D2 showed scattered, widely dispersed areas of influenza virus NP expression (predominantly localized around airways) in the frozen as well as the formalin-fixed lung tissue samples. NP expression was not shown in the other viscera or in the central nervous system. In addition, influenza virus was isolated in MDCK cells (and confirmed by immunocytochemistry and HA gene sequencing) from a sample of the frozen lung tissue.

Acute respiratory distress syndrome (ARDS) in neonatal foals has been documented as a consequence of bacterial sepsis (Wilkins, 2003; Hoffman et al., 1993), perinatal EHV-1 (Frymus et al., 1986; Gilkerson et al., 1999) and EHV-4 (Gilkerson et al., 1999), and equine viral arteritis infection (Del Piero et al., 1997). Less severe lower airway disease occasionally is documented with adenovirus and EHV-2 infections, particularly in the immunocompromised patient (Webb et al., 1981; Murray et al., 1996). Bronchointerstitial pneumonia and ARDS are high-mortality respiratory diseases of older foals with several potential causes, including bacterial and viral infections (Lakritz et al., 1993). Whether it occurs in neonates experiencing septic shock or

in older foals with diffuse bronchointerstitial pneumonia, ARDS is characterized by acute-onset, rapidly progressive, severe tachypnea. The increased respiratory effort, worsening cyanosis, hypoxemia, and hypercapnia that accompany ARDS frequently are poorly responsive to aggressive therapy (Wilkins, 2003; Lakritz et al., 1993). It is a category of respiratory disease with several potential etiologies and a mortality rate that frequently exceeds 30% despite intensive treatment with antimicrobials, oxygen, anti-inflammatory agents, bronchodilators, and thermoregulatory control. Equine influenza is a well-documented cause of upper respiratory disease in horses worldwide (Wilkins, 2003; Van Maanen et al., 2002; Wilson, 1993), but very little information exists in the literature about the manifestations of this disease in neonates. A single report describes bronchointerstitial pneumonia in a 7-day-old foal from which equine influenza A was isolated (Britton et al., 2002); this foal resembles the foal described herein.

The foal detailed in this study was one of several hospitalized horses that developed fever, mucopurulent nasal discharge, and coughing during a 2- or 3-week period. Clinical signs in the other affected horses, including high-risk neonates, generally were confined to the upper respiratory tract, except for mild systemic signs of fever and inappetence. The reason for the severity of the pulmonary failure in this foal is unclear. Treatment did include the potentially immunosuppressive drug dexamethasone and general anesthesia in a diagnostic procedure, both of which may have predisposed the foal to the development of pneumonia. The impact of the foal's neurologic disease on the development and progression of respiratory disease also is unclear. The histologic findings of diffuse vacuolization, decreased myelin throughout the central nervous system, and absent molecular layer within the cerebellum do not fit any specific clinical or histopathologic diagnosis. The foal could have had impaired central control of respiration, because the areas of the brain involved in the control of respiration (the pons and medulla oblongata) showed diffuse vacuolization and diminished myelin staining. Any subsequent impairment of ventilation would likely have been a terminal event given the normalcy of ventilatory function until several days after hospitalization. However, the abnormal mentation from birth, the vacuolization, the decreased myelination in the central nervous system, and the cerebellar abnormalities are suggestive of a concurrent, congenital neurologic abnormality, which may have compromised the foal's ability to respond to worsening respiratory function. The focal hemorrhage observed on the caudal ventral aspect of the brain was mild and was possibly a consequence of trauma during one of the seizures the foal experienced.

The mare had been vaccinated semiannually against influenza for the past 2 years with a killed product and was given a booster vaccination in late pregnancy. Considering the evidence of adequate passive transfer in this foal, these antibodies apparently did not confer adequate protection for the foal. Furthermore, phylogenetic analysis of the isolate obtained from the foal characterized it as an H3N8 subtype, and the commercial product used to vaccinate the mare in late pregnancy contained an influenza virus strain of the same subtype, suggesting that passive transfer cannot be guaranteed to protect against natural infection under certain circumstances. This lack of vaccine efficacy is consistent with a recent study by Mumford et al. (2003) that describes the failure of commercially available H7N7 and H3N8 equine influenza virus vaccines to protect adults against clinical respiratory disease that results from a natural infec-

tion with certain H3N8 virus strains. The transtracheal recovery of 2 bacterial species that were resistant to the antimicrobial regimen in place at the time of death confounds the conclusion that influenza was the sole cause of death. However, postmortem examination identified no gross or histopathologic evidence of sepsis, and synergism occurs between the influenza virus and some bacterial pathogens, combining to cause pneumonia with increased mortality (McCullers et al., 2003; Simonsen, 1999). Furthermore, the isolation of the infectious virus and the immunohistochemical demonstration of viral antigen from the lung tissue obtained postmortem, 6 days after the virus initially was recovered by a nasopharyngeal swab, provide strong evidence of a pathologic contribution from influenza virus in this foal's respiratory failure.

To compare the growth characteristics of avian, equine, human, and porcine lineage viruses in primary canine respiratory epithelial cells and to investigate the species influence on their growth characteristics, cultured cells were infected at an MOI of 3 with viruses including A/Equine/Wisconsin/1/03 and incubated for up to 10 hours. The other viruses included six human and swine influenza A virus isolates (A/Phillippines/08/98, A/Panama/2002/99, A/Costa Rica/07/99; A/Swine/NorthCarolina/44173/00, A/Swine/Minnesota/593/99, A/Swine/Ontario/00130/97, and two equine influenza viruses (A/Equine/Kentucky/81 and A/Equine/Kentucky/91). At the end of the experiment, the cells were formalin fixed for immunocytochemistry and flow cytometry analyses.

The six human and swine influenza virus isolates mentioned above readily infected substantially all (80-90%) of the canine respiratory epithelial cells and grew to high titers ($10^{5.3}$ - 10^7 TCID₅₀/ml) in those cells. A/Equine/Kentucky/81 and A/Equine/Kentucky/91 were highly restricted in their infectivity (<10% of the cells infected) with little ($10^{1.7}$ TCID₅₀/ml for A/Equine/Kentucky/81) or no (for A/Equine/Kentucky/91) detectable viral growth. In contrast, A/Equine/Wisconsin/1/03 infected a larger percentage (about 30%) of the primary canine respiratory epithelial cells and grew to substantially higher titers (about $10^{3.8}$ TCID₅₀/ml) in those cells. The results demonstrated that all influenza A viruses tested were able to infect canine primary respiratory epithelial cells. However, the infectivity and replication characteristics of the viruses were strongly lineage-dependent.

Dubovi et al. (2004) noted recurrent outbreaks of severe respiratory disease characterized by coughing and fever in greyhound dogs at racing kennels in Florida. Most affected dogs recovered, but some succumbed to a fatal hemorrhagic pneumonia. Lung tissues from 5 of the dogs that died from the hemorrhagic pneumonia syndrome were subjected to virus isolation studies in African green monkey kidney epithelial cells (Vero), Madin-Darby canine kidney epithelial cells (MDCK), primary canine kidney epithelial cells, primary canine lung epithelial cells, primary bovine testicular epithelial cells, canine tumor fibroblasts (A-72), and human colorectal adenocarcinoma epithelial cells (HRT-18) (Dubovi et al., 2004). Cytopathology in the MDCK cells was noted on the first passage of lung homogenate from one of the dogs, and the loss of cytopathology upon subsequent passage to cells cultured without trypsin coupled with the presence of hemagglutinating activity in culture supernatants suggested the presence of an influenza virus (Dubovi et al., 2004). The virus was initially identified as influenza virus by PCR using primers specific for the matrix gene. The canine influenza virus has been designated as the A/Canine/Florida/43/04 strain. Based on virus isolation from the lungs, the presence of viral antigens in lung tissues by immuno-

histochemistry, and seroconversion data, Dubovi et al. (2004) concluded that the isolated influenza virus was most likely the etiological agent responsible for the fatal hemorrhagic pneumonia in racing greyhounds during the Jacksonville 2004 outbreak, and that this was the first report of an equine influenza virus associated with respiratory disease in dogs (Dubovi et al., 2004). The HA protein of the canine isolate differs from the A/Equine/Wisconsin/1/03 strain by only 6 amino acids.

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All publications, patents and patent applications are incorporated herein by reference. While in the foregoing specification this invention has been described in relation to certain preferred embodiments thereof, and many details have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the details described herein may be varied considerably without departing from the basic principles of the invention.

SEQUENCE LISTING

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Leu Tyr Ile Gln Glu Ser Gly Arg Val Thr Val Ser Thr Lys Arg Ser
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Gln Gln Thr Ile Ile Pro Asn Ile Gly Ser Arg Pro Trp Val Arg Gly
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Gln Ser Gly Arg Ile Ser Ile Tyr Trp Thr Ile Val Lys Pro Gly Asp
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Phe Lys Leu Lys Thr Gly Lys Ser Ser Val Met Arg Ser Asp Val Pro
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Cys Asn Ile Cys Ile
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Tyr Tyr Met Asn Asn Thr Glu Pro Leu Cys Glu Ala Gln Gly Phe Ala
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Pro Phe Ser Lys Asp Asn Gly Ile Arg Ile Gly Ser Arg Gly His Val
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Phe Val Ile Arg Glu Pro Phe Val Ser Cys Ser Pro Ser Glu Cys Arg
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Val Thr Gly Pro Asp Asn Gln Ala Ile Ala Val Val Asn Tyr Gly Gly
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Cys Leu Val Pro Cys Phe Trp Val Glu Met Ile Arg Gly Lys Pro Glu
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Glu Thr Thr Ile Trp Thr Ser Ser Ser Ser Ile Val Met Cys Gly Val
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Gly Tyr Ala Gln Thr Asp Cys Val Leu Glu Ala Met Ala Phe Leu Glu
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Glu Ser His Pro Gly Ile Phe Glu Asn Ser Cys Leu Glu Thr Met Glu
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Tyr Asp Trp Thr Leu Asn Arg Asn Gln Pro Ala Ala Thr Ala Leu Ala
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Lys	Ser	Tyr	Ile	Asn	Arg	Thr	Gly	Thr	Phe	Glu	Phe	Thr	Ser	Phe	Phe
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Tyr	Arg	Tyr	Gly	Phe	Val	Ala	Asn	Phe	Ser	Met	Glu	Leu	Pro	Ser	Phe
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Gln	Met	Ala	Leu	Gln	Leu	Phe	Ile	Lys	Asp	Tyr	Arg	Tyr	Thr	Tyr	Arg
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Asp	Gly	Gly	Pro	Asn	Leu	Tyr	Asn	Ile	Arg	Asn	Leu	His	Ile	Pro	Glu
			595				600								605
Val	Cys	Leu	Lys	Trp	Glu	Leu	Met	Asp	Glu	Asp	Tyr	Lys	Gly	Arg	Leu
			610				615								620
Cys	Asn	Pro	Leu	Asn	Pro	Phe	Val	Ser	His	Lys	Glu	Ile	Glu	Ser	Val
			625				630				635				640
Asn	Ser	Ala	Val	Val	Met	Pro	Ala	His	Gly	Pro	Ala	Lys	Ser	Met	Glu
			645								650				655
Tyr	Asp	Ala	Val	Ala	Thr	Thr	His	Ser	Trp	Ile	Pro	Lys	Arg	Asn	Arg
			660								665				670

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Ser Ile Leu Asn Thr Ser Gln Arg Gly Ile Leu Glu Asp Glu Gln Met
 675 680 685

Tyr Gln Lys Cys Cys Asn Leu Phe Glu Lys Phe Phe Pro Ser Ser Ser
 690 695 700

Tyr Arg Arg Pro Val Gly Ile Ser Ser Met Val Glu Ala Met Val Ser
 705 710 715 720

Arg Ala Arg Ile Asp Ala Arg Ile Asp Phe Glu Ser Gly Arg Ile Lys
 725 730 735

Lys Asp Glu Phe Ala Glu Ile Met Lys Ile Cys Ser Thr Ile Glu Glu
 740 745 750

Leu Arg Arg Gln Lys
 755

<210> SEQ ID NO 4
 <211> LENGTH: 759
 <212> TYPE: PRT
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 4

Met Glu Arg Ile Lys Glu Leu Arg Asp Leu Met Leu Gln Ser Arg Thr
 1 5 10 15

Arg Glu Ile Leu Thr Lys Thr Thr Val Asp His Met Ala Ile Ile Lys
 20 25 30

Lys Tyr Thr Ser Gly Arg Gln Glu Lys Asn Pro Ala Leu Arg Met Lys
 35 40 45

Trp Met Met Ala Met Lys Tyr Pro Ile Thr Ala Asp Lys Arg Ile Met
 50 55 60

Glu Met Ile Pro Glu Arg Asn Glu Gln Gly Gln Thr Leu Trp Ser Lys
 65 70 75 80

Thr Asn Asp Ala Gly Ser Asp Arg Val Met Val Ser Pro Leu Ala Val
 85 90 95

Thr Trp Trp Asn Arg Asn Gly Pro Thr Thr Ser Thr Ile His Tyr Pro
 100 105 110

Lys Val Tyr Lys Thr Tyr Phe Glu Lys Val Glu Arg Leu Lys His Gly
 115 120 125

Thr Phe Gly Pro Val His Phe Arg Asn Gln Val Lys Ile Arg Arg Arg
 130 135 140

Val Asp Val Asn Pro Gly His Ala Asp Leu Ser Ala Lys Glu Ala Gln
 145 150 155 160

Asp Val Ile Met Glu Val Val Phe Pro Asn Glu Val Gly Ala Arg Ile
 165 170 175

Leu Thr Ser Glu Ser Gln Leu Thr Ile Thr Lys Glu Lys Lys Glu Glu
 180 185 190

Leu Gln Asp Cys Lys Ile Ala Pro Leu Met Val Ala Tyr Met Leu Glu
 195 200 205

Arg Glu Leu Val Arg Lys Thr Arg Phe Leu Pro Val Ala Gly Gly Thr
 210 215 220

Ser Ser Val Tyr Ile Glu Val Leu His Leu Thr Gln Gly Thr Cys Trp
 225 230 235 240

Glu Gln Met Tyr Thr Pro Gly Gly Glu Val Arg Asn Asp Asp Ile Asp
 245 250 255

Gln Ser Leu Ile Ile Ala Ala Arg Asn Ile Val Arg Arg Ala Thr Val
 260 265 270

Ser Ala Asp Pro Leu Ala Ser Leu Leu Glu Met Cys His Ser Thr Gln

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275					280					285					
Ile	Gly	Gly	Ile	Arg	Met	Val	Asp	Ile	Leu	Lys	Gln	Asn	Pro	Thr	Glu
	290						295					300			
Glu	Gln	Ala	Val	Asp	Ile	Cys	Lys	Ala	Ala	Met	Gly	Leu	Arg	Ile	Ser
305					310					315					320
Ser	Ser	Phe	Ser	Phe	Gly	Gly	Phe	Thr	Phe	Lys	Arg	Thr	Ser	Gly	Ser
			325						330					335	
Ser	Val	Lys	Arg	Glu	Glu	Glu	Met	Leu	Thr	Gly	Asn	Leu	Gln	Thr	Leu
			340						345				350		
Lys	Ile	Arg	Val	His	Glu	Gly	Tyr	Glu	Glu	Phe	Thr	Met	Val	Gly	Arg
		355					360						365		
Arg	Ala	Thr	Ala	Ile	Leu	Arg	Lys	Ala	Thr	Arg	Arg	Leu	Ile	Gln	Leu
	370					375					380				
Ile	Val	Ser	Gly	Arg	Asp	Glu	Gln	Ser	Ile	Ala	Glu	Ala	Ile	Ile	Val
385					390					395					400
Ala	Met	Val	Phe	Ser	Gln	Glu	Asp	Cys	Met	Ile	Lys	Ala	Val	Arg	Gly
				405					410					415	
Asp	Leu	Asn	Phe	Val	Asn	Arg	Ala	Asn	Gln	Arg	Leu	Asn	Pro	Met	His
			420					425					430		
Gln	Leu	Leu	Arg	His	Phe	Gln	Lys	Asp	Ala	Lys	Val	Leu	Phe	Gln	Asn
		435					440						445		
Trp	Gly	Ile	Glu	Pro	Ile	Asp	Asn	Val	Met	Gly	Met	Ile	Gly	Ile	Leu
	450					455					460				
Pro	Asp	Met	Thr	Pro	Ser	Thr	Glu	Met	Ser	Leu	Arg	Gly	Val	Arg	Val
465					470					475					480
Ser	Lys	Met	Gly	Val	Asp	Glu	Tyr	Ser	Ser	Thr	Glu	Arg	Val	Val	Val
				485					490					495	
Ser	Ile	Asp	Arg	Phe	Leu	Arg	Val	Arg	Asp	Gln	Arg	Gly	Asn	Ile	Leu
			500					505					510		
Leu	Ser	Pro	Glu	Glu	Val	Ser	Glu	Thr	Gln	Gly	Thr	Glu	Lys	Leu	Thr
		515					520					525			
Ile	Ile	Tyr	Ser	Ser	Ser	Met	Met	Trp	Glu	Ile	Asn	Gly	Pro	Glu	Ser
		530				535					540				
Val	Leu	Val	Asn	Thr	Tyr	Gln	Trp	Ile	Ile	Arg	Asn	Trp	Glu	Ile	Val
545					550					555					560
Lys	Ile	Gln	Trp	Ser	Gln	Asp	Pro	Thr	Met	Leu	Tyr	Asn	Lys	Ile	Glu
				565					570					575	
Phe	Glu	Pro	Phe	Gln	Ser	Leu	Val	Pro	Arg	Ala	Thr	Arg	Ser	Gln	Tyr
			580					585					590		
Ser	Gly	Phe	Val	Arg	Thr	Leu	Phe	Gln	Gln	Met	Arg	Asp	Val	Leu	Gly
		595				600						605			
Thr	Phe	Asp	Thr	Ala	Gln	Ile	Ile	Lys	Leu	Leu	Pro	Phe	Ala	Ala	Ala
		610				615					620				
Pro	Pro	Glu	Gln	Ser	Arg	Met	Gln	Phe	Ser	Ser	Leu	Thr	Val	Asn	Val
625					630					635					640
Arg	Gly	Ser	Gly	Met	Arg	Ile	Leu	Val	Arg	Gly	Asn	Ser	Pro	Val	Phe
				645					650					655	
Asn	Tyr	Asn	Lys	Ala	Thr	Lys	Arg	Leu	Thr	Val	Leu	Gly	Lys	Asp	Ala
			660					665					670		
Gly	Ala	Leu	Thr	Glu	Asp	Pro	Asp	Glu	Gly	Thr	Ala	Gly	Val	Glu	Ser
		675					680					685			
Ala	Val	Leu	Arg	Gly	Phe	Leu	Ile	Leu	Gly	Lys	Glu	Asn	Lys	Arg	Tyr
		690				695					700				

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Gly Pro Ala Leu Ser Ile Asn Glu Leu Ser Lys Leu Ala Lys Gly Glu
705 710 715 720

Lys Ala Asn Val Leu Ile Gly Gln Gly Asp Val Val Leu Val Met Lys
725 730 735

Arg Lys Arg Asp Ser Ser Ile Leu Thr Asp Ser Gln Thr Ala Thr Lys
740 745 750

Arg Ile Arg Met Ala Ile Asn
755

<210> SEQ ID NO 5
<211> LENGTH: 716
<212> TYPE: PRT
<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 5

Met Glu Asp Phe Val Arg Gln Cys Phe Asn Pro Met Ile Val Glu Leu
1 5 10 15

Ala Glu Lys Ala Met Lys Glu Tyr Gly Glu Asp Pro Lys Ile Glu Thr
20 25 30

Asn Lys Phe Ala Ala Ile Cys Thr His Leu Glu Val Cys Phe Met Tyr
35 40 45

Ser Asp Phe His Phe Ile Asn Glu Leu Ser Glu Ser Val Val Ile Glu
50 55 60

Ser Gly Asp Pro Asn Ala Leu Leu Lys His Arg Phe Glu Ile Ile Glu
65 70 75 80

Gly Arg Asp Arg Thr Met Ala Trp Thr Val Val Asn Ser Ile Cys Asn
85 90 95

Thr Thr Arg Ala Glu Lys Pro Lys Phe Leu Pro Asp Leu Tyr Asp Tyr
100 105 110

Lys Glu Asn Arg Phe Val Glu Ile Gly Val Thr Arg Arg Glu Val His
115 120 125

Ile Tyr Tyr Leu Glu Lys Ala Asn Lys Ile Lys Ser Glu Lys Thr His
130 135 140

Ile His Ile Phe Ser Phe Thr Gly Glu Glu Met Ala Thr Lys Ala Asp
145 150 155 160

Tyr Thr Leu Asp Glu Glu Ser Arg Ala Arg Ile Lys Thr Arg Leu Phe
165 170 175

Thr Ile Arg Gln Glu Met Ala Ser Arg Gly Leu Trp Asp Ser Phe Arg
180 185 190

Gln Ser Glu Arg Gly Glu Glu Thr Ile Glu Glu Arg Phe Glu Ile Thr
195 200 205

Gly Thr Met Arg Lys Leu Ala Asn Tyr Ser Leu Pro Pro Asn Phe Ser
210 215 220

Ser Leu Glu Asn Phe Arg Val Tyr Val Asp Gly Phe Glu Pro Asn Gly
225 230 235 240

Cys Ile Glu Ser Lys Leu Ser Gln Met Ser Lys Glu Val Asn Ala Arg
245 250 255

Ile Glu Pro Phe Ser Lys Thr Thr Pro Arg Pro Leu Lys Met Pro Gly
260 265 270

Gly Pro Pro Cys His Gln Arg Ser Lys Phe Leu Leu Met Asp Ala Leu
275 280 285

Lys Leu Ser Ile Glu Asp Pro Ser His Glu Gly Glu Gly Ile Pro Leu
290 295 300

Tyr Asp Ala Ile Lys Cys Met Lys Thr Phe Phe Gly Trp Lys Glu Pro

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305	310	315	320
Ser Ile Val Lys Pro His Glu Lys Gly Ile Asn Pro Asn Tyr Leu Gln	325	330	335
Thr Trp Lys Gln Val Leu Ala Glu Leu Gln Asp Leu Glu Asn Glu Glu	340	345	350
Lys Asp Pro Lys Thr Lys Asn Met Lys Lys Thr Ser Gln Leu Lys Trp	355	360	365
Ala Leu Ser Glu Asn Met Ala Pro Glu Lys Val Asp Phe Glu Asp Cys	370	375	380
Lys Asp Ile Ser Asp Leu Lys Gln Tyr Asp Ser Asp Glu Pro Glu Thr	385	390	395
Arg Ser Leu Ala Ser Trp Ile Gln Ser Glu Phe Asn Lys Ala Cys Glu	405	410	415
Leu Thr Asp Ser Ser Trp Ile Glu Leu Asp Glu Ile Gly Glu Asp Val	420	425	430
Ala Pro Ile Glu Tyr Ile Ala Ser Met Arg Arg Asn Tyr Phe Thr Ala	435	440	445
Glu Val Ser His Cys Arg Ala Thr Glu Tyr Ile Met Lys Gly Val Tyr	450	455	460
Ile Asn Thr Ala Leu Leu Asn Ala Ser Cys Ala Ala Met Asp Glu Phe	465	470	475
Gln Leu Ile Pro Met Ile Ser Lys Cys Arg Thr Lys Glu Gly Arg Arg	485	490	495
Lys Thr Asn Leu Tyr Gly Phe Ile Val Lys Gly Arg Ser His Leu Arg	500	505	510
Asn Asp Thr Asp Val Val Asn Phe Val Ser Met Glu Phe Ser Leu Thr	515	520	525
Asp Pro Arg Phe Glu Pro His Lys Trp Glu Lys Tyr Cys Val Leu Glu	530	535	540
Ile Gly Asp Met Leu Leu Arg Thr Ala Val Gly Gln Val Ser Arg Pro	545	550	555
Met Phe Leu Tyr Val Arg Thr Asn Gly Thr Ser Lys Ile Lys Met Lys	565	570	575
Trp Gly Met Glu Met Arg Arg Cys Leu Leu Gln Ser Leu Gln Gln Ile	580	585	590
Glu Ser Met Ile Glu Ala Glu Ser Ser Val Lys Glu Lys Asp Met Thr	595	600	605
Lys Glu Phe Phe Glu Asn Lys Ser Glu Thr Trp Pro Ile Gly Glu Ser	610	615	620
Pro Lys Gly Val Glu Glu Gly Ser Ile Gly Lys Val Cys Arg Thr Leu	625	630	635
Leu Ala Lys Ser Val Phe Asn Ser Leu Tyr Ala Ser Pro Gln Leu Glu	645	650	655
Gly Phe Ser Ala Glu Ser Arg Lys Leu Leu Leu Ile Val Gln Ala Leu	660	665	670
Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Ile Gly Gly Leu Tyr Glu	675	680	685
Ser Ile Glu Glu Cys Leu Ile Asn Asp Pro Trp Val Leu Leu Asn Ala	690	695	700
Ser Trp Phe Asn Ser Phe Leu Thr His Ala Leu Lys	705	710	715

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<211> LENGTH: 498

<212> TYPE: PRT

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 6

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Met Ala Ser Gln Gly Thr Lys Arg Ser Tyr Glu Gln Met Glu Thr Asp
 1          5          10          15
Gly Glu Arg Gln Asn Ala Thr Glu Ile Arg Ala Ser Val Gly Arg Met
 20          25          30
Val Gly Gly Ile Gly Arg Phe Tyr Val Gln Met Cys Thr Glu Leu Lys
 35          40          45
Leu Asn Asp His Glu Gly Arg Leu Ile Gln Asn Ser Ile Thr Ile Glu
 50          55          60
Arg Met Val Leu Ser Ala Phe Asp Glu Arg Arg Asn Lys Tyr Leu Glu
 65          70          75          80
Glu His Pro Ser Ala Gly Lys Asp Pro Lys Lys Thr Gly Gly Pro Ile
 85          90          95
Tyr Arg Arg Lys Asp Gly Lys Trp Met Arg Glu Leu Ile Leu His Asp
100          105          110
Lys Glu Glu Ile Met Arg Ile Trp Arg Gln Ala Asn Asn Gly Glu Asp
115          120          125
Ala Thr Ala Gly Leu Thr His Met Met Ile Trp His Ser Asn Leu Asn
130          135          140
Asp Thr Thr Tyr Gln Arg Thr Arg Ala Leu Val Arg Thr Gly Met Asp
145          150          155          160
Pro Arg Met Cys Ser Leu Met Gln Gly Ser Thr Leu Pro Arg Arg Ser
165          170          175
Gly Ala Ala Gly Ala Ala Val Lys Gly Val Gly Thr Met Val Met Glu
180          185          190
Leu Ile Arg Met Ile Lys Arg Gly Ile Asn Asp Arg Asn Phe Trp Arg
195          200          205
Gly Glu Asn Gly Arg Arg Thr Arg Ile Ala Tyr Glu Arg Met Cys Asn
210          215          220
Ile Leu Lys Gly Lys Phe Gln Thr Ala Ala Gln Arg Ala Met Met Asp
225          230          235          240
Gln Val Arg Glu Gly Arg Asn Pro Gly Asn Ala Glu Ile Glu Asp Leu
245          250          255
Ile Phe Leu Ala Arg Ser Ala Leu Ile Leu Arg Gly Ser Val Ala His
260          265          270
Lys Ser Cys Leu Pro Ala Cys Val Tyr Gly Leu Ala Val Thr Ser Gly
275          280          285
Tyr Asp Phe Glu Lys Glu Gly Tyr Ser Leu Val Gly Ile Asp Pro Phe
290          295          300
Lys Leu Leu Gln Asn Ser Gln Ile Phe Ser Leu Ile Arg Pro Lys Glu
305          310          315          320
Asn Pro Ala His Lys Ser Gln Leu Val Trp Met Ala Cys His Ser Ala
325          330          335
Ala Phe Glu Asp Leu Arg Val Leu Asn Phe Ile Arg Gly Thr Lys Val
340          345          350
Ile Pro Arg Gly Gln Leu Thr Thr Arg Gly Val Gln Ile Ala Ser Asn
355          360          365
Glu Asn Met Glu Thr Ile Asp Ser Ser Thr Leu Glu Leu Arg Ser Lys
370          375          380
Tyr Trp Ala Ile Arg Thr Arg Ser Gly Gly Asn Thr Ser Gln Gln Arg

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385          390          395          400
Ala Ser Ala Gly Gln Ile Ser Val Gln Pro Thr Phe Ser Val Gln Arg
          405          410          415
Asn Leu Pro Phe Glu Arg Ala Thr Ile Met Ala Ala Phe Thr Gly Asn
          420          425          430
Thr Glu Gly Arg Thr Ser Asp Met Arg Thr Glu Ile Ile Arg Met Met
          435          440          445
Glu Asn Ala Lys Ser Glu Asp Val Ser Phe Gln Gly Arg Gly Val Phe
          450          455          460
Glu Leu Ser Asp Glu Lys Ala Thr Asn Pro Ile Val Pro Ser Phe Asp
465          470          475          480
Met Ser Asn Glu Gly Ser Tyr Phe Phe Gly Asp Asn Ala Glu Glu Phe
          485          490          495

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Asp Ser

<210> SEQ ID NO 7

<211> LENGTH: 252

<212> TYPE: PRT

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 7

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Met Ser Leu Leu Thr Glu Val Glu Thr Tyr Val Leu Ser Ile Val Pro
 1          5          10          15
Ser Gly Pro Leu Lys Ala Glu Ile Ala Gln Arg Leu Glu Asp Val Phe
          20          25          30
Ala Gly Lys Asn Thr Asp Leu Glu Ala Leu Met Glu Trp Leu Lys Thr
          35          40          45
Arg Pro Ile Leu Ser Pro Leu Thr Lys Gly Ile Leu Gly Phe Val Phe
          50          55          60
Thr Leu Thr Val Pro Ser Glu Arg Gly Leu Gln Arg Arg Arg Phe Val
65          70          75          80
Gln Asn Ala Leu Ser Gly Asn Gly Asp Pro Asn Asn Met Asp Arg Ala
          85          90          95
Val Lys Leu Tyr Arg Lys Leu Lys Arg Glu Ile Thr Phe His Gly Ala
          100          105          110
Lys Glu Val Ala Leu Ser Tyr Ser Thr Gly Ala Leu Ala Ser Cys Met
          115          120          125
Gly Leu Ile Tyr Asn Arg Met Gly Thr Val Thr Thr Glu Val Ala Phe
          130          135          140
Gly Leu Val Cys Ala Thr Cys Glu Gln Ile Ala Asp Ser Gln His Arg
145          150          155          160
Ser His Arg Gln Met Val Thr Thr Thr Asn Pro Leu Ile Arg His Glu
          165          170          175
Asn Arg Met Val Leu Ala Ser Thr Thr Ala Lys Ala Met Glu Gln Met
          180          185          190
Ala Gly Ser Ser Glu Gln Ala Ala Glu Ala Met Glu Val Ala Ser Arg
          195          200          205
Ala Arg Gln Met Val Gln Ala Met Arg Thr Ile Gly Thr His Pro Ser
          210          215          220
Ser Ser Ala Gly Leu Lys Asp Asp Leu Leu Glu Asn Leu Gln Ala Tyr
225          230          235          240
Gln Lys Arg Met Gly Val Gln Met Gln Arg Phe Lys
          245          250

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<210> SEQ ID NO 8
 <211> LENGTH: 230
 <212> TYPE: PRT
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 8

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Met Asp Ser Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp
 1          5          10         15
His Val Arg Lys Arg Phe Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe
          20          25          30
Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser
          35          40          45
Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr His Ala Gly Lys Gln Ile
          50          55          60
Val Glu Gln Ile Leu Glu Lys Glu Ser Asp Glu Ala Leu Lys Met Thr
          65          70          75          80
Ile Ala Ser Val Pro Thr Ser Arg Tyr Leu Thr Asp Met Thr Leu Asp
          85          90          95
Glu Met Ser Arg Asp Trp Phe Met Leu Met Pro Lys Gln Lys Val Thr
          100         105         110
Gly Ser Leu Cys Ile Arg Met Asp Gln Ala Ile Met Asp Lys Asn Ile
          115         120         125
Ile Leu Lys Ala Asn Phe Ser Val Ile Phe Glu Arg Leu Glu Thr Leu
          130         135         140
Ile Leu Leu Arg Ala Phe Thr Glu Glu Gly Ala Val Val Gly Glu Ile
          145         150         155         160
Ser Pro Leu Pro Ser Leu Pro Gly His Thr Asn Glu Asp Val Lys Asn
          165         170         175
Ala Ile Gly Val Leu Ile Gly Gly Leu Lys Trp Asn Asp Asn Thr Val
          180         185         190
Arg Ile Ser Glu Thr Leu Gln Arg Phe Ala Trp Arg Ser Ser His Glu
          195         200         205
Asn Gly Arg Pro Ser Phe Pro Ser Lys Gln Lys Arg Lys Met Glu Arg
          210         215         220
Thr Ile Lys Pro Lys Ile
          225         230

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<210> SEQ ID NO 9
 <211> LENGTH: 1701
 <212> TYPE: DNA
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 9

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tcatgaagac aaccattatt ttgatactac tgaccattg ggcttacagt caaaacccaa    60
tcagtggcaa caacacagcc acattgtgtc tgggacacca tgcagtagca aatggaacat    120
tggtaaaaac aataagtgat gatcaaattg aggtgacaaa tgctacagaa ttagttcaaa    180
gcatttcaat ggggaaaata tgcaacaact catatagaat tctagatgga agaaattgca    240
cattaataga tgcaatgcta ggagaccccc actgtgacgc ctttcagtat gagaattggg    300
acctctttat agaaagaagc agcgctttca gcaattgcta cccatattgac atccctgact    360
atgcatcgct cccgatccatt gtagcatcct caggaacatt ggaattcaca gcagagggat    420
tcacatggac aggtgtcact caaaacggaa gaagtggagc ctgcaaaagg ggatcagccc    480
atagtttctt tagccgactg aattggctaa caaaatctgg aagctcttac cccacattga    540
atgtgacaat goctaacaat aaaaatttcg acaagctata catctggggg attcatcacc    600

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cgagctcaaa tcaagagcag acaaaattgt acatccaaga atcaggacga gtaacagtct 660
caacaaaaag aagtcaacaa acaataatcc ctaacatcgg atctagaccg tgggtcagag 720
gtcaatcagg taggataagc atatactgga ccattgtaaa acctggagat atcctaataga 780
taaacagtaa tggcaactta gttgcaccgc ggggatattt taaattgaaa acagggaaaa 840
gctctgtaat gagatcagat gtacccatag acatttgtgt gtctgaatgt attacaccaa 900
atggaagcat ctccaacgac aagccattcc aaaatgtgaa caaagttaaa tatggaaaaat 960
gccccaaagta tatcaggcaa aacactttaa agctggccac tgggatgagg aatgtaccag 1020
aaaagcaaat cagaggaatc tttggagcaa tagcgggatt catcgaaaac ggctgggaag 1080
gaatggttga tgggtggtat gggttccgat atcaaaactc tgaaggaaca gggcaagctg 1140
cagatctaaa gagcactcaa gcagccatcg accagattaa tggaaagtta aacagagtga 1200
ttgaaagaac caatgagaaa ttccatcaaa tagagaagga attctcagaa gtgaaaagaa 1260
gaattcagga cttggagaaa tatgtagaag acacaaaaat agacctatgg tcctacaatg 1320
cagaattgct ggtggctcta gaaaaatcaac atacaattga cttaacagat gcagaaatga 1380
ataaattatt tgagaagact agacgccagt taagagaaaa cgcagaagac atgggaggtg 1440
gatgtttcaa gatttaccac aaatgtgata atgcatgcat tggatcaata agaaatggga 1500
catatgacca ttacatatac agagatgaag cattaacaaa ccgatttcag atcaaaaggtg 1560
tagagttgaa atcaggctac aaagattgga tactgtggat ttcattcggc atatcatgct 1620
tettaatttg cgttgttcta ttgggtttca ttatgtgggc ttgccccaaa ggcaacatca 1680
gatgcaacat ttgcatttga g 1701

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<210> SEQ ID NO 10

<211> LENGTH: 1413

<212> TYPE: DNA

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 10

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atgaatccaa atcaaaagat aatagcaatt ggatttgcatt cattggggat attaatcatt 60
aatgtcattc tccatgtagt cagcattata gtaacagtac tggtoctcaa taacaataga 120
acagatctga actgcaaagg gacgatcata agagagtaca atgaaacagt aagagtagaa 180
aaaattactc aatggtataa taccagtaca attaagtaca tagagagacc ttcaaatgaa 240
tactacatga acaaacactga accactttgt gaggcccaag gctttgcacc attttccaaa 300
gataatggaa tacgaattgg gtcgagagggc catgtttttg tgataagaga accttttgta 360
tcatgttcgc cctcagaatg tagaaccttt ttcctcacac agggctcatt actcaatgac 420
aaacattcta acggcacagt aaaggaccga agtccgtata ggactttgat gagtgtcaaa 480
atagggcaat cacctaattg atataagct aggtttgaat cgggtggcatg gtcagcaaca 540
gcatgccatg atggaaaaaa atggatgaca gttggagtca cagggccccg caatcaagca 600
attgcagtag tgaactatgg aggtgttccg gttgatatta ttaattcatg ggcaggggat 660
atthtaagaa cccaagaatc atcatgcacc tgcattaaag gagactgtta ttgggtaatg 720
actgatggac cggcaaatag gcaagctaaa tataggatat tcaaagcaaa agatggaaga 780
gtaattggac agactgatat aagtttcaat gggggacaca tagaggagtg ttcttgttac 840
cccaatgaag ggaaggtgga atgcatatgc agggacaatt ggactggaac aaatagacca 900
attctggtaa tatcttctga tctatcgtac acagttggat atttgtgtgc tggcattccc 960

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actgacactc ctaggggaga ggatagtcaa ttcacaggct catgtacaag tcctttggga 1020
aataaaggat acggtgtaaa aggtttcggg ttcgacaag gaactgacgt atgggccgga 1080
aggacaatta gtaggacttc aagatcagga ttcgaaataa taaaaatcag gaatggttg 1140
acacagaaca gtaaagacca aatcaggagg caagtgatta tcgatgaccc aaattggtca 1200
ggatatagcg gttctttcac attgccggtt gaactaaca aaaagggatg tttggcccc 1260
tgtttctggg ttgaaatgat tagaggtaaa cctgaagaaa caacaatatg gacctctagc 1320
agctccattg tgatgtgtgg agtagatcat aaaattgcc a gttggtcatg gcacgatgga 1380
gctattcttc cctttgacat cgataagatg taa 1413

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<210> SEQ ID NO 11

<211> LENGTH: 2277

<212> TYPE: DNA

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 11

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atggatgtca atccgactct acttttctta aaggtgccag cgcaaatgc tataagcaca 60
acatttcctt atactggaga tcctccctac agtcatggaa caggacagc atacaccatg 120
gatactgtca acagaacaca ccaatattca gaaaaaggga aatggacaac aaacactgag 180
attggagcac cacaacttaa tccaatcgat ggaccacttc ctgaagacaa tgaaccaagt 240
gggtacgccc aaacagattg tgtattggaa gcaatggctt tccttgaaga atcccatccc 300
ggaatctttg aaaattcgtg tcttgaacg atggaggtga ttcagcagac aagagtggac 360
aaactaacac aaggccgaca aacttatgat tggaccttga ataggaatca acctgccgca 420
acagcacttg ctaatacgat tgaagtattc agatcaaatg gtctgacttc caatgaatcg 480
gggagattga tggacttctc caaagatgtc atggagtcca tgaacaagga agaaatggaa 540
ataacaacac acttccaacg gaagagaaga gtaagagaca acatgacaaa gagaatggta 600
acacagagaa ccatagggaa gaaaaaaca cgattaaaca gaaagagcta tctaatcaga 660
acattaaccc taaacacaat gaccaaggac gctgagagag ggaattgaa acgacgagca 720
atcgctacco cagggatgca gataagaggg tttgtatatt ttgttgaac actagcccga 780
agaatatgtg aaaagcttga acaatcagga ttgccagttg gcggtaatga gaaaaaggcc 840
aaactggcta atgtcgtcag aaaaatgatg actaattccc aagacactga actctccttc 900
accatcactg gggacaatac caaatggaat gaaaatcaga acccagcat attcctggca 960
atgatcacat acataactag aaaccagcca gaatggttca gaaatgttct aagcattgca 1020
ccgattatgt tctcaataa aatggcaaga ctggggaaag gatatatgtt tgaaagcaaa 1080
agtatgaaat tgagaactca aataccagca ggaatgcttg caagcattga cctgaaatat 1140
ttcaatgatc caacaaaaaa gaaattgaa aagatacgac cacttctggt tgacgggact 1200
gcttctactg atcctggcat gatgatggga atgttcaaca tgttgagcac tgtgctaggt 1260
gtatccatat taaacctggg ccagaggaaa tacacaaaga ccacatactg gtgggatggt 1320
ctgcaatcat ccgatgactt tgctttgata gtgaatgctc ctaatcatga aggaatacaa 1380
gctggagtag acagattcta taggacttgc aaactggctg ggatcaacat gagcaaaaag 1440
aagtccatac taaatagaac tggaacattc gaattcaaaa gctttttcta ccggtatggt 1500
ttttagacca atttcagcat ggaactacce agttttgggg tttccggaat aaatgaatct 1560
gcagacatga gcattggagt gacagtcac aaaaaaaca tgataataa tgatctcgggt 1620
cctgccaccg cacaaatggc actccaactc ttcattaagg attatcggt a cacataccgg 1680

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tgccatagag	gtgataccca	gatacaaaacc	agaagatctt	ttgagttgaa	gaaactgtgg	1740
gaacagactc	gatcaaagac	tgggtctactg	gtatcagatg	ggggtccaaa	cctatataac	1800
atcagaaaacc	tacacatccc	ggaagtctgt	ttaaaatggg	agctaatagga	tgaagattat	1860
aaggggagcg	tatgcaatcc	attgaatcct	ttcgtagtc	acaaagaaat	tgaatcagtc	1920
aacagtgcag	tagtaatgcc	tgcgcatggc	cctgccaaaa	gcatggagta	tgatgctggt	1980
gcaacaacac	attcttggat	ccccaaagag	aaccggtcca	tattgaacac	aagccaaagg	2040
ggaatactcg	aagatgagca	gatgtatcag	aaatgctgca	acctgtttga	aaaattcttc	2100
cccagcagct	catacagaag	accagtcggg	atctctagta	tggttgaggc	catggtgtcc	2160
agggcccgca	ttgatgcacg	aattgacttc	gaatctggac	ggataaagaa	ggatgagttc	2220
gctgagatca	tgaagatctg	ttccaccatt	gaagagctca	gacggcaaaa	atagtga	2277

<210> SEQ ID NO 12

<211> LENGTH: 2281

<212> TYPE: DNA

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 12

atggagagaa	taaaagaact	gagagatctg	atgttacaat	cccgaccg	cgagatacta	60
acaaaaacta	ctgtggacca	catggccata	atcaagaaat	acacatcagg	aagacaagag	120
aagaaccctg	cacttaggat	gaaatggatg	atggcaatga	aatacccaat	tacagcagat	180
aagaggataa	tggagatgat	tctgagaga	aatgaacagg	gacaaacctt	ttggagcaaa	240
acgaacgatg	ctggctcaga	ccgctaatg	gtatcacctc	tggcagtgac	atggtggaat	300
aggaatggac	caacaacaag	cacaattcat	tatccaaaag	tctacaaaac	ttattttgaa	360
aaggttgaaa	gattgaaaca	cggaaacctt	ggccccgttc	atcttaggaa	tcaagtcaag	420
ataagacgaa	gagttgatgt	aaacctggt	cacgcgacc	tcagtgccaa	agaagcacia	480
gatgtgatca	tggagattgt	tttcccaat	gaagtgggag	ccagaattct	aacatcggaa	540
tcacaactaa	caataaccaa	agagaaaaag	gaagaacttc	aggactgcaa	aattgctccc	600
ttgatggtag	catacatgct	agaaagagag	ttggtccgaa	aaacaagggt	cctcccagta	660
gcaggcggaa	caagcagtg	atacattgaa	gtgttgcac	tgactcaggg	aacatgctgg	720
gagcaaatgt	acaccccagg	aggagaagtt	agaaacgatg	atattgatca	aagtttaatt	780
attgcagccc	ggaacatagt	gagaagagca	acagtatcag	cagatccact	agcatcccta	840
ctggaatgt	gccacagtac	acagattggt	ggaataagga	tggtagacat	ccttaagcag	900
aatccaacag	aggaacaagc	tgtggatata	tgcaaagcag	caatgggatt	gagaattagc	960
tcacattca	gctttggtg	attcaccttc	aagagaacaa	gtggatcatc	agtcaagaga	1020
gaagaagaaa	tgcttacggg	caaccttcaa	acattgaaaa	taagagtgca	tgagggctat	1080
gaagaattca	caatggctcg	aagaagagca	acagccattc	tcagaaaggc	aaccagaaga	1140
ttgattcaat	tgatagtaag	tgggagagat	gaacagtcaa	ttgctgaagc	aataattgta	1200
gccatggtgt	tttcgcaaga	agattgcatg	ataaaagcag	ttcgaggcga	tttgaacttt	1260
gttaatagag	caaatcagcg	cttgaacccc	atgcatcaac	tcttgaggca	tttccaaaag	1320
gatgcaaaa	tgcttttcca	aaattggggg	attgaaccca	tcgacaatgt	aatgggaatg	1380
attggaatat	tgccctgacat	gaccccagc	accgagatgt	cattgagagg	agtgagagtc	1440
agcaaaatgg	gagtgatga	gtactccagc	actgagagag	tggtggtgag	cattgaccgt	1500

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ttttaagag ttcgggatca aaggggaaac atactactgt cccctgaaga agtcagtgaa 1560
acacaaggaa cggaaaagct gacaataatt tattcgtcat caatgatgtg ggagattaat 1620
ggtcccgaat cagtgttggt caatacttat caatggatca tcaggaactg ggaaattgta 1680
aaaattcagt ggtcacagga ccccaaatg ttatacaata agatagaatt tgagccattc 1740
caatccctgg tccctagggc taccagaagc caatacagcg gtttcgtaag aaccctgttt 1800
cagcaaatgc gagatgtact tggaacatth gatactgctc aaataataaa actcctccct 1860
tttgccgctg ctctccgga acagagtagg atgcagttct cttctttgac tgtaaatgta 1920
agaggttcgg gaatgaggat acttgtaaga ggcaattccc cagtgttcaa ctacaataaa 1980
gccactaaaa ggctcacagt cctcggaaag gatgcagggt cgcttactga ggaccagat 2040
gaaggtacgg ctggagtaga atctgctggt ctaagagggt ttctcattt aggtaaagaa 2100
aataagagat atggcccagc actaagcadc aatgaactaa gcaacttgc aaaaggggag 2160
aaagccaatg tactaattgg gcaaggggac gtagtgttgg taatgaaacg gaaacgtgac 2220
tctagcatac ttactgacag ccagacagcg accaaaagga ttcggatggc catcaattag 2280
t 2281

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<210> SEQ ID NO 13

<211> LENGTH: 2209

<212> TYPE: DNA

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 13

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atggaagact ttgtgcgaca atgcttcaat ccaatgatcg tcgagcttgc ggaaaaggca 60
atgaaagaat atggagagga cccgaaaatc gaaacaaaca aatttgagc aatagcact 120
cacttggaag tctgcttcat gtactcggat ttccacttta ttaatgaact gagtgagtca 180
gtggtcatag agtctggtga cccaaatgct cttttgaaac acagatttga aatcattgag 240
gggagagatc gaacaatggc atggacagta gtaaacagca tctgcaacac cacaagagct 300
gaaaaaccta aatttcttcc agatttatac gactataagg agaacagatt tghtgaaatt 360
gggtgacaaa ggagagaagt tcacatatac tacttgaga aggccaacaa aataaagtct 420
gagaaaaacac atatccacat tttctcattt acaggagagg aatggctac aaaagcggac 480
tatactcttg atgaagagag tagagccagg atcaagacca gactattcac tataagacaa 540
gaaatggcca gtagaggcct ctgggattcc tttcgtcagt ccgagagagg cgaagagaca 600
attgaagaaa gatttgaat cacagggagc atgcgcaagc ttgccaatta cagtctccca 660
ccgaacttct ccagccttga aaattttaga gtctatgtgg atggattcga accgaacggc 720
tgcattgaga gtaagctttc tcaaatgtcc aaagaagtaa atgccagaat cgaaccattt 780
tcaaagacaa caccocgacc actcaaaatg ccaggtggtc caccctgcca tcagcgatct 840
aaattcctgc taatggatgc tctgaaactg agcattgagg acccaagtca cgagggagag 900
ggaataccac tatatgatgc catcaaatgc atgaaaactt tctttggatg gaaagagccc 960
agtattgtta aaccacatga aaaggtata aaccggaact atctccaac ttggaagcaa 1020
gtattagcag aattacaaga ccttgagaac gaagaaaagg accccaagac caagaatatg 1080
aaaaaaacaa gccaatgaa atgggcactt agtgaaaata tggcaccaga gaaagtggat 1140
tttgaggatt gtaaagacat cagtgattta aaacagtatg acagtgatga gccagaaaca 1200
aggtctcttg caagttggat tcaaagttag ttcaacaaag cttgtgaaact gacagattca 1260
agctggatag agctcgatga aattggggag gatgttgccc caatagaata cattcgcagc 1320

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atgaggagaa attatnttac tgctgaggtt tccattgta gagcaacaga atatataatg 1380
aaggagagt acatcaacac tgctctactc aatgcatcct gtgctgcat ggatgaattc 1440
caattaatc cgatgataag taaatgcagg accaaagaag ggagaaggaa gacaaattta 1500
tatggattca tagtaaaggg aaggtcccat ttaagaaatg atactgacgt ggtgaacttt 1560
gtaagtatgg aatnttctct cactgatcca agatttgagc cacacaaatg ggaaaaatac 1620
tgcttctag aamtggaga catgcttota agaactgctg taggtcaagt gtcaagacc 1680
atgntttgt atgtaaggac aatggaacc tctaaaatta aatgaaatg gggaatggaa 1740
atgaggcgt gctccttca gtctctgcaa cagattgaaa gcatgatcga agctgagtc 1800
tcagtcaaag aaaaggacat gaccaaagaa tnttttgaga acaaatcaga gacatggcct 1860
ataggagagt ccccaagg agtggagag ggtcaatcg ggaaggttg caggacctta 1920
ttagcaaat ctgtgtttaa cagttgtat gcatctccac aactggaagg gtttcagct 1980
gaatctagga aattacttct cattgttcag gctcttagg ataactgga acctggaacc 2040
ttgatattg ggggttata tgaatcaatt gaggagtgc tgattaatga tccctgggt 2100
ttgctaatg catcttggt caactcttc cttacacatg cactgaagta gttgtggcaa 2160
tgctactatt tgctatccat actgtccaaa aaagtacctt gtttctact 2209

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<210> SEQ ID NO 14

<211> LENGTH: 1498

<212> TYPE: DNA

<213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 14

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atggcgtctc aaggcaccac acgatcctat gaacagatgg aaactgatgg ggaacgccag 60
aatgcaactg aatcagagc atctgtcgga aggatggtg gaggaatcg ccggttttat 120
gttcagatgt gtactgagct taaactaac gaccatgaag ggcgctgat tcagaacagc 180
ataacaatag aaaggatggt actttcgca ttcgacgaaa gaagaacaa gtatctcgag 240
gagcatccca gtgctgggaa agaccctaa aaaacaggag gcccgatata cagaaggaaa 300
gatgggaaat ggatgagga actcatctc catgataaag aagaaatcat gagaatctgg 360
cgtcaggcca acaatggtga agacgctact gctggtctta ctcatatgat gatctggcac 420
tccaatctca atgacaccac ataccaaaaga acaagggtc ttgttcggac tgggatggat 480
cccagaatgt gctctctgat gcaaggctca accctccac ggagatctgg agccgctggt 540
gtgacagtaa aagtggttg aacaatggt atggaactca tcagaatgat caaacgcgga 600
ataaatgatc ggaatntctg gagaggtgaa aatggtcgaa gaaccagaat tgcttatgaa 660
agaatgtgca atatctcaa agggaaatnt cagacagcag cacaacgggc tatgatggac 720
caggtgaggg aaggccgcaa tcttgaaac gctgagattg aggatctcat tntctggca 780
cgatcagcac ttatnttgag aggatcagta gcccataaat catgcctacc tgctgtggt 840
tatggccttg cagtaaccag tgggtatgac tttgagaagg aaggatactc tctggttgg 900
attgatcctt tcaactact ccagaacagt caaatnttca gtctaactag accaaaagaa 960
aaccagcac acaagagcca gttggtggt atggcatgcc attctgcagc attgaggac 1020
ctgagagttt taaatntcat tagaggaacc aaagtaatcc caagaggaca gttaacaacc 1080
agaggagttc aatagcttc aatgaaaac atggagacaa tagattctag cacacttgaa 1140
ctgagaagca aatattgggc aataaggacc agaagcggag gaaacaccag tcaacagaga 1200

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gcatctgcag gacagataag tgtgcaacct actttctcag tacagagaaa tcttccttt	1260
gagagagcaa ccattatggc tgcattcact ggtaacactg aaggaggac ttccgacatg	1320
agaacggaaa tcataaggat gatggaaaat gccaaatcag aagatgtgtc tttccagggg	1380
cggggagtct tcgagctctc ggacgaaaag gcaacgaacc cgatcgtgcc ttctttgac	1440
atgagcaatg aagggtctta tttcttcgga gacaatgctg aggagtgtga cagttaa	1498

<210> SEQ ID NO 15
 <211> LENGTH: 982
 <212> TYPE: DNA
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 15

atgagtcttc taaccgaggt cgaaacgtac gttctctcta tcgtaccatc agggcccctc	60
aaagccgaga tcgcgagag acttgaagat gtctttgcag ggaagaacac cgatcctgag	120
gcaactcatgg aatggctaaa gacaagacca atcctgtcac ctctgactaa agggatttta	180
ggatttगत tcaagctcac cgtgccagt gagcgaggac tgcagcgtag acgctttgtc	240
caaaatgccc ttagtgaaa cggagatcca aacaacatgg acagagcagt aaaactgtac	300
aggaagctta aaagagaaat aacattccat ggggcaaaag aggtggcact cagctattcc	360
actggtgcac tagccagctg catgggactc atatacaaca gaatgggaac tgttacaacc	420
gaagtggcat ttggcctggt atgcgccaca tgtgaacaga ttgctgattc ccagcatcgg	480
tctcacaggc agatgggtgac aacaaccaac ccattaatca gacatgaaa cagaatggta	540
ttagccagta ccacggctaa agccatggaa cagatggcag gatcagatga gcaggcagca	600
gaggccatgg aggttgctag tagggctagg cagatggtag agggcaatgag aaccattggg	660
accacccta gctccagtgc cggtttgaaa gatgatctcc ttgaaaattt acaggcctac	720
cagaaacgga tgggagtgca aatgcagcga ttcaagtgat cctctcgta tgcagcaag	780
tatcattggg atcttgcact tgatattgtg gattcttgat cgtctttct tcaaattcat	840
ttatcgtcgc cttaaatagc ggttgaaaag agggccttct acggaaggag tacctgagtc	900
tatgagggaa gaatatcggc aggaacagca gaatgctgtg gatgttgacg atggtcattt	960
tgtcaacata gagctggagt aa	982

<210> SEQ ID NO 16
 <211> LENGTH: 838
 <212> TYPE: DNA
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 16

atggattcca aactgtgtc aagctttcag gtagactggt ttctttggca tgtccgaaa	60
cgattcgcag accaagaact gggatgatgcc ccattccttg accggcttcg ccgagaccag	120
aagtccctaa ggggaagagg tagcactctt ggtctggaca tcgaaacagc cactcatgca	180
ggaaagcaga tagtggagca gattctggaa aaggaatcag atgaggcact taaaatgacc	240
attgcctctg ttcctacttc acgctactta actgacatga ctcttgatga gatgtcaaga	300
gactggttca tgctcatgcc caagcaaaaa gtaacaggct ccctatgtat aagaatggac	360
caggcaatca tggataagaa catcactact aaagcaaaact ttagtgtgat tttcgaaagg	420
ctggaacac taatactact tagagccttc accgaagaag gagcagtcgt tggcgaaatt	480
tcaccattac cttctcttcc aggacatact aatgaggatg tcaaaaatgc aattggggtc	540
ctcatcggag gacttaaatg gaatgataat acggttagaa tctctgaaac tctacagaga	600

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ttogcttga gaagcagtc tgagaatggg agaccttcat tcccttcaa gcagaaaaga 660
aaatggaga gaacaattaa gcaaaaaatt tgaagaaata agatggttga ttgaagaagt 720
gcgacataga ttgaaaaata cagaaaatag ttttgaacaa ataacattta tgcaagcctt 780
acaactattg cttgaagtag aacaagagat aagaactttc tcgtttcagc ttatttaa 838

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<210> SEQ ID NO 17
<211> LENGTH: 97
<212> TYPE: PRT
<213> ORGANISM: Influenza A Virus

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<400> SEQUENCE: 17

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Met Ser Leu Leu Thr Glu Val Glu Thr Pro Thr Arg Asn Gly Trp Glu
 1           5              10             15
Cys Lys Cys Ser Asp Ser Ser Asp Pro Leu Val Ile Ala Ala Ser Ile
 20          25          30
Ile Gly Ile Leu His Leu Ile Leu Trp Ile Leu Asp Arg Leu Phe Phe
 35          40          45
Lys Phe Ile Tyr Arg Arg Leu Lys Tyr Gly Leu Lys Arg Gly Pro Ser
 50          55          60
Thr Glu Gly Val Pro Glu Ser Met Arg Glu Glu Tyr Arg Gln Glu Gln
 65          70          75          80
Gln Asn Ala Val Asp Val Asp Asp Gly His Phe Val Asn Ile Glu Leu
 85          90          95

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Glu

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<210> SEQ ID NO 18
<211> LENGTH: 119
<212> TYPE: PRT
<213> ORGANISM: Influenza A Virus

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<400> SEQUENCE: 18

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Met Asp Ser Asn Thr Val Ser Ser Phe Gln Leu Met Arg Met Ser Lys
 1           5              10             15
Met Gln Leu Gly Ser Ser Ser Glu Asp Leu Asn Gly Met Ile Ile Arg
 20          25          30
Leu Glu Ser Leu Lys Leu Tyr Arg Asp Ser Leu Gly Glu Ala Val Met
 35          40          45
Arg Met Gly Asp Leu His Ser Leu Gln Ser Arg Asn Glu Lys Trp Arg
 50          55          60
Glu Gln Leu Ser Gln Lys Phe Glu Glu Ile Arg Trp Leu Ile Glu Glu
 65          70          75          80
Val Arg His Arg Leu Lys Asn Thr Glu Asn Ser Phe Glu Gln Ile Thr
 85          90          95
Phe Met Gln Ala Leu Gln Leu Leu Leu Glu Val Glu Gln Glu Ile Arg
 100         105         110
Thr Phe Ser Phe Gln Leu Ile
 115

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<210> SEQ ID NO 19
<211> LENGTH: 344
<212> TYPE: PRT
<213> ORGANISM: Influenza A Virus

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<400> SEQUENCE: 19

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Met Lys Thr Thr Ile Ile Leu Ile Leu Leu Thr His Trp Ala Tyr Ser
 1           5              10             15

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Gln Asn Pro Ile Ser Gly Asn Asn Thr Ala Thr Leu Cys Leu Gly His
 20 25 30

His Ala Val Ala Asn Gly Thr Leu Val Lys Thr Ile Ser Asp Asp Gln
 35 40 45

Ile Glu Val Thr Asn Ala Thr Glu Leu Val Gln Ser Ile Ser Met Gly
 50 55 60

Lys Ile Cys Asn Asn Ser Tyr Arg Ile Leu Asp Gly Arg Asn Cys Thr
 65 70 75 80

Leu Ile Asp Ala Met Leu Gly Asp Pro His Cys Asp Ala Phe Gln Tyr
 85 90 95

Glu Asn Trp Asp Leu Phe Ile Glu Arg Ser Ser Ala Phe Ser Asn Cys
 100 105 110

Tyr Pro Tyr Asp Ile Pro Asp Tyr Ala Ser Leu Arg Ser Ile Val Ala
 115 120 125

Ser Ser Gly Thr Leu Glu Phe Thr Ala Glu Gly Phe Thr Trp Thr Gly
 130 135 140

Val Thr Gln Asn Gly Arg Ser Gly Ala Cys Lys Arg Gly Ser Ala Asp
 145 150 155 160

Ser Phe Phe Ser Arg Leu Asn Trp Leu Thr Lys Ser Gly Ser Ser Tyr
 165 170 175

Pro Thr Leu Asn Val Thr Met Pro Asn Asn Lys Asn Phe Asp Lys Leu
 180 185 190

Tyr Ile Trp Gly Ile His His Pro Ser Ser Asn Gln Glu Gln Thr Lys
 195 200 205

Leu Tyr Ile Gln Glu Ser Gly Arg Val Thr Val Ser Thr Lys Arg Ser
 210 215 220

Gln Gln Thr Ile Ile Pro Asn Ile Gly Ser Arg Pro Trp Val Arg Gly
 225 230 235 240

Gln Ser Gly Arg Ile Ser Ile Tyr Trp Thr Ile Val Lys Pro Gly Asp
 245 250 255

Ile Leu Met Ile Asn Ser Asn Gly Asn Leu Val Ala Pro Arg Gly Tyr
 260 265 270

Phe Lys Leu Lys Thr Gly Lys Ser Ser Val Met Arg Ser Asp Val Pro
 275 280 285

Ile Asp Ile Cys Val Ser Glu Cys Ile Thr Pro Asn Gly Ser Ile Ser
 290 295 300

Asn Asp Lys Pro Phe Gln Asn Val Asn Lys Val Thr Tyr Gly Lys Cys
 305 310 315 320

Pro Lys Tyr Ile Arg Gln Asn Thr Leu Lys Leu Ala Thr Gly Met Arg
 325 330 335

Asn Val Pro Glu Lys Gln Ile Arg
 340

<210> SEQ ID NO 20
 <211> LENGTH: 344
 <212> TYPE: PRT
 <213> ORGANISM: Influenza A Virus

<400> SEQUENCE: 20

Met Lys Thr Thr Ile Ile Leu Ile Leu Leu Thr His Trp Ala Tyr Ser
 1 5 10 15

Gln Asn Pro Ile Ser Gly Asn Asn Thr Ala Thr Leu Cys Leu Gly His
 20 25 30

His Ala Val Ala Asn Gly Thr Leu Val Lys Thr Ile Ser Asp Asp Gln

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	35					40					45				
Ile	Glu	Val	Thr	Asn	Ala	Thr	Glu	Leu	Val	Gln	Ser	Ile	Ser	Met	Gly
	50					55					60				
Lys	Ile	Cys	Asn	Asn	Ser	Tyr	Arg	Ile	Leu	Asp	Gly	Arg	Asn	Cys	Thr
65					70					75					80
Leu	Ile	Asp	Ala	Met	Leu	Gly	Asp	Pro	His	Cys	Asp	Val	Phe	Gln	Tyr
				85					90					95	
Glu	Asn	Trp	Asp	Leu	Phe	Ile	Glu	Arg	Ser	Ser	Ala	Phe	Ser	Asn	Cys
			100					105					110		
Tyr	Pro	Tyr	Asp	Ile	Pro	Asp	Tyr	Ala	Ser	Leu	Arg	Ser	Ile	Val	Ala
		115					120						125		
Ser	Ser	Gly	Thr	Leu	Glu	Phe	Thr	Ala	Glu	Gly	Phe	Thr	Trp	Thr	Gly
	130					135					140				
Val	Thr	Gln	Asn	Gly	Arg	Ser	Gly	Ala	Cys	Lys	Arg	Gly	Ser	Ala	Asp
145					150						155				160
Ser	Phe	Phe	Ser	Arg	Leu	Asn	Trp	Leu	Thr	Lys	Ser	Gly	Asn	Ser	Tyr
				165					170						175
Pro	Thr	Leu	Asn	Val	Thr	Met	Pro	Asn	Asn	Lys	Asn	Phe	Asp	Lys	Leu
			180					185					190		
Tyr	Ile	Trp	Gly	Ile	His	His	Pro	Ser	Ser	Asn	Gln	Glu	Gln	Thr	Lys
		195					200						205		
Leu	Tyr	Ile	Gln	Glu	Ser	Gly	Arg	Val	Thr	Val	Ser	Thr	Lys	Arg	Ser
	210					215						220			
Gln	Gln	Thr	Ile	Ile	Pro	Asn	Ile	Gly	Ser	Arg	Pro	Trp	Val	Arg	Gly
225					230					235					240
Gln	Ser	Gly	Arg	Ile	Ser	Ile	Tyr	Trp	Thr	Ile	Val	Lys	Pro	Gly	Asp
			245						250						255
Ile	Leu	Met	Ile	Asn	Ser	Asn	Gly	Asn	Leu	Val	Ala	Pro	Arg	Gly	Tyr
			260					265							270
Phe	Lys	Leu	Lys	Thr	Gly	Lys	Ser	Ser	Val	Met	Arg	Ser	Asp	Ala	Pro
	275						280						285		
Ile	Asp	Ile	Cys	Val	Ser	Glu	Cys	Ile	Thr	Pro	Asn	Gly	Ser	Ile	Ser
	290					295					300				
Asn	Asp	Lys	Pro	Phe	Gln	Asn	Val	Asn	Lys	Val	Thr	Tyr	Gly	Lys	Cys
305					310					315					320
Pro	Lys	Tyr	Ile	Arg	Gln	Asn	Thr	Leu	Lys	Leu	Ala	Thr	Gly	Met	Arg
				325					330						335
Asn	Val	Pro	Glu	Lys	Gln	Ile	Arg								
															340

What is claimed is:

1. A vaccine comprising an isolated H3 influenza virus comprising a gene segment with sequences for a HA or a HA-1 having at least 95% amino acid sequence identity to SEQ ID NO:1, where the HA or the HA-1 has an alanine at position 78 and a serine at position 159, and has a residue other than methionine at position 29, a residue other than lysine at position 54, a residue other than serine at position 83, a residue other than asparagine at position 92, a residue other than leucine at position 222, a residue other than alanine at position 272, or a residue other than threonine at position 328, in an amount effective to induce a prophylactic or therapeutic response against influenza infection, wherein the virus is inactivated, the vaccine is in freeze-dried form or the vaccine further comprises an adjuvant.

2. The vaccine of claim 1 wherein the HA or HA-1 has at least 96% amino acid sequence identity to SEQ ID NO:1.

3. The vaccine of claim 1 wherein the HA or HA-1 has at least 99% amino acid sequence identity to SEQ ID NO:1.

4. The vaccine of claim 1 further comprising a different isolated influenza virus.

5. The vaccine of claim 1 wherein the virus is inactivated.

6. The vaccine of claim 1 which comprises the adjuvant.

7. The vaccine of claim 1 further comprising a pharmaceutically acceptable carrier.

8. The vaccine of claim 7 wherein the carrier is suitable for intranasal or intramuscular administration.

9. The vaccine of claim 1 which is in freeze-dried form.

10. The vaccine of claim 1 wherein the virus comprises at least one of the following gene segments: a gene segment with sequences for a NA having SEQ ID NO:2 or having at least 95% amino acid sequence identity to SEQ ID NO:2, a gene segment with sequences for a PB1 having SEQ ID

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NO:3 or having at least 95% amino acid sequence identity to SEQ ID NO:3, a gene segment with sequences for a PB2 having SEQ ID NO:4 or having at least 95% amino acid sequence identity to SEQ ID NO:4, a gene segment with sequences for a PA having SEQ ID NO:5 or having at least 95% amino acid sequence identity to SEQ ID NO:5, a gene segment with sequences for a NP having SEQ ID NO:6 or having at least 95% amino acid sequence identity to SEQ ID NO:6, a gene segment with sequences for a M1 having SEQ ID NO:7 or having at least 95% amino acid sequence identity to SEQ ID NO:7, a gene segment with sequences for a M2 having SEQ ID NO:17 or having at least 95% amino acid sequence identity to SEQ ID NO:17, a gene segment with sequences for a NS1 having SEQ ID NO:8 or having at least 95% amino acid sequence identity to SEQ ID NO:8, or a gene segment with sequences for a NS2 having SEQ ID NO:18 or having at least 95% amino acid sequence identity to SEQ ID NO:18.

11. A method to immunize a mammal against influenza, comprising administering to the mammal an effective amount of a composition comprising an H3 influenza virus comprising a gene segment with sequences for a HA or a HA-1 having at least 95% amino acid sequence identity to SEQ ID NO:1, wherein the HA or the HA-1 has an alanine at position 78 and a serine at position 159, and has a residue other than methionine at position 29, a residue other than lysine at position 54, a residue other than serine at position 83, a residue other than asparagine at position 92, a residue other than leucine at position 222, a residue other than alanine at position 272, or a residue other than threonine at position 328.

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12. The method of claim 11 wherein the mammal is a dog.

13. The method of claim 11 wherein the mammal is a horse.

14. The method of claim 11 wherein the composition further comprises a different influenza virus.

15. The method of claim 11 wherein the H3 influenza virus is an attenuated virus or is a reassortant virus.

16. The method of claim 11 wherein the composition further comprises an adjuvant or a pharmaceutically acceptable carrier.

17. The method of claim 16 wherein the carrier is suitable for intranasal or intramuscular administration.

18. The method of claim 11 wherein the composition further comprises a pathogen other than the H3 influenza virus.

19. An immunogenic composition comprising an isolated HA polypeptide having a HA polypeptide having at least 95% amino acid sequence identity to SEQ ID NO:1, or a HA-1 portion of the HA polypeptide, wherein the HA or the HA-1 has an alanine at position 78 and a serine at position 159, and has a residue other than methionine at position 29, a residue other than lysine at position 54, a residue other than serine at position 83, a residue other than asparagine at position 92, a residue other than leucine at position 222, a residue other than alanine at position 272, or a residue other than threonine at position 328, a pharmaceutically acceptable carrier, and an adjuvant.

20. The composition of claim 19 wherein the HA or the HA-1 has at least 99% amino acid sequence identity to SEQ ID NO:1.

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