



US 20180028643A1

(19) **United States**

(12) **Patent Application Publication**

Kingstad-Bakke et al.

(10) **Pub. No.: US 2018/0028643 A1**

(43) **Pub. Date: Feb. 1, 2018**

(54) **ZIKA VIRUS VACCINES USING VIRUS-LIKE PARTICLES**

Publication Classification

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(51) **Int. Cl.**

A61K 39/12 (2006.01)

C12N 7/00 (2006.01)

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(52) **U.S. Cl.**

CPC **A61K 39/12** (2013.01); **C12N 7/00**
(2013.01); **A61K 2039/55** (2013.01)

(21) Appl. No.: **15/629,503**

(22) Filed: **Jun. 21, 2017**

(57)

ABSTRACT

Related U.S. Application Data

(60) Provisional application No. 62/352,904, filed on Jun. 21, 2016, provisional application No. 62/384,967, filed on Sep. 8, 2016.

A flavivirus virus-like particle and methods of making and using that particle, and antibodies raised to a plurality of those particles, are provided.

FIG. 1A.

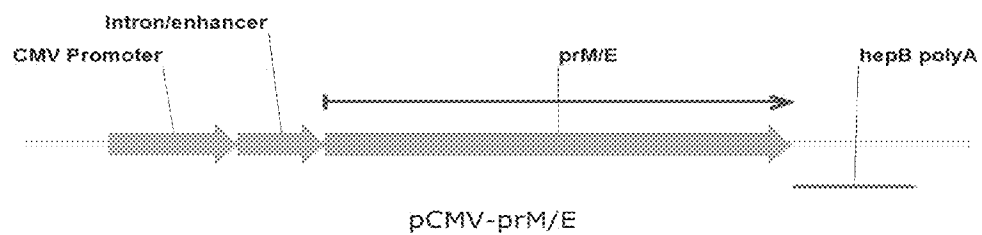


FIG. 1B.

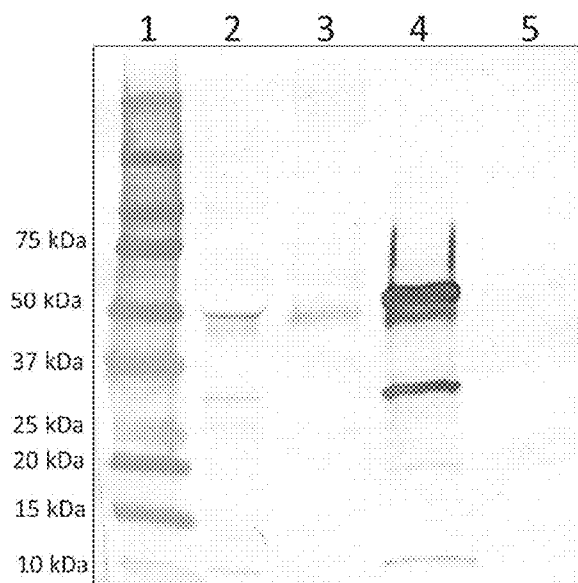


FIG. 1C.

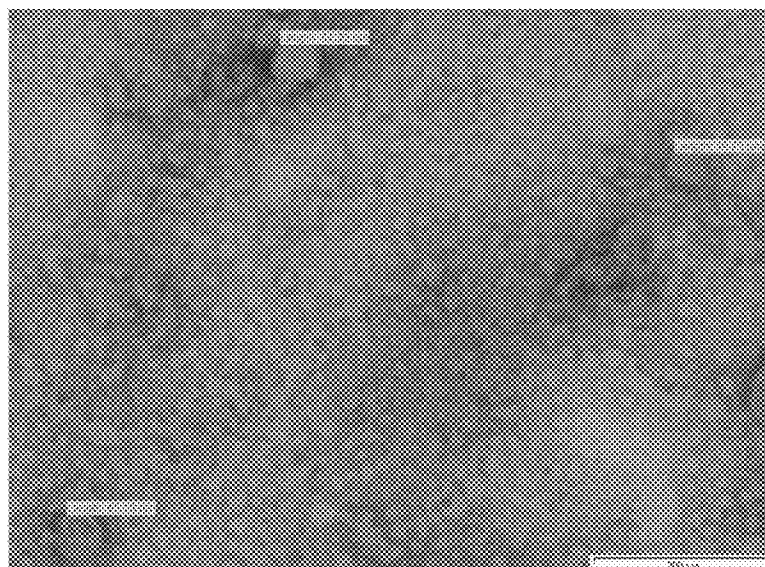


FIG. 1D.

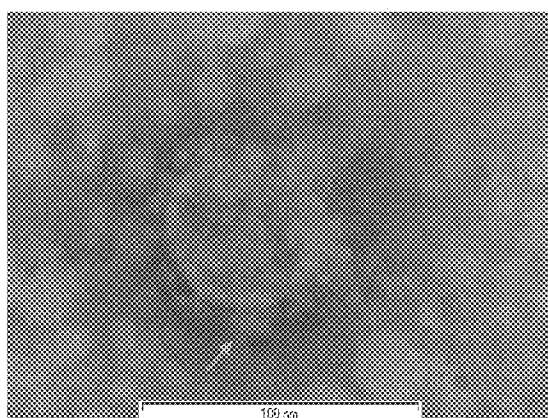


FIG. 1E.

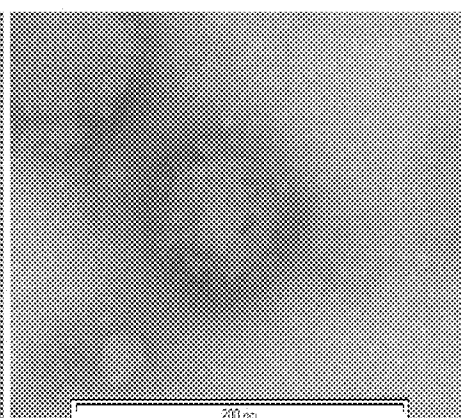


FIG. 2A.

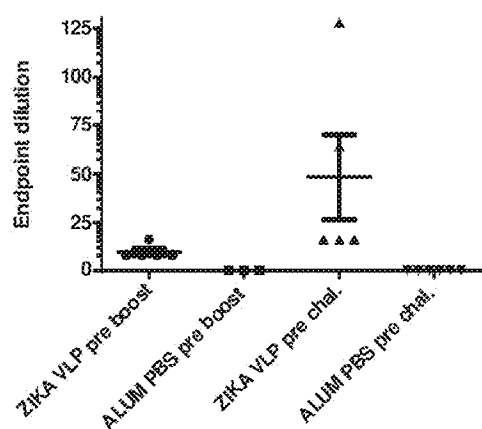


FIG. 2B.

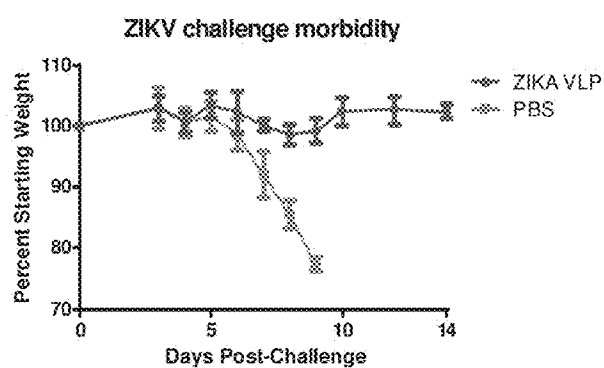


FIG. 2C.

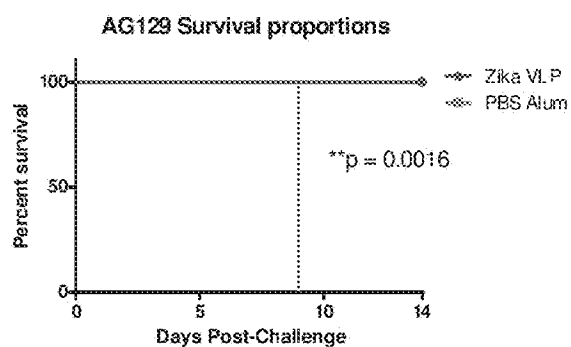


FIG. 2D.

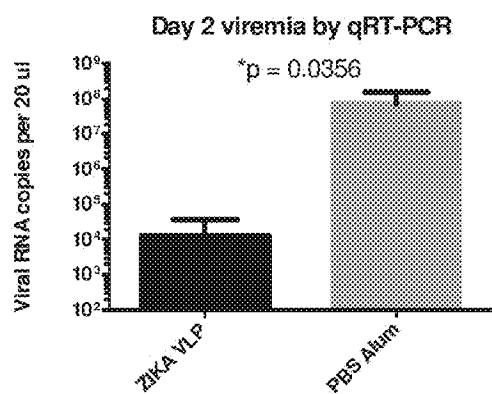


FIG. 2E.

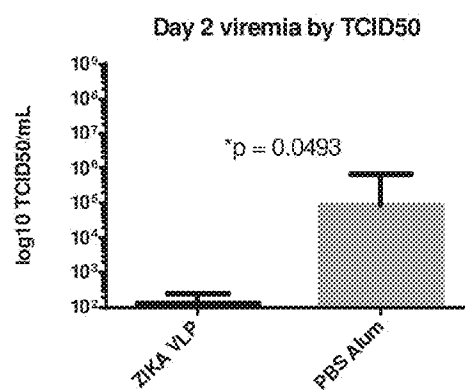


FIG 2F.

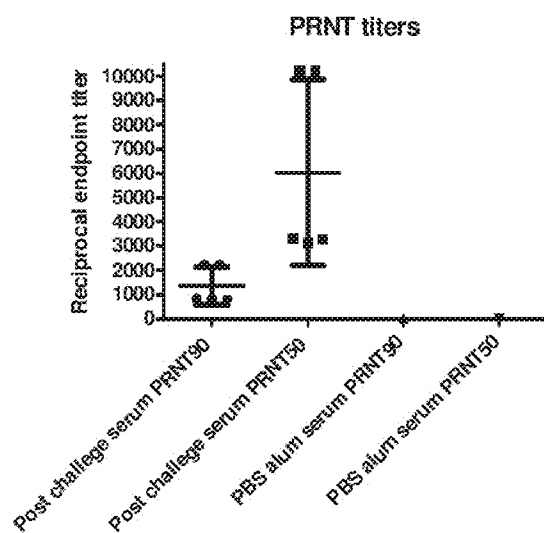


FIG. 3A.

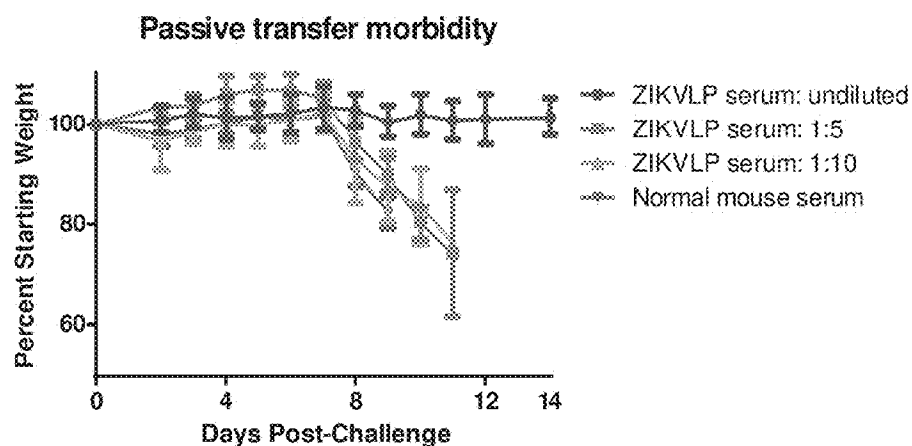


FIG. 3B.

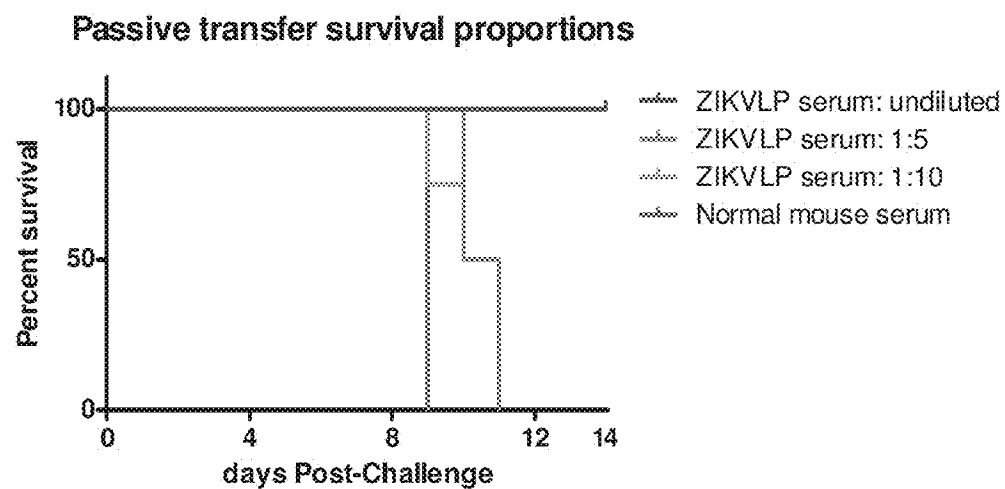


FIG. 4

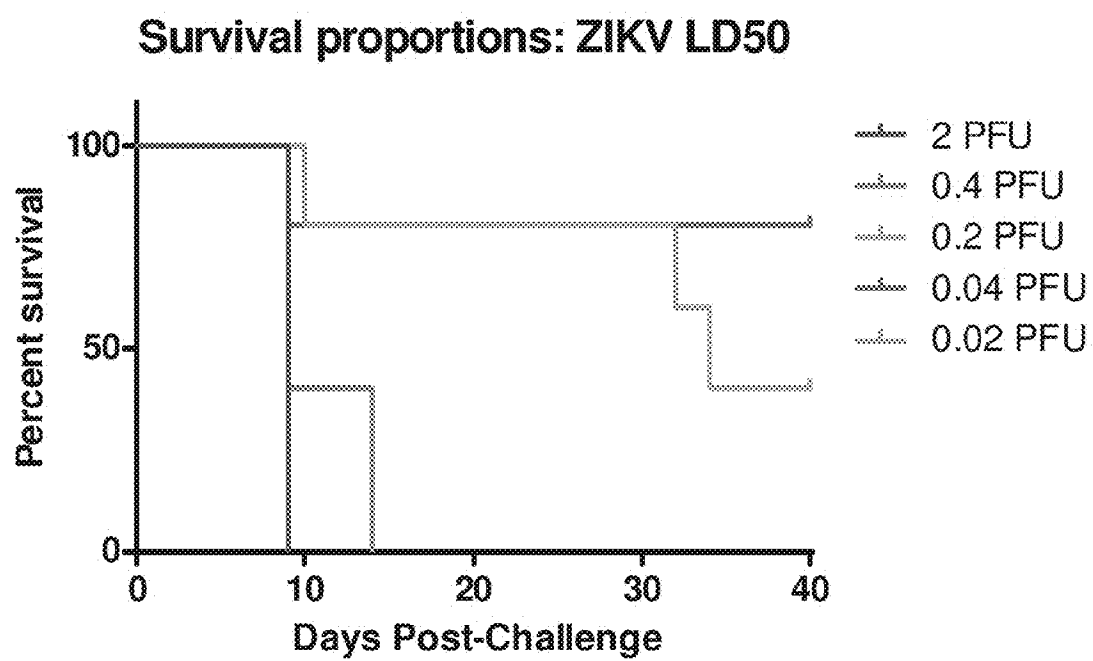


FIG. 5A

WEIGHT LOSS OF AG129 AFTER ID CHALLENGE WITH 20 PFU ZIKV OVER A 12 DAY PERIOD.
Passive transfer morbidity

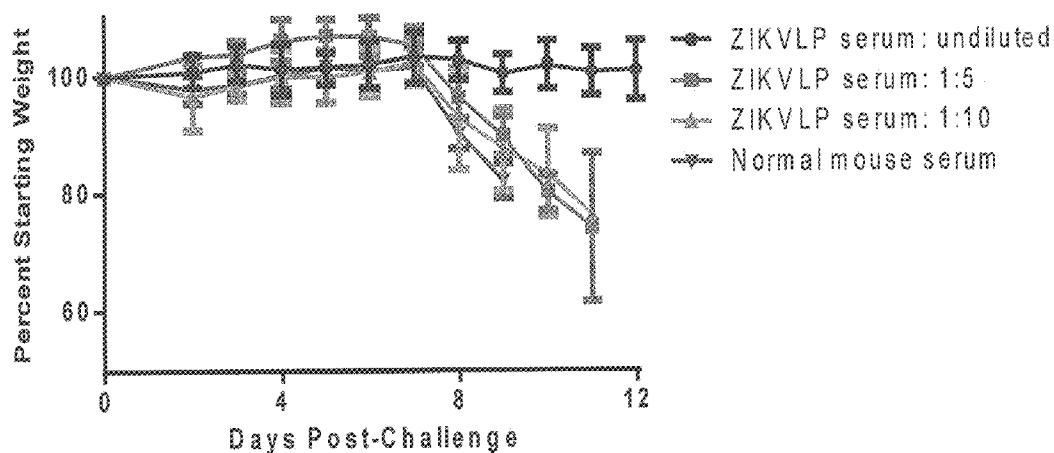
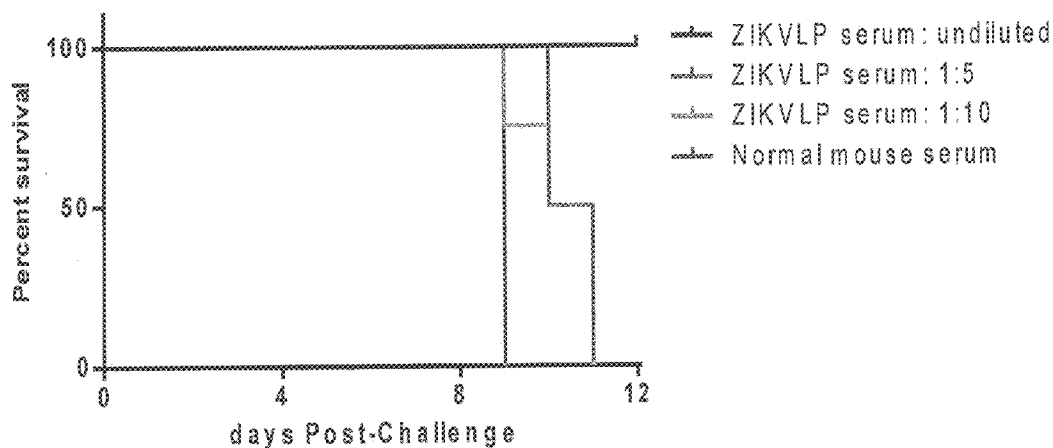


FIG. 5B

SURVIVAL OF AG129 AFTER ID CHALLENGE WITH 200 PFU ZIKV OVER A 12 DAY PERIOD.
Passive transfer survival proportions



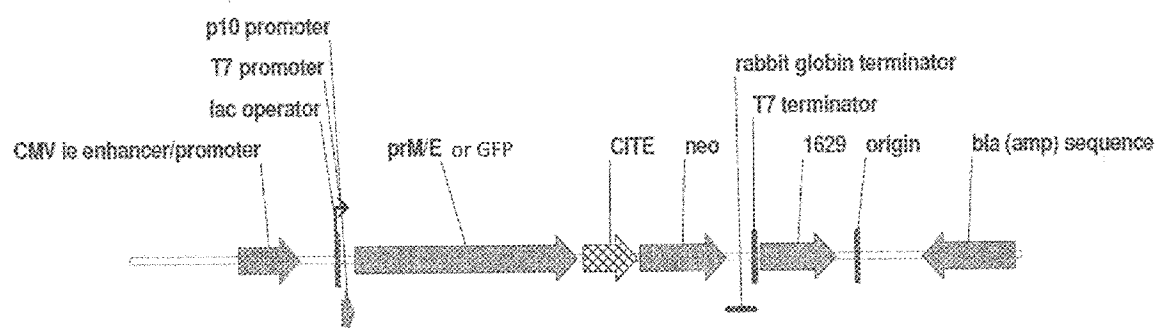
101	1	TGTTGATACT	CATACTCTTC	CTTTTTCATA	TTATTGAAGC	ATTTATCAGG	GTTATTGTCT	CATGAGCG	
		ACAACATATGA	GTATGAGAAG	GAAAAAGTAT	AATAACTTCG	TAAATAGTCC	CAATAACAGA	GTACTCGG	
101		ANAANTNNNG	GTTCOCOCGN	CATTNCCCCG	AAAAGTGCCA	CCTGACGTCG	NCGGATCGGG	AGATCTCG	
		TNTTNANNCC	CAAGGGGCGN	GTAANGGGGC	TTTTACGGGT	GGACTSCAGC	NGCCTIAGCC	TCTAGAGC	
201		GCTCTGATGC	CGCATAGTAA	AGCCAGTATC	TGCTCCCTGC	TTGTGTGTTC	GAGGTGCGTG	AGTAGTGC	
		CGAGACTACG	GCGTATCAAT	TCGGTCATAG	ACGAGGGACG	AACACACAAC	CTCCAGCGAC	TCATCACG	
301		CTTGACCGAC	AATTGCATGA	AGAATCTGCT	TAGGGTTAGG	CGTTTIGGGC	TGCTTCGCGA	TGTACGGC	
		GAACTGGCTG	TTAACGTACT	TCTTAGACCA	ATCCCAATCC	GCAAAACGCG	ACGAAGCGGT	ACATGCCG	
401		AGTTATTAAT	AGTAATCAAT	TACGGGGTCA	TTAGTTCATA	GCCCATATAT	GGAGTTCGGC	GTTACATG	
		TCATAATTA	TCATTAGTAA	ATGCCCCAGT	AATCAAGTAT	CGGGTATATA	CCTCAAGGCG	CAATGTAT	
501		CCAACGACCC	CCGCCCATTC	ACGTCAATAA	TGACGTATGT	TCCCATAGTA	ACGCCAATAG	GGACTTTT	
		GSTTGTCTGG	GGCCGGTTAC	TGCAGTTATT	ACTGCATACA	AGGGTATCAT	TGCGGTTATC	CCTGAACG	
601		AACTGCCCCA	TTGGCAGTAC	ATCAAGTGTA	TCATATGCCA	AGTCCGCCCC	CTATTGACGT	CAATGACG	
		TTGACGGGTG	AACCGTCATG	TAGTTTACAT	AGTATACGGT	TCAGGCGGGG	GATAACTSCA	GTTACTGT	
701		ATGACCTTAC	GGGACTTTCC	TACTTGGCAG	TACATCTACG	TATTAGTCAAT	CGCTATTACC	ATGGTGAT	
		TACTGGAATG	CCCTGAAAGG	ATGAACCGTC	ATGTAGATGC	ATAATCAGTA	GCGATAATGG	TACCACTT	
801		AGCGGTTTGA	CTCACGGGGA	TTTCCAAGTC	TCCACCCCAT	TGACGTCAAT	GGGAGTTTGT	TTTGGGCA	
		TCGCCAAACT	GAGTGCCCTT	AAAGGTTTCA	AGGTGGGGTA	ACTGCAGTTA	CCCTCAACA	AAACCGTC	
901		TAACCCCGCC	CCGTGACGCG	AAATGGGGCG	TAGGCGTGTA	CGGTGGGAGG	TCTATATAAG	CAGAGCTT	
		ATTGGGGCGG	GGCAACTGGC	TTTACCCGCC	ATCCGCACAT	GCCACCTCC	AGATATATTC	GTCTCGAA	
1001		CATCCACGCT	GTTTGAACCT	CCATAGAAGA	CACCGGGACC	GATCCAGCCT	CCGGGCGCGG	GAACGGTG	
		GTAGTGCGGA	CAAACTGGA	GGTATCTTCT	CTGSCCCTGG	CTAGGTGCGA	GGCGCGCGCC	CTTGCCCA	
1101		ACTCACCGTC	CGGATCTCAG	CAAGCAGGTA	TGTACTCTCC	AGGGTGGGCC	TGGCTTCCCC	AGTCAAGG	
		TGAGTGGCAG	GCCTAGAGTC	GTTGCTCCAT	ACATGAGAGG	TCCCACCCGG	ACCGAAGGGG	TCAGTTCT	
1201		CTCTTACATG	TACCTTTTGC	TTGCCTCAAC	CCTGACTATC	TTCCAGGTCA	GGATCCCAGA	GTGAGGGG	
		GAGAATGTAC	ATGGAAAACG	AACGGAGTTG	GGACTGATAG	AAGGTCCAGT	CCTAGGGTCT	CAGTCCCC	
1301		GAACAGTAAA	CCCTGCTCCG	AATATTGCTT	CTCACATCTC	GTCAATCTCC	GCGAGGACTG	GGGACCCC	
		CTTGTCATTT	GGGACGAGGC	TTATAACGGA	GAGTGTAGAG	CAGTTAGAGG	CGCTCCTGAC	CCCTGGGT	
1401		CTCTCTGCTG	ACCACAGCTA	TGGCAGCGGA	GGTCACTAGA	CGTGGGAGTG	CATACTATAT	GTACTTGT	
		GGAGGACGAC	TGGTGTGCTT	ACCGTCGCCCT	CCAGTGATCT	GCACCCCTAC	GTATGATATA	CATGAACG	
1501		CCAACCACAT	TGGGGATGAA	TAACTGTTAT	ATACAGATCA	TGGATCTTGG	ACACATGTTG	GATGCCAC	
		GGTTGGTGTA	ACCCCTACTT	ATTACAAATA	TATGTCTAGT	ACCTAGAACC	TGTGTACACA	CTACGGTG	
1601		GGGTGGAACC	AGATGACGTC	GATTGTTGGT	GCAACACGAC	GTCAACTTGG	GTTGTGTACG	GAACCTGT	
		CCCACCTTGG	TCTACTGCAG	CTAACCAACCA	CGTTGTGCTG	CAGTTGAACC	CAACACATGC	CTTGCACG	
1701		AAGAGCTGTG	ACGCTCCCTT	CCCATTCCAC	TAGGAAAGCTG	CAAACGCGGT	CGCAAACCTG	GTTGGAAAT	
		TTCTCGACAC	TGGAGGGGGA	GGGTAAGGTG	ATCCTTCGAC	GTTTGGGCCA	GCCTTGGGAC	CAACCTTT	
1801		GAAAATTGGA	TATTAGGAA	CCCTGGCTTC	GCGTTAGCAG	CAGCTGCCAT	CGCTTGGCTT	TTGGGAAAT	
		CTTTTAACTT	ATAAGTCTTT	GGCACCGAAG	CGCAATCGTC	GTCCGACGGTA	GCGAACCGAA	AACCTTTT	
1901		TGATACTGCT	GATTGCCCGG	GCATACAGCA	TCAGGTGCTAT	AGGAGTCAGC	AATAGGGACT	TTGTGGAA	
		ACTATGACGA	CTAACGGGGC	CCTATGTCTT	AGTCCACGTA	TCCTCACTCG	TTATCCCTGA	AACACCTT	
2001		CTTGGAACAT	GGAGGTTGTG	TCACCGTAAT	GGCACAGGAC	AAACCGACTG	TGCACATAGA	GCTGGTTT	
		GAACCTTGTG	CCTCCAAACAC	AGTGGCATTA	CCGTGTCTCT	TTTGGCTGAC	AGCTGTATCT	CGACCAAT	
2101		TCCTACTGCT	ATGAGGCATC	AATATCGGAC	ATGGCTTCGG	ACAGCCGCTG	CCCAACACAA	GGTGAAGG	
		AGGATGACGA	TACTCCGTAG	TTATAGCCTG	TACCGAAGCC	TGTCCGGGAC	GGGTTGTGTT	CCACTTCC	
2201		TCTGCAAAAG	AACGTTAGTG	GACAGAGGCT	GGGGAATGCG	ATGTGGACIT	TTTGGCAAAAG	GGAGCCTT	
		AGACGTTTTT	TTGCAATCAC	CTGTCTCCGA	CCCCTTTACC	TACACCTGAA	AAACCGTTTC	CCTCGGAC	

FIG. 6A

2301	AATGACCGGG AAGAGCATCC AGCCACAGAA TCTGGAGTAC CGGATAATGC TGTCACTTCA TGGCTCCG
	TTACTGGCCC TTCTCGTAGG TCGGTCTCTT AGACCTCATG GCCTATTACG ACAGTCAAGT ACCGAGGC
2401	CATGAAACTG ATCAGAATAG AGCGAAGSIT GAGATAACGC CCAATTCACC AAGAGCCGAA GCCACCCG
	GTACTTTGAC TACTCTTATC TCGCTTCCAA CTCTATTGGG GGTAAAGTGG TTCTCGGCTT CGGTGGG
2501	AACCGAGGAC AGGCCTTGAC TTTTCAGAIT TGTATTACTT GACTATGAAT AACAAGCACT GGTGGGT
	TTGGCTCCTG TCCGGAACCTG AAAAGTCTAA ACATAATGAA CTGATACTTA TTGTTCTGTA CCAACCA
2601	TTGGCAGGCT GGGGCAGACA CCGGAACCTC ACACTGGAAC AACAAAGAAG CACTGGTAGA GTTCAAC
	AACCGTCCGA CCCCCTCTGT GGCCTTGAGG TGTACCTTG TTGTTTCTTC GTGAACATCT CAAGTTC
2701	CTAGGGAGTC AAGAAGGAGC AGTTCACACG CCCCCTTGCTG GAGCTCTGGA GGCTGAGATG GATGGTGC
	GATCCCTCAG TTCTTCTCTG TCAAGTGTGC CGGGAACGAC CTCGAGACCT CCSACTCTAC CTACCAC
2801	GTCCGCTGAA AATGGATAAA CTTAGAITGA AGGGCGTGTG ATACTCCTTG TGTACCGCAG CGTTCAC
	CAGCGGACTT TTACCTATTT GAATCTAACT TCCCGCACAG TATGAGGAAC ACATGGCGTC GCAAGTGT
2901	GACAGTCACA GTGAGGTAC AGTACCCAGG GACAGATGGA CCTTGCAAGG TTCCAGCTCA GATGGCG
	CTGTCACTGT CACCTCCATG TCATGCGTCC CTGTCTACCT GGAACGTTCC AAGGTCTGAT CTACCGC
3001	TTGATAACCG CTAACCCCGT AATCACTGAA AGCACTGAGA ACTCTAAGAT GATGCTGGAA CTTGATC
	AACATITGGC GATTGGGGCA TTAGTCACTT TCGTGACTCT TGAGATTCTA CTACGACCTT GAACTAG
3101	TCGGGGAGAA GAAGATCACC CACCACTGCC ACAGGAGTGG CAGCACCATT GGAAAAGCAT TTGAAGC
	AGCCCCCTCTT CTCTAGTGG GTGGTGACCG TGTCTCACC GTCGTGGTAA CCTTTTCGTA AACTTCG
3201	GGGAGACACA GCCTGGGACT TTGGATCAGT TGGAGGGGCT CTCAACTCAT TGGGCAAGGG CATCCAT
	CCCTCTGTGT CGGACCCTGA AACCTAGTCA ACCTCCCGA GAGTTGAGTA ACCCGTTCCC GTAGGTAC
3301	GGAGGAATGT CCTGGTTCTC ACAAATTCTC ATTGGAACGT TGCTGATGTG GTTGGGTCTG AACACAA
	CCTCCTTACA GGACCAAGAG TGTTTAAGAG TAACCTTGCA ACCACTACAC CAACCCAGAC TTGTGTT
3401	TAGGGGGAGT GTTGATCTTC TTATCCACAG CTGTCTCTGC TGATGTGGGG TGCTCGGTGT GAGGATCT
	ATCCCCCTCA CAACTAGAAG AATAGGTGTC GACAGAGACG ACTACACCCC ACAGGCCACA CTCTAG
3501	CCCTAAACTT CATGGGTTAC GTAATTGGAA GTTGGGGGAC ATTCCACAA GATCATATIG TACAAAAC
	GGGATTGAA GTACCCCAATG CATTAACTT CAACCCCTG TAACGGTGT CTAGTATAAC ATGTTTTT
3601	CAGGCCATTT GATTGGAAG TATGTCAAAG GATTGTGGGT CTTTTGGGT TTGCTGCTCC ATTTACAC
	GTCCGGATAA CTAACCTTTC ATACAGTTTC CTAACACCCA GAAAACCCGA AACGACGAGG TAAATGT
3701	GCATGTATAC AAGCTAAACA GGCTTTCACT TTCTCGCCAA CTTACAAGGC CTTCTAAGT AAACAGT
	CGTACATATG TTGGATTGT CCGAAAGTGA AAGAGCGGTT GAATGTTCCG GAAAGATTCA TTGTCTAC
3801	CTGGTCTGTG CCAAGTGTIT GCTGACGCAA CCCCCACTGG CTGGGGCTTG GCCATAGGCC ATCAGCG
	GACCAGACAC GGTTCACAAA CGACTGCCGT GGGCGTGACC GACCCCGAAC CGGTATCCGG TAGTCGG
3901	CCATACTGGG GAACTCCTAG CCGCTTGTIT TGCTCGCAGC CGGTCTGGAG CAAAGCTCAT AGGAAC
	GGTATGACGC CTTGAGGATC GCGCAACAAA ACGAGCGTGG GCCAGACCTC GTTTCGAGTA TCCTTGA
4001	TCGTTTCGAT CTACGTATGA TCTTTTTCCC TCTGCCAAAA ATTATGGGGA CATCATGAAG CCGCTTG
	AGCAAAAGCTA GATGCATACT AGAAAAAGGG AGACGGTTTT TAATACCCCT GTAGTACTTC GGGGAAC
	EcoRI
4101	TTTTCATTCG AATAGTGTGT TGAATTTTT TGTGTCTCTC ACTCGGAAGG AATTCTGCAT TAATGAAC
	AAAAGTAACG TTATCACACA ACCTTAAAAA ACACAGAGAG TGAGCCTTCC TTAAGACGTA ATTACTT
4201	TTGGGGGCTC TTCCGGTTCC TCGCTCACTG ANCTCGNTGC GCTTCGGTGC T
	AACCCGCGAG AAGGCGAAGG AGCGAGTGAC TNGAGCNACG CGAAGCCAGC A

FIG. 6B

pTriex4-neo Expression cassette



- CITE: IRES
- 1629: Poly A site
- Fairly "run of the mill" expression vector

FIG. 7

pTriex4-neo GFP expression in HEK-293

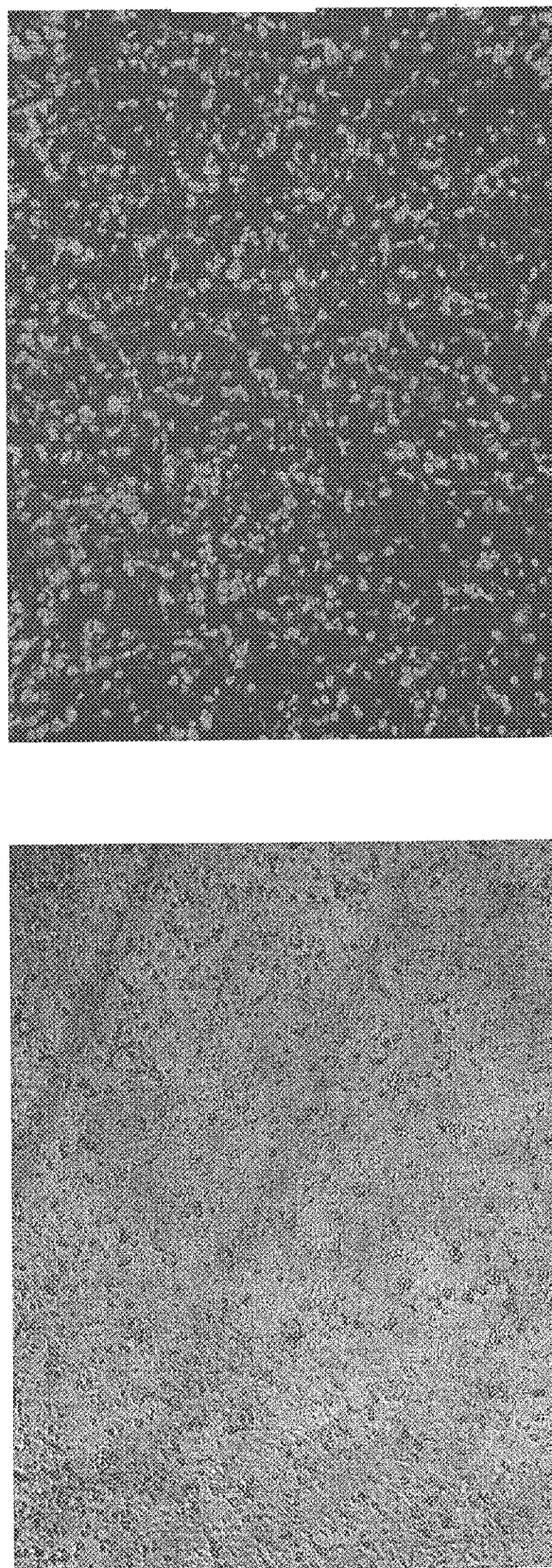


FIG. 8A

pCMV GFP expression in HEK-293

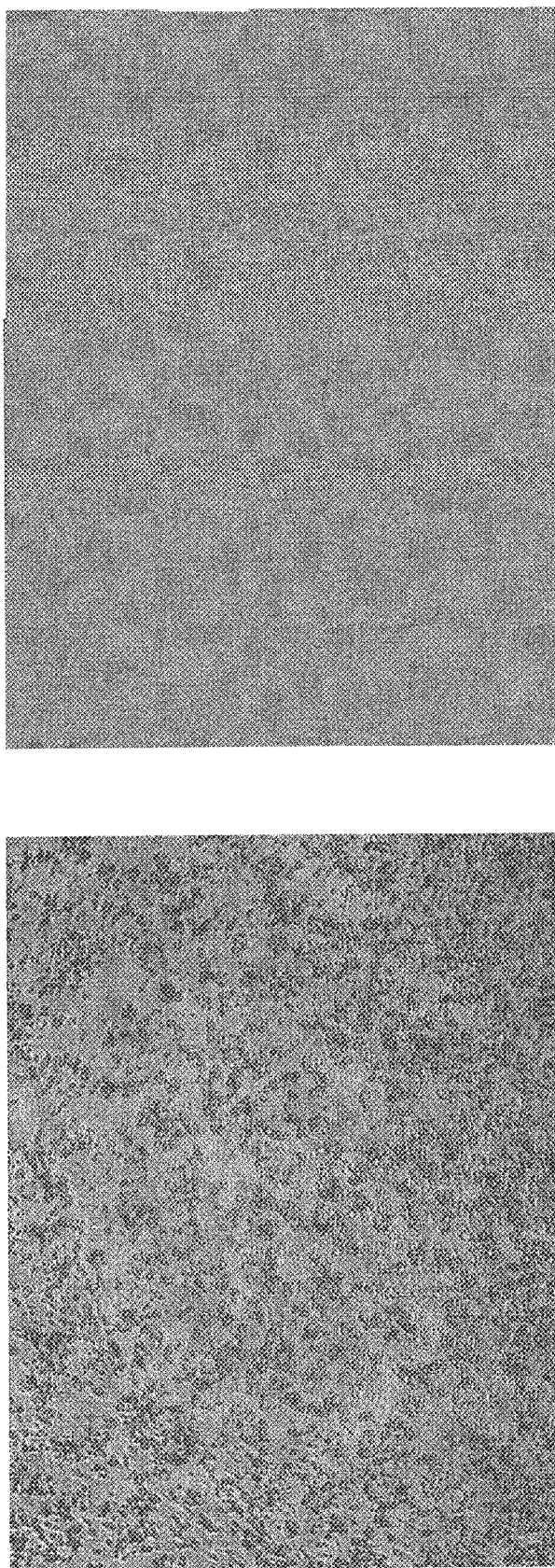
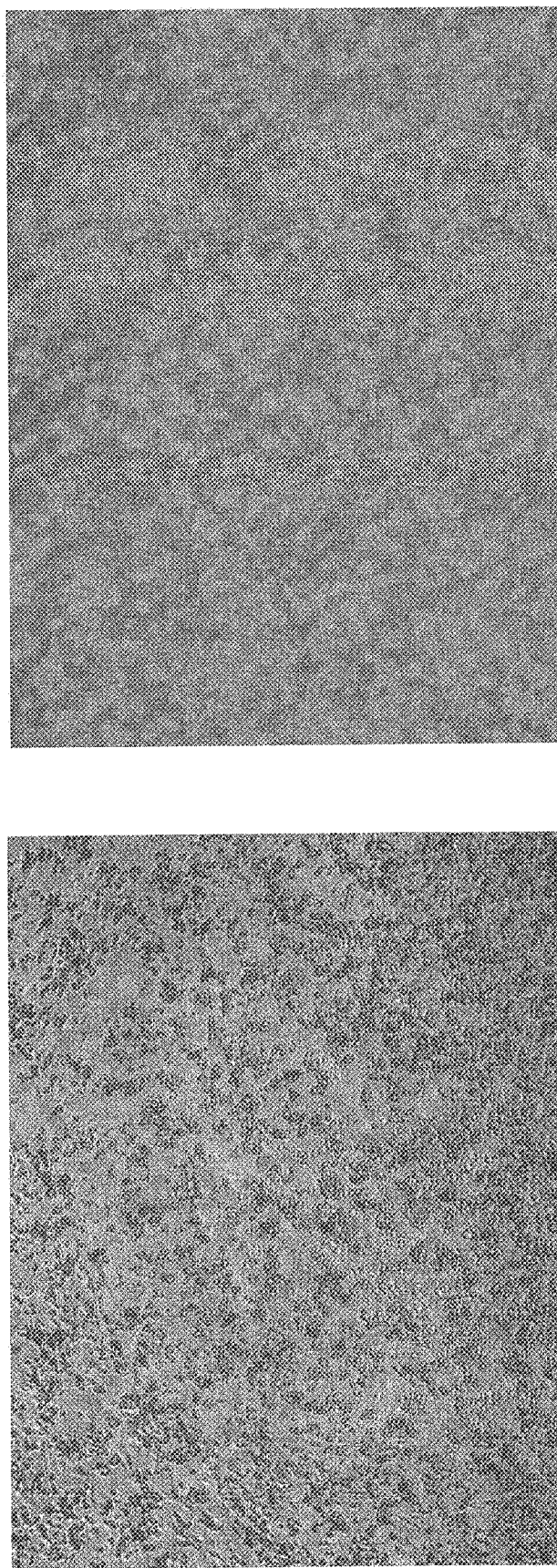


FIG. 8B

pCMV GFP expression in HEK-293



- Lamp turned down about 70%

FIG. 8C

pTriex vs pCMV prM/E expression

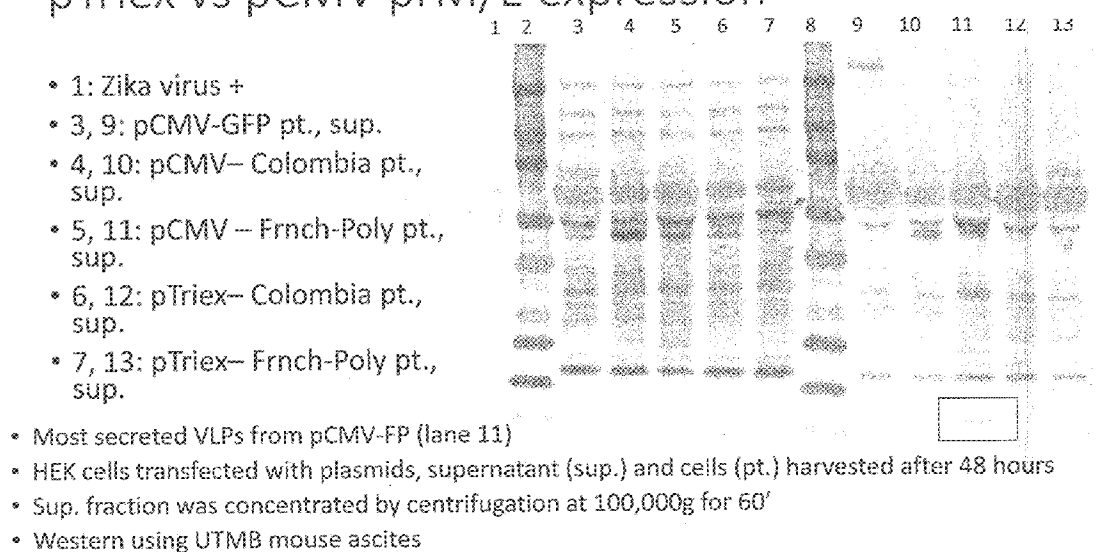


FIG. 9

Preliminary studies in mice

- Mice were injected IP with $\sim 10^6$ TCID₅₀ of ZIKV
- 5 weeks later bleed, then injected with crude VLP supernatant
- Mice were bled 7 days after injection and antibodies analyzed by ZIKV ELISA

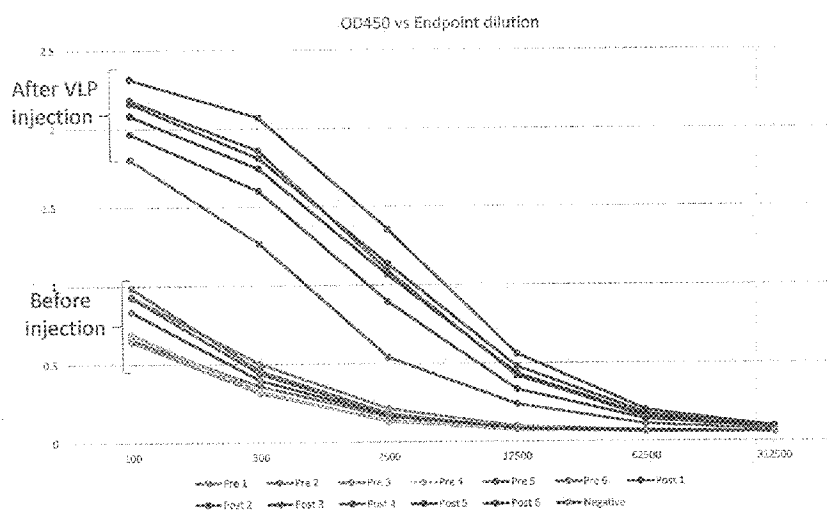
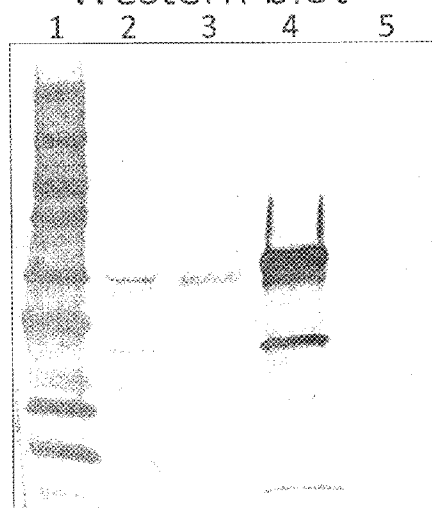


FIG. 10

Zika VLP purification: post sucrose purification Western blot



- 1: Marker
- 2: VLP 100,000g precipitation (previous slide)
- 3: Zika virus +
- 4: pCMV – Frnch-Poly post sucrose purification
- 5: pCMV-GFP post sucrose purification

Supernatant from T-75 flasks transfected with pCMV-prM/E, or pCMV-GFP were collected after 3 days, clarified by centrifugation (15,000g, 30'), then layered onto a 20% sucrose cushion, pelleted at 112,000g for 3.5hr.

FIG. 11

Sucrose fractional analysis

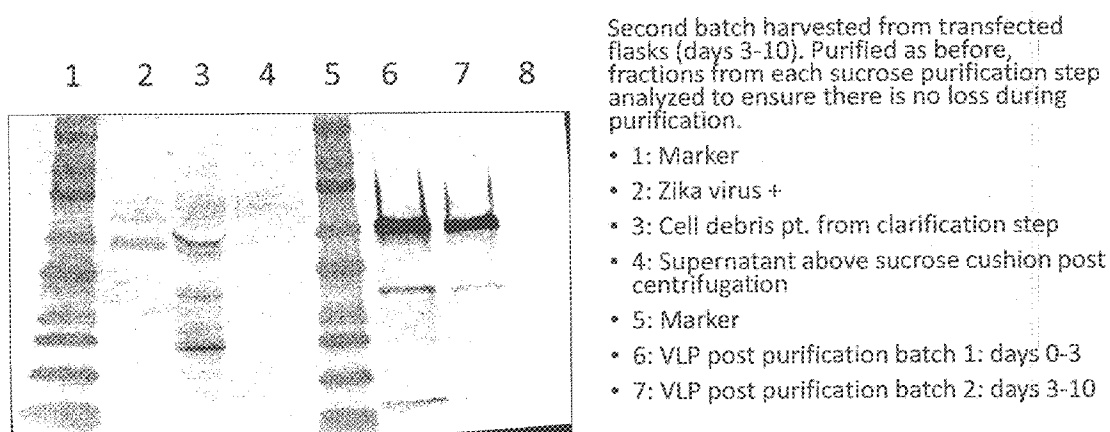


FIG. 12

pCMV expressed much more protein than pTriex

- pCMV-Col. and all pTriex constructs didn't express significant levels of VLPs

more protein from the second batch of VLPs (days 3-10)
about 60ug total from about 100ml

productivity of the cells was highest in the first three days,
yielding ~50ug per 15ml = 3.3ug per ml, or 3.3 mg/liter

clone Zeocin resistance gene into pCMV vector for stable cell
line

- Generated cell line with pTriex, but terminated after western data

FIG. 13

Mouse study

- Mice: 11 AG129 mixed sex, age
- Route of vaccinations: IM
- Adjuvant: 1mg Alum
- Route of challenge: ID footpad
- Challenge Dose: 1×10^2 PFU

Group	Constituent	IM	Sex	Age (weeks)	Dose	N
1	VLP+ adjuvant	IM	F (n=2) M (n=3)	12 (n=5)	2 μ g total protein	5
2	Adjuvant	IM	F (n=2) M (n=3)	12 (n=3) 9 (n=3)	N/A	6

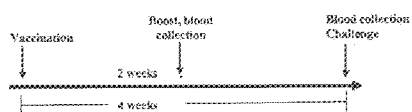
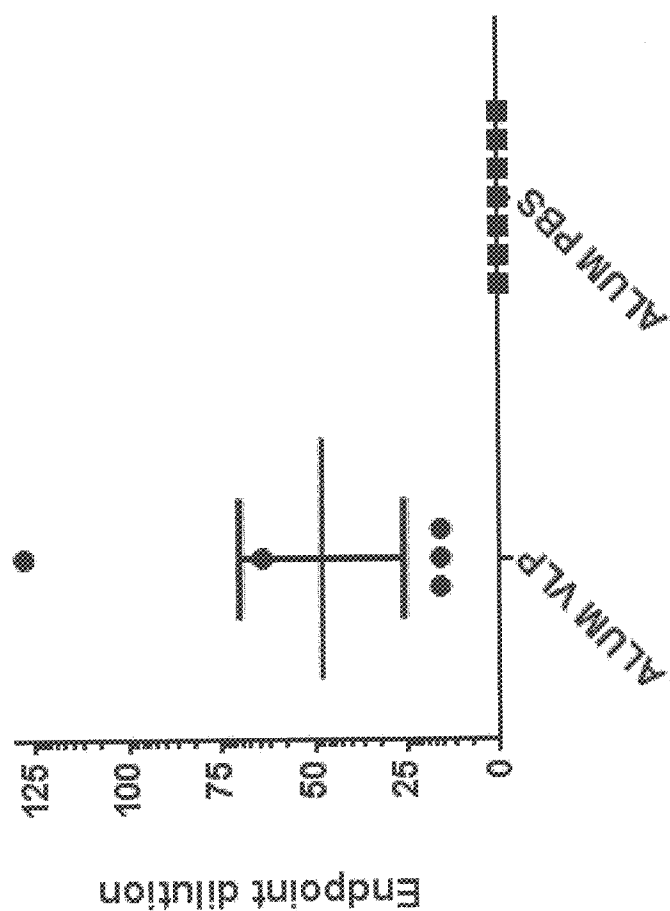


FIG. 14

Antibody two weeks post boost

- Normal for alum/protein vaccine



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Survival and morbidity

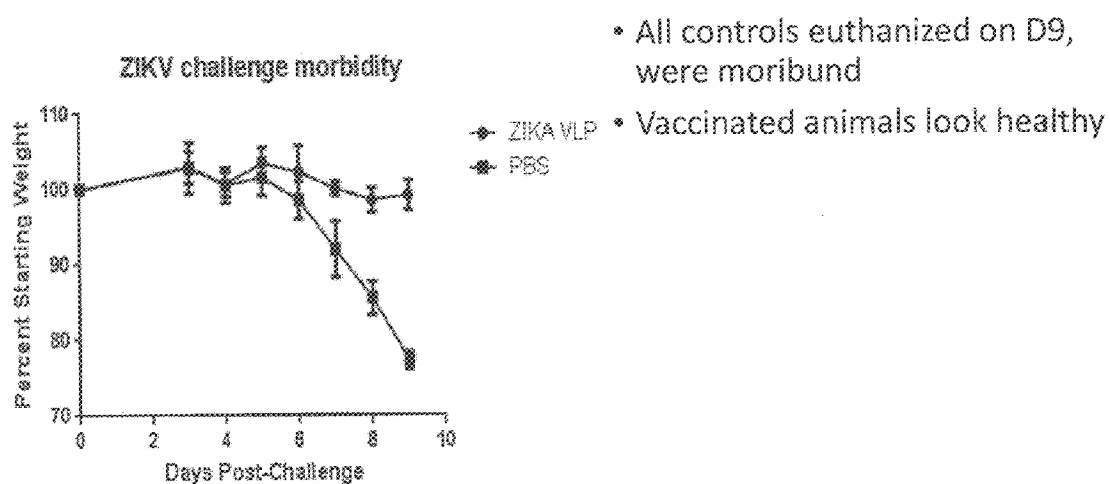


FIG. 16

FIG. 17A.

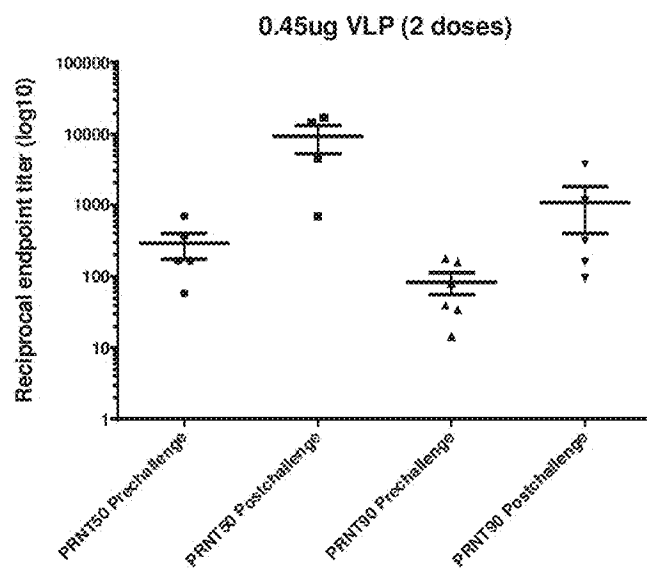


FIG. 17B.

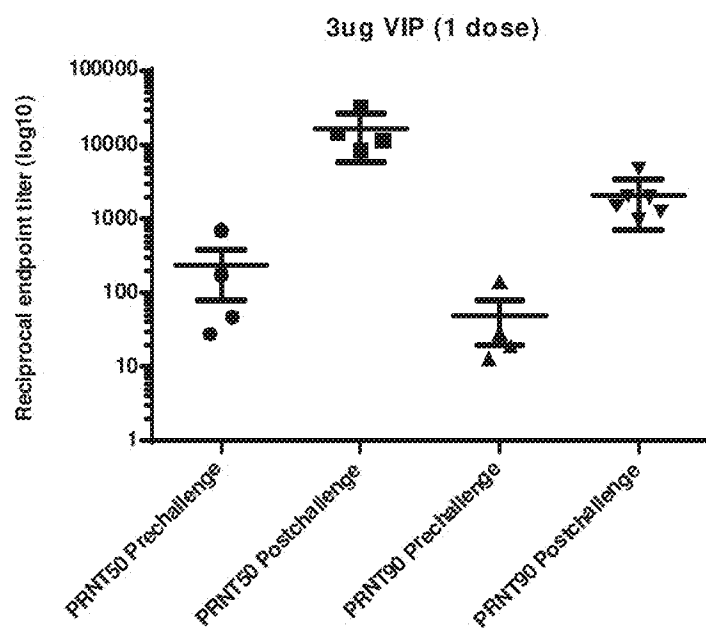


FIG. 17C

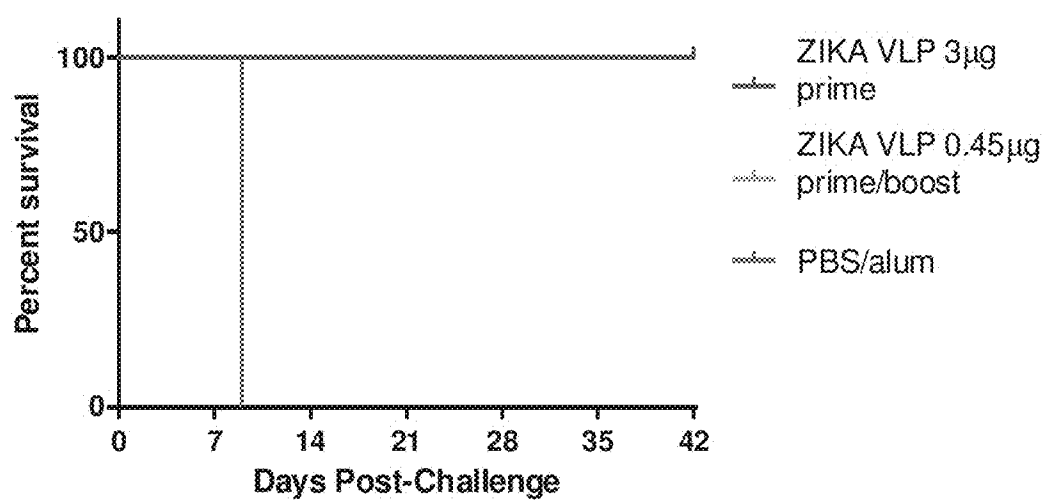


FIG. 18A.

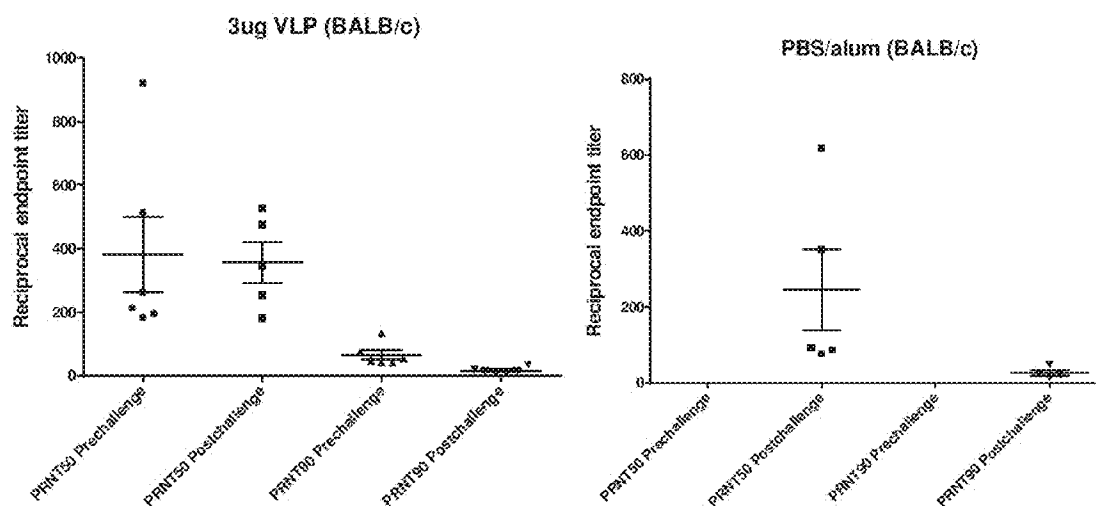


FIG. 18B.

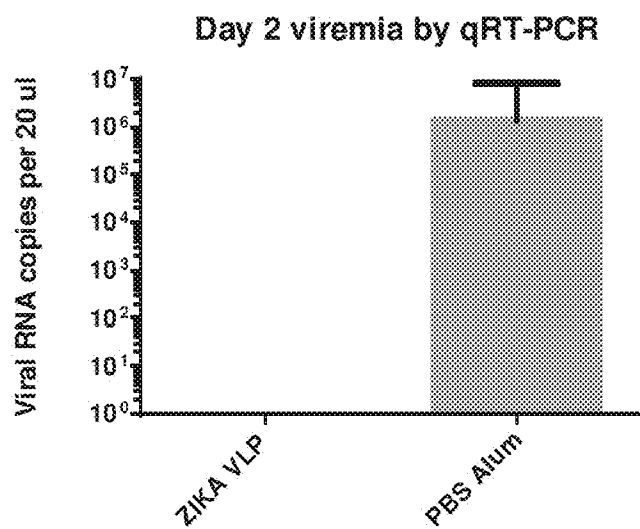
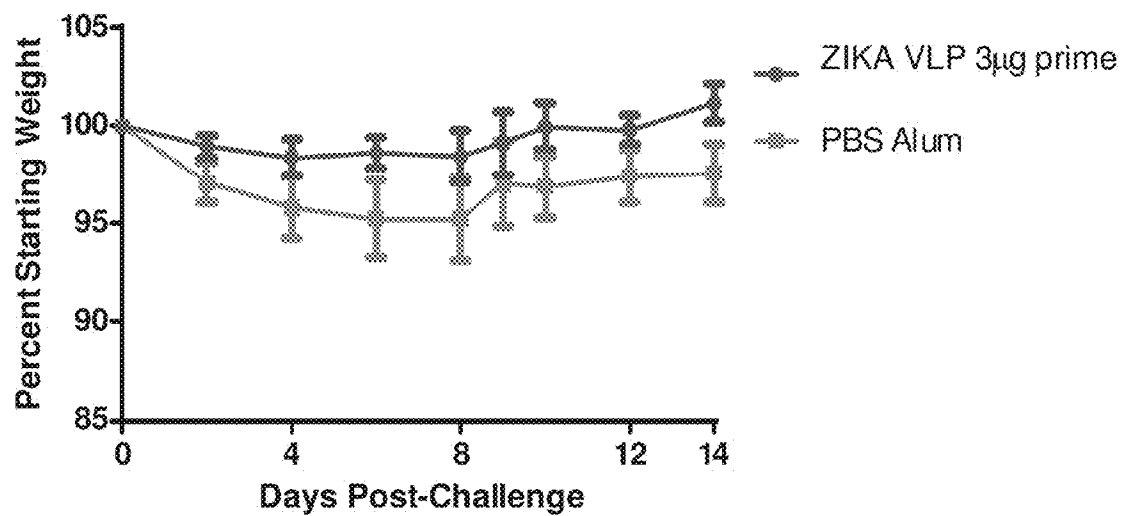


FIG. 18C.



ZIKA VIRUS VACCINES USING VIRUS-LIKE PARTICLES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the benefit of the filing date of U.S. application Ser. No. 62/352,904, filed on Jun. 21, 2016, and U.S. application Ser. No. 62/384,967, filed on Sep. 8, 2016, the disclosure of which are incorporated by reference herein.

BACKGROUND

[0002] Zika virus (ZIKV; Flaviviridae, Flavivirus) is an emerging arbovirus, transmitted by *Aedes* mosquitoes (loos et al., 2014). ZIKV has a positive-sense, single-stranded RNA genome, approximately 11 kilobases in length that encodes three structural proteins: the capsid (C), premembrane/membrane (prM), and envelope (E), and seven non-structural proteins (NS1, NS2A, NS2B, NS3, NS4A, 2K, NS4B, and NS5). Based on a genetic study using nucleotide sequences derived from the NS5 gene, there are three ZIKV lineages: East African, West African, and Asian (Musso, 2015; Faye et al., 2014). ZIKV emerged out of Africa and previously caused outbreaks of febrile disease in the Yap islands of the Federated states of Micronesia (Duffy et al., 2009), French Polynesia (Cao-Lormeau et al., 2014), and Oceania. Currently, several Latin American countries are experiencing the first-ever reported local transmission of ZIKV in the Americas (Hennessey et al., 2016). The current outbreak in the Americas is cause for great concern, because of the fast and uncontrolled autochthonous spread. Clinically, infection with ZIKV resembles dengue fever and several other arboviral diseases (Dyer, 2015), but it has been linked to neurological syndromes and congenital malformation (Pinto Junior et al., 2015). Alarming, the rate of microcephaly (small head, reduced brain size, impaired neurocognitive development) in infants born to pregnant women has increased significantly (20-fold in 2015) in areas with high ZIKV incidence in Brazil (Oliveira Melo et al., 2016) (Butler, 2016). In February 2016, the World Health Organization declared the Zika virus an international public health emergency, prompted by its link to microcephaly. As many as four million people could be infected by the end of the year (Galland, 2016).

[0003] To date, there are no vaccines or antiviral therapy for ZIKV, although successful vaccines have been developed for other flavivirus infections (dengue, Japanese encephalitis and yellow fever).

SUMMARY

[0004] Mosquito-borne Zika virus (ZIKV) typically causes a mild and self-limiting illness known as Zika fever, which often is accompanied by maculopapular rash, headache, and myalgia. However, more serious consequences have been reported for ZIKV infection during pregnancy, microcephaly of the fetus. As described herein, Zika virus-like particles (VLPs) were developed and their immunogenicity and protective efficacy were evaluated in a small animal model for wild-type ZIKV. The prM and E genes of ZIKV strain 33 H/PF/2013 with a nascent signal sequence in the 3' coding region of the capsid protein were cloned into a pCMV expression vector under the control of a cytomegalovirus (CMV) promoter and CMV polyadenylation signal.

Following transfection of HEK293 cells, ZIKV-VLPs expression was confirmed by Western blot and transmission electron microscopy. ZIKV-VLPs (about 0.45 μ g) were formulated with 0.2% Imject alum and used to inject groups of six-week-old AG129 mice by the intramuscular (IM) route, followed by a boost administration two weeks later. Control groups received PBS mixed with alum. At five weeks post-initial vaccination all animals were challenged with 200 PFU (>400 LD50s) of ZIKV strain H/PF/2013 by injection into the right hind footpad. All control animals (n=6) died 9 days post challenge, while vaccinated mice survived with no morbidity or weight loss and had significantly lower viremia. This was in contrast to Dengue VLPs produced from prM and E, which did not produce a protective immune response (Pijlman, 2015). Significant levels of neutralizing antibodies were observed in all ZIKV-VLP vaccinated mice compared to control groups. The role of neutralizing antibodies in protecting mice was demonstrated by antibody passive transfer studies; naive AG129 mice that received pooled serum from VLP vaccinated animals were fully protected. Thus, the present findings demonstrate the protective efficacy of the ZIKV-VLP vaccine and highlight the role that neutralizing antibodies play in protection against ZIKV infection.

[0005] One advantage of VLPs is that VLPs structurally mimic the conformation of native viruses but do not contain any viral genetic material (no viral replication) and are therefore non-infectious. This is in contrast to a live attenuated vaccine (which has genetic material) or in the case of insufficient inactivation of killed vaccines (resulting in viral replication). A VLP vaccine approach eliminates concerns associated with such replication for pregnant women and other populations at high risk for suffering the effects of ZIKV infections.

[0006] In one embodiment, a recombinant nucleic acid vector is provided comprising a heterologous promoter operably linked to a sequence encoding flavivirus, e.g., ZIKV, prM/E. In one embodiment, the vector lacks nucleic acid sequences encoding one or more of flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks nucleic acid sequences encoding functional flavivirus capsid, e.g., a protein that aggregates so as to form a viral capsid having a diameter of about 50 to 60 nm or about 45 nm to 70 nm. In one embodiment, the heterologous promoter is expressed in mammalian cells. In one embodiment, the heterologous promoter is a heterologous viral promoter. In one embodiment, the heterologous promoter comprises a CMV promoter, a SV40 promoter, an EF-1 α promoter or a PGK1 promoter. In one embodiment, the flavivirus is a Zika virus. In one embodiment, the vector sequences are from a Zika virus from the East African or West African lineage. In one embodiment, only a portion of flavivirus capsid sequences is included, e.g., a C-terminal portion of a flavivirus capsid that is linked to prM/E sequences as in the poly-protein that is expressed by wild-type flavivirus. In one embodiment, the portion of the capsid sequence includes amino acids 98 to 112 of the capsid protein encoded by SEQ ID NO:1 or a protein having at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 99% or more amino acid sequence identity thereto. In one embodiment, the prM/E sequences have at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 98%, 99% or more amino acid sequence identity to the prM/E sequences encoded by any one of SEQ ID Nos. 1-3, 5 or 11-13. In one embodiment, the portion of the capsid

sequence lacks a NS2B-3 cleavage site, e.g., KEKKRR (SEQ ID NO:10). In one embodiment, the prM/E sequences are operably linked to a heterologous secretion signal. In one embodiment, the vector further comprises an intron and/or enhancer sequence, e.g., 5' to a prM/E coding sequence. In one embodiment, the vector further comprises an intron, internal ribosome entry sequence, or an enhancer sequence, or any combination thereof.

[0007] A recombinant host cell comprising the vector is also provided. In one embodiment, the cell is a mammalian, e.g., Vero cell, HeLa cell or CHO cell, insect or yeast cell. In one embodiment, the cell is a human or simian cell. In one embodiment, the genome of the cell is augmented, e.g., stably augmented, with nucleic acid sequences encoding flavivirus NS2B, e.g., the source of NS2B may be heterologous or homologous to the source for prM/E. In one embodiment, the genome of the cell is augmented, e.g., stably augmented, with nucleic acid sequences encoding flavivirus capsid, e.g., the capsid may be heterologous or homologous to prM/E, which sequences are optionally integrated into the genome of the cell. In one embodiment, the genome of the cell is augmented with nucleic acid sequences encoding flavivirus NS2B, which sequences are optionally integrated into the genome of the cell. In one embodiment, the vector is integrated into the genome of the host cell.

[0008] Also provided is a method to prepare flavivirus VLPs. The method includes contacting a culture of isolated host cells that do not express one or more of flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally do not express functional flavivirus capsid, with the recombinant vector and collecting VLPs from supernatant of the culture. Thus, in one embodiment, the isolated host cells do not have flavivirus sequences prior to contact with the vector. In one embodiment, the collected particles have a diameter of about 10 to 100 nm, e.g., 20 to 60 nm, 40 to 70 nm or 40 to 60 nm. In one embodiment, the host cell expresses flavivirus NS2B. In one embodiment, the host cell expresses flavivirus capsid protein and optionally NS2B.

[0009] Further provided is a preparation comprising a flavivirus VLPs. The VLP comprises a lipid bilayer comprising flavivirus prM/E but lacks one or more of a flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks functional flavivirus capsid. Such a preparation may be used in a vaccine or immunogenic composition. The vaccine or immunogenic composition may have about 10 µg to 1000 µg, e.g., 200 µg to 400 µg or 400 µg to 800 µg, about 0.5 µg to 100 µg, about 1 µg to 50 µg, about 5 µg to 75 µg, about 1 to 500 mg, e.g., about 20 to 50 mg, about 100 to 300 or about 300 to 400 mg, of VLP. The vaccine or immunogenic composition may further comprise one or more adjuvants. In one embodiment, the adjuvant comprises alum, monophosphoryl lipid A (MPLA), squalene, a TLR4 agonist, dimethyldioctadecylammonium, tripalmitoyl-S-glycerol cysteine, trehalose dibehenate; saponin, MF59, AS03, virosomes, AS04, CpG, imidazoquinoline, poly I:C, flagellin, or any combination thereof. In one embodiment, an adjuvant is included at about 0.001 mg to about 10 mg, about 0.01 to about 10 mg, about 1 to about 20 mg, or about 10 mg to about 100 mg.

[0010] Further provided is a method to prevent, inhibit or treat flavivirus infection in a mammal. The method includes administering an effective amount of the recombinant vector, a host cell having the vector or the vaccine or immunogenic composition having the VLPs. In one embodiment,

the mammal is a female mammal. In one embodiment, the vector, host cell, vaccine or immunogenic composition is administered subcutaneously, intradermally, intramuscularly or intravenously to the mammal.

[0011] In one embodiment, a method to passively prevent, inhibit or treat flavivirus infection in a mammal is provided. The method includes obtaining serum or plasma having anti-flavivirus antibodies from a mammal exposed to flavivirus and optionally isolating antibodies from the serum or plasma; and administering an effective amount of the serum or plasma, or isolated antibodies, to a different mammal at risk of or having a flavivirus infection. In one embodiment, the mammal is immunocompromised. In one embodiment, the anti-flavivirus antibodies are isolated from the serum before administration. In one embodiment, the mammal is a human.

BRIEF DESCRIPTION OF THE FIGURES

[0012] FIGS. 1A-E. In vitro characterization of Zika virus like particles. A) Schematic of pCMV-prM/E expression cassette. B) Western blot analysis of Zika virus like particles. Lanes are, 1) Bio-rad precision plus kaleidoscope protein standards. 2) pCMV-prM/E transfection pre sucrose cushion purification supe. 3) 3.5×10^4 PFU ZIKV positive control. 4) pCMV-prM/E transfection post sucrose cushion purification pt. 5) pCMV-GFP transfection post sucrose cushion purification pt. C-E) Sucrose cushion purified Zika VLPs observed using transmission electron microscopy. C) VLPs stained with Tungsten. Diameter is indicated. Background protein staining also apparent. D) VLP stained with Tungsten. Membrane proteins visible on the surface of VLP are indicated with arrow. Background protein staining apparent. E) VLP stained with Uranyl acetate. Membrane proteins visible on the surface of VLP are indicated with an arrow.

[0013] FIGS. 2A-F. Protection of ZIKVLPs in AG129 mice. A) Neutralizing antibody titers (+/-SD) of vaccinated AG129 mice pre boost and pre challenge. B) Average weight loss (+/-SD) of AG129 after ID challenge with 200 PFU ZIKV over a 14 day period. C) Survival of 11 week old AG129 after ID challenge with 200 PFU ZIKV over a 14 day period. D) Viremia (+/-SD) in serum samples from mice two days post challenge by qRT-PCR. Values are total RNA copies per reaction. E) Viremia (+/-SD) in serum samples from mice two days post challenge by TCID₅₀. F) PRNT₅₀ and PRNT₉₀ values (+/-SD) of serum samples taken from ZIKVLP vaccinated AG129 mice post challenge, and pre challenge serum from PBS/alum mice.

[0014] FIGS. 3A-B. ZIKVLP serum transfer to naïve AG129 mice. A) Average weight loss (+/-SD) of 8 week AG129 transferred serum from mice vaccinated with ZIKVLPs after ID challenge with 20 PFU of ZIKV over a 14 day period. B) Survival of AG129 after challenge with ZIKV over a 14 day period.

[0015] FIG. 4. LD50 of ZIKV in AG129 mice. Survival of AG129 after ZIKV over a 14 day period.

[0016] FIG. 5A-B. A) Weight loss of AG129 after ID challenge with 20 PFU ZIKV over a 12 day period. B) Survival of AG129 after ID challenge with 200 PFU ZIKV over a 12 day period.

[0017] FIGS. 6A-B. Sequence of a vector with an exemplary coding sequence to express prM/E (SEQ ID NO:5).

[0018] FIG. 7. Schematic of a pCMV (A) and pTriex4-neo (B) vector for expression of prM/E.

[0019] FIG. 8A-C. Images showing GFP expression in HEK293 cells. A) pTri px4-neo GFP expression, B) pCMV GFP expression, and C) pCMV GFP expression.

[0020] FIG. 9. Western blot analysis of pTriex versus pCMV prM/E expression. Lane 1: Zika virus +; lanes 3,9: pCMV-GFP cells (pt.) and supernatant (sup.); lanes 4,10: pCMV-Columbia pt., sup.; lanes 5,11: pCMV-French-Poly pt., sup.; lanes 6, 12: pTriex-Columbia pt., sup.; and lanes 7, 13: pTriex-French-Poly pt., sup.

[0021] FIG. 10. Anti-Zika antibodies in mice before and after VLP exposure. Mice were injected IP with about 10^6 TCID₅₀ of ZIKV. 5 weeks later the mice were bled, then injected with crude VLP supernatant. Mice were bled 7 days after injection and antibodies analyzed by ZIKV ELISA.

[0022] FIG. 11. Western blot of sucrose purified VLPs. Lane 1: marker; lane 2: VLP 100,000 g precipitation; lane 3: Zika virus +; lane 4: pCMV—French-Poly post sucrose purification; and lane 5: pCMV-GFP post sucrose purification. Cells in T-75 flasks were transfected with pCMV-prM/E, or pCMV-GFP, and supernatants were collected after 3 days, then clarified by centrifugation (15,000 g, 30 minutes), then layered onto a 20% sucrose cushion, and pelleted at 112,000 g for 3.5 hours.

[0023] FIG. 12. Sucrose fractional analysis. Lane 1: marker; lane 2: Zika virus +; lane 3: Cell debris (pt.) from clarification step; lane 4: Supernatant above sucrose cushion post centrifugation; lane 5: marker; lane 6: VLP post purification batch 1: days 0-3; and lane 7: VLP post purification batch 2: days 3-10. A second batch was harvested from transfected flasks (days 3-10). Purified as before, fractions from each sucrose purification step were analyzed to ensure there was no loss during purification.

[0024] FIG. 13. Comparison of protein expression for VLPs produced from pCMV and pTriex constructs.

[0025] FIG. 14. Mouse study. 11 AG129 mice of mixed sex and age were used. VLPs were administered IM along with 1 mg Alum. Challenge virus (100 PFU) was administered ID.

[0026] FIG. 15. Antibody levels two weeks post boost.

[0027] FIG. 16. Survival and morbidity. All controls were moribund on day 9.

[0028] FIGS. 17A-C. Dose response of ZIKVLPs in AG129 mice. A-B) PRNT₅₀ and PRNT₉₀ values (+/-SD) of serum samples taken from AG129 mice administered a prime and boost of 0.45 µg (A) or a prime only of 3.0 µg (B) ZIKVLPs pre and post challenge. C) Survival of 11 week old AG129 after ID challenge with 200 PFU ZIKV over a 14 day period.

[0029] FIGS. 18A-C. Protection of ZIKVLPs in BALB/c mice. A) PRNT₅₀ and PRNT₉₀ values (+/-SD) of serum samples taken from BALB/c mice administered a prime only of 3.0 µg ZIKVLPs post challenge. B) Viremia (+/-SD) in serum samples from mice two days post challenge by qRT-PCR. Values are total RNA copies per reaction. C) Average weight loss (+/-SD) of BALB/c mice after ID challenge with 200 PFU ZIKV over a 14 day period.

DETAILED DESCRIPTION

Definitions

[0030] As used herein, the terms “isolated” refers to in vitro preparation, isolation of a nucleic acid molecule such as a vector or plasmid of the invention or a virus-like particle of the invention, so that it is not associated with in vivo

substances, or is substantially purified from in vitro substances. An isolated virus-like particle preparation is generally obtained by in vitro culture and propagation and is substantially free from infectious agents. As used herein, “substantially free” means below the level of detection for a particular infectious agent using standard detection methods for that agent. As used herein, the term “recombinant nucleic acid” or “recombinant DNA sequence or segment” refers to a nucleic acid, e.g., to DNA, that has been derived or isolated from a source, that may be subsequently chemically altered in vitro, so that its sequence is not naturally occurring, or corresponds to naturally occurring sequences that are not positioned as they would be positioned in the native genome. An example of DNA “derived” from a source, would be a DNA sequence that is identified as a useful fragment, and which is then chemically synthesized in essentially pure form. An example of such DNA “isolated” from a source would be a useful DNA sequence that is excised or removed from said source by chemical means, e.g., by the use of restriction endonucleases, so that it can be further manipulated, e.g., amplified, for use in the invention, by the methodology of genetic engineering.

[0031] A signal peptide (sometimes referred to as signal sequence, secretory signal, e.g., an Oikosin 15 secretory signal, targeting signal, localization signal, localization sequence, transit peptide, leader sequence or leader peptide) is a short (about 5 to 30 amino acids long) peptide present at the N-terminus of proteins that are destined towards the secretory pathway. These proteins include those that reside either inside certain organelles (the endoplasmic reticulum, golgi or endosomes), secreted from the cell, or inserted into most cellular membranes. Although most type I membrane-bound proteins have signal peptides, the majority of type II and multi-spanning membrane-bound proteins are targeted to the secretory pathway by their first transmembrane domain, which biochemically resembles a signal sequence except that it is not cleaved. Signal sequences generally have a tripartite structure, consisting of a hydrophobic core region (h-region) flanked by an n- and c-region. The latter contains the signal peptidase (SPase) consensus cleavage site. Usually, signal sequences are cleaved off co-translationally, the resulting cleaved signal sequences are termed signal peptides.

Exemplary Embodiments

[0032] Zika virus infection transmitted by *Aedes* mosquitoes is now receiving considerable attention due to its associated with microcephaly and Guillain-Barre syndrome. According to the CDC, there have been over 500 cases of travel-related Zika infections in America to date, with no locally-acquired vector-borne cases reported; in contrast, over 700 cases have been reported in US territories, of which nearly all were locally-transmitted.

[0033] Computational analysis has identified ZIKV envelope glycoproteins as a good candidate for vaccine development, as these are the most immunogenic (Shawan, 2015). Several approaches are currently being explored to develop a ZIKV vaccine, including inactivated, recombinant live-attenuated viruses, protein subunit vaccines, or DNA vaccines. A VLP vaccine approach against ZIKV may eliminate concerns of live attenuated vaccines and insufficient inactivation of killed vaccines for pregnant women and other populations at high risk of suffering the devastating effects of ZIKV infections.

[0034] VLPs are structurally mimic the conformation of native virions but do not generate progeny viruses (VLPs are “non-infectious”) and do not contain any viral genetic material. VLPs are known to be highly immunogenic and elicit higher titer neutralizing antibody responses than sub-unit vaccines based on individual proteins (Wang et al., 2013). Such VLPs present viral spikes and other surface components that display linear or conformational epitopes in a repetitive array that effectively results in recognition by B-cells (Metz and Pijlman, 2016). This recognition leads to B cell signaling and MHC class II up-regulation that facilitates the generation of high titer specific antibodies. VLPs from viruses, including hepatitis B virus, West Nile virus and Chikungunya virus, elicit high titer neutralizing antibody responses that contribute to protective immunity in preclinical animal models and in humans (Akahata et al., 2010; Spohn et al., 2010; Wang et al., 2012).

[0035] As mentioned above, a VLP vaccine approach against ZIKV eliminates concerns of live attenuated vaccines and insufficient inactivation of killed vaccines for pregnant women and other populations at high risk of suffering the devastating effects of ZIKV infections. The generation of ZIKV-VLPs containing the prM and E genes as well as the immunogenicity and efficacy testing in the AG129 mouse model is described herein. A position in the secretory signal was identified that likely allows for higher than normal levels of VLP secretion, due to the absence of an auto (NS2b-3) cleavage signal. Using bioinformatic signal sequence prediction tools, the putative signal sequences of ZIKV starting from positions aa 98-aa 112 were examined, and a site was selected that putatively resulted in the highest secretion score. The prM and E genes from ZIKV (Colombian isolate; GenBank accession no. KU646827) were combined with a secretory signal (positions aa 98-aa 112), were cloned into a mammalian expression vector (pCMV-prM/E). HEK-293 cells were transfected and supernatants were harvested from the cells at approximately 10 days post transfection. Transfected HEK-293 cells secreted VLPs with relatively high yields, likely due to the inclusion of a secretory signal that allows for higher than normal levels of VLP secretion. The cell supernatants contained a fraction of extracellular particles that were purified by ultracentrifugation through a sucrose cushion. These particles reacted with known ZIKV antibodies by Western Blot. Western blot analysis also revealed relatively high yields of VLPs after purification, indicating the potential for scalable production. To test the efficacy of this VLP vaccine, AG129 mice susceptible to ZIKV were vaccinated with 2 µg of total protein (about 400-500 ng of VLPs) formulated with 1 mg of adjuvant, and the mice boosted with the same vaccine two weeks later. At two weeks post boost, serum from vaccinated animals was collected and tested for anti-ZIKV neutralizing antibodies. Three weeks post boost mice were challenged with 200 PFU of ZIKV (about 400 LD₅₀s). All control animals (n=6) died by 9 days post challenge, while vaccinated mice survived with no morbidity/illness (as of 11 days post-challenge). Passive transfer of antibodies from vaccinated mice was efficacious in protecting susceptible mice from Zika infections. Thus, the present findings show the protective efficacy of a ZIKV-VLP vaccine and highlight the important role that neutralizing antibodies play in protection against ZIKV infection. Further, passive transfer may be employed as a treatment for immune-compromised patients that cannot receive a vaccine.

[0036] In one embodiment, a recombinant nucleic acid vector is provided comprising a heterologous promoter operably linked to a sequence encoding ZIKV, prM/E. In one embodiment, the vector lacks nucleic acid sequences encoding ZIKV NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks nucleic acid sequences encoding functional ZIKV capsid, e.g., a protein that aggregates so as to form a viral capsid having a diameter of about 50 to 60 nm. In one embodiment, the heterologous promoter is expressed in mammalian cells. In one embodiment, the heterologous promoter is a heterologous viral promoter. In one embodiment, only a portion of ZIKV capsid sequences is included, e.g., a C-terminal portion of a ZIKV capsid that is linked to prM/E sequences as in the polypeptide that is expressed by wild-type flavivirus. In one embodiment, the portion of the capsid sequence includes amino acids 98 to 112 of the capsid protein encoded by SEQ ID NO:1 or a protein having at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 99% or more amino acid sequence identity thereto. In one embodiment, the prM/E sequences have at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 99% or more amino acid sequence identity to the prM/E sequences encoded by any one of SEQ ID Nos. 1-3 or 5. In one embodiment, the portion of the capsid sequence lacks a NS2B-3 cleavage site. In one embodiment, the prM/E sequences are operably linked to a heterologous secretion signal. In one embodiment, the vector further comprises an intron and/or enhancer sequence, e.g., 5' to a prM/E coding sequence.

[0037] A recombinant host cell comprising the vector is also provided. In one embodiment, the cell is a mammalian cell. In one embodiment, the cell is a human or simian cell. In one embodiment, the genome of the cell is augmented, e.g., stably augmented, with nucleic acid sequences encoding ZIKV NS2B, e.g., the source of NS2B may be heterologous or homologous to the source for prM/E. In one embodiment, the genome of the cell is augmented, e.g., stably augmented, with nucleic acid sequences encoding ZIKV capsid, e.g., the capsid may be heterologous or homologous to prM/E. In one embodiment, the vector is integrated into the genome of the host cell.

[0038] Also provided is a method to prepare ZIKV VLPs. The method includes contacting a culture of isolated host cells that do not express ZIKV NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally do not express functional ZIKV capsid, with the recombinant vector and collecting VLPs from supernatant of the culture. Thus, in one embodiment, the isolated host cells do not have ZIKV sequences prior to contact with the vector. In one embodiment, the collected particles have a diameter of about 10 to 100 nm, e.g., 20 to 60 nm, 40 to 70 nm or 40 to 60 nm. In one embodiment, the host cell expresses ZIKV NS2B. In one embodiment, the host cell expresses ZIKV capsid protein and optionally NS2B.

[0039] Further provided is a preparation comprising a ZIKV VLPs. The VLP comprises a lipid bilayer comprising ZIKV prM/E but lacks ZIKV NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks functional ZIKV capsid. Such a preparation may be used in a vaccine or immunogenic composition. The vaccine or immunogenic composition may have about 10 to 1000 µg, e.g., 200 to 400 µg or 400 to 800 µg, or about 1 to about 500 mg, e.g., about 20 to 50 mg, about 100 to 300 or about 300 to 400 mg, of VLP. The vaccine or immunogenic composition may further comprise one or more adjuvants. In one embodiment, an

adjuvant is included at about 0.01 to about 10 mg, about 1 to about 20 mg, or about 10 mg to about 100 mg.

[0040] Further provided is a method to prevent, inhibit or treat ZIKV infection in a mammal. The method includes administering an effective amount of the recombinant vector, a host cell having the vector or the vaccine or immunogenic composition having the VLPs. In one embodiment, the mammal is a female mammal. In one embodiment, the vector, host cell, vaccine or immunogenic composition is administered intradermally, intramuscularly or intravenously to the mammal.

[0041] In one embodiment, a method to passively prevent, inhibit or treat ZIKV infection in a mammal is provided. The method includes obtaining serum or plasma having anti-ZIKV antibodies from a mammal exposed to ZIKV and optionally isolating antibodies from the serum or plasma; and administering an effective amount of the serum or plasma, or isolated antibodies, to a different mammal at risk of or having a ZIKV infection. In one embodiment, the mammal is immunocompromised. In one embodiment, the anti-flavivirus antibodies are isolated from the serum before administration. In one embodiment, the mammal is a human.

Exemplary Adjuvants

[0042] Adjuvants are compounds that enhance the specific immune response against co-inoculated antigens. Adjuvants can be used for various purposes: to enhance the immunogenicity of highly purified or recombinant antigens; to reduce the amount of antigen or the number of immunizations needed for protective immunity; to prime the efficacy of vaccines in newborns, the elderly or immunocompromised persons; or as antigen delivery systems for the uptake of antigens by the mucosa. Ideally, adjuvants should not induce immune responses against themselves and promote an appropriate immune response (i.e., cellular or antibody immunity depending on requirements for protection). Adjuvants can be classified into three groups: active immunostimulants, being substances that increase the immune response to the antigen; carriers being immunogenic proteins that provide T-cell help; and vehicle adjuvants, being oil emulsions or liposomes that serve as a matrix for antigens as well as stimulating the immune response.

[0043] Adjuvant groups include but are not limited to mineral salt adjuvants, e.g., alum-based adjuvants and salts of calcium, iron and zirconium; tensioactive adjuvants, e.g., Quil A which is a saponin derived from an aqueous extract from the bark of *Quillaja saponaria*; Saponins induce a strong adjuvant effect to T-dependent as well as T-independent antigens. Other adjuvant groups are bacteria-derived substances including cell wall peptidoglycan or lipopolysaccharide of Gram-negative bacteria, that enhance immune response against co-administered antigens and which is mediated through activation of Toll-like receptors; lipopolysaccharides (LPS) which are potent B-cell mitogens, but also activate T cells; and trehalose dimycolate (TCM), which simulates both humoral and cellular responses.

[0044] Other adjuvants are emulsions, e.g., oil in water or water in oil emulsions such as FIA (Freund's incomplete adjuvant), Montanide, Adjuvant 65, and Lipovant; liposomes, which may enhance both humoral and cellular immunity; polymeric adjuvants such as biocompatible and biodegradable microspheres; cytokines; carbohydrates; inulin-derived adjuvants, e.g., gamma inulin, a carbohydrate derived from plant roots of the Compositae family, is a

potent humoral and cellular immune adjuvant and algamulin, which is a combination of γ -inulin and aluminium hydroxide. Other carbohydrate adjuvants include polysaccharides based on glucose and mannose including but not limited to glucans, dextrans, lentinans, glucomannans, galactomannans, levans and xylans.

[0045] Some well known parenteral adjuvants, like MDP, monophosphoryl lipid A (MPL) and LPS, also act as mucosal adjuvants. Other mucosal adjuvants poly(DL-lactide-coglycolide) (DL-PLG), cellulose acetate, iminocarbonates, proteinoid microspheres, polyanhydrides, dextrans, as well as particles produced from natural materials like alginates, gelatine and plant seeds.

[0046] Adjuvants for DNA immunizations include different cytokines, polylactic microspheres, polycarbonates and polystyrene particles.

[0047] In one embodiment, adjuvants useful in the vaccines, compositions and methods described herein include, but are not limited to, mineral salts such as aluminum salts, calcium salts, iron salts, and cerium salts, saponin, e.g., Quid A including QS21, squalene (e.g., AS03), TLR ligands, bacterial MDP (N-acetyl muramyl-L-alanyl-D-isoglutamine), lipopolysaccharide (LPS), Lipid A, montanide, Adjuvant 65, Lipovant, Incomplete Freund's adjuvant (IFA), liposomes, microparticles formed of, for example, poly(D,L-lactide (coglycolide)), cytokines, e.g., IFN-gamma or GM-CSF, or carbohydrates such as gamma inulin, glucans, dextrans, lentinans, glucomannans and/or galactomannans.

Pharmaceutical Compositions

[0048] Pharmaceutical compositions of the present invention, suitable for inoculation or for parenteral or oral administration, comprise flavivirus VLPs, optionally further comprising sterile aqueous or non-aqueous solutions, suspensions, and emulsions. The compositions can further comprise auxiliary agents or excipients, as known in the art. See, e.g., Berkow et al., 1987; *Avery's Drug Treatment*, 1987. The composition of the invention is generally presented in the form of individual doses (unit doses).

[0049] Vaccines may contain about 0.1 to 500 ng, 0.1 to 500 μ g, or 1 to 100 μ g, of VLPs. In one embodiment, the vaccine may contain about 100 μ g to about 500 μ g of VLPs. In one embodiment, the vaccine may contain about at least 100 ng of VLPs. In one embodiment, the vaccine may contain about at least 500 ng of VLPs. In one embodiment, the vaccine may contain about at least 1000 ng of VLPs. In one embodiment, the vaccine may contain about at least 50 μ g of VLPs. In one embodiment, the vaccine may contain less than about 750 μ g of VLPs. In one embodiment, the vaccine may contain less than about 250 μ g of VLPs. In one embodiment, the vaccine may contain less than about 100 μ g of VLPs. In one embodiment, the vaccine may contain less than about 40 μ g of VLPs. The vaccine forming the main constituent of the vaccine composition of the invention may comprise a combination of different flavivirus VLPs, for example, at least two of the three types, Chinese, West African or East African.

[0050] Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and/or emulsions, which may contain auxiliary agents or excipients known in the art. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Carriers or occlusive dressings can be used to increase skin

permeability and enhance antigen absorption. Liquid dosage forms for oral administration may generally comprise a liposome solution containing the liquid dosage form. Suitable forms for suspending liposomes include emulsions, suspensions, solutions, syrups, and elixirs containing inert diluents commonly used in the art, such as purified water. Besides the inert diluents, such compositions can also include adjuvants, wetting agents, emulsifying and suspending agents, or sweetening, flavoring, or perfuming agents. See, e.g., Avery's, 1987.

[0051] When a composition of the present invention is used for administration to an individual, it can further comprise salts, buffers, adjuvants, or other substances which are desirable for improving the efficacy of the composition. For vaccines, adjuvants, substances which can augment a specific immune response, can be used. Normally, the adjuvant and the composition are mixed prior to presentation to the immune system, or presented separately, but into the same site of the organism being immunized. Examples of materials suitable for use in vaccine compositions are provided.

[0052] A pharmaceutical composition according to the present invention may further or additionally comprise at least one chemotherapeutic compound, for example, immunosuppressants, anti-inflammatory agents or immune enhancers, chemotherapeutics including, but not limited to, gamma globulin, amantadine, guanidine, hydroxybenzimidazole, interferon- α , interferon- β , interferon- γ , tumor necrosis factor- α , thiosemicarbazones, methisazone, rifampin, ribavirin, a pyrimidine analog, a purine analog, foscarnet, phosphonoacetic acid, acyclovir, dideoxynucleosides, a protease inhibitor, or ganciclovir.

[0053] The composition can also contain variable but small quantities of endotoxin-free formaldehyde, and preservatives, which have been found safe and not contributing to undesirable effects in the organism to which the composition is administered.

Pharmaceutical Purposes

[0054] The administration of the composition (or the antisera that it elicits) may be for either a "prophylactic" or "therapeutic" purpose. When provided prophylactically, the compositions of the invention which are vaccines, are provided before any symptom of a pathogen infection becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate any subsequent infection or one or more symptoms associated with the disease.

[0055] When provided therapeutically, a VLP vaccine is provided upon the detection of a symptom of actual infection. The therapeutic administration of the vaccine serves to attenuate any actual infection. See, e.g., Avery, 1987.

[0056] Thus, a VLP vaccine composition of the present invention may thus be provided either before the onset of infection (so as to prevent or attenuate an anticipated infection) or after the initiation of an actual infection.

[0057] A composition is said to be "pharmacologically acceptable" if its administration can be tolerated by a recipient patient. Such an agent is said to be administered in a "therapeutically effective amount" if the amount administered is physiologically significant. A composition of the present invention is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient, e.g., enhances at least one primary or

secondary humoral or cellular immune response against at least one strain of an infectious flavivirus.

[0058] The "protection" provided need not be absolute, i.e., the flavivirus infection need not be totally prevented or eradicated, if there is a statistically significant improvement compared with a control population or set of patients. Protection may be limited to mitigating the severity or rapidity of onset of symptoms of the flavivirus infection.

Pharmaceutical Administration

[0059] A composition of the present invention may confer resistance to one or more pathogens, e.g., one or more flavivirus strains, by either passive immunization or active immunization. In active immunization, an inactivated or attenuated live vaccine composition is administered prophylactically to a host (e.g., a mammal), and the host's immune response to the administration protects against infection and/or disease. For passive immunization, the elicited antisera can be recovered and administered to a recipient suspected of having an infection caused by at least one flavivirus strain.

[0060] In one embodiment, the vaccine or immune serum is provided to a mammalian female (at or prior to pregnancy or parturition), under conditions of time and amount sufficient to cause the production of an immune response which serves to protect both the female and the fetus or newborn (via passive incorporation of the antibodies across the placenta or in the mother's milk).

[0061] The present invention thus includes methods for preventing or attenuating a disorder or disease, e.g., an infection. As used herein, a vaccine is said to prevent or attenuate an infection if its administration results either in the total or partial attenuation (i.e., suppression) of a symptom or condition of the infection, or in the total or partial immunity of the individual to the disease.

[0062] At least one VLP or composition thereof, of the present invention may be administered by any means that achieve the intended purposes, using a pharmaceutical composition as previously described.

[0063] For example, administration of such a composition may be by various parenteral routes such as subcutaneous, intravenous, intradermal, intramuscular, intraperitoneal, intranasal, oral or transdermal routes. Parenteral administration can be by bolus injection or by gradual perfusion over time. One mode of using a pharmaceutical composition of the present invention is by intramuscular or subcutaneous application. See, e.g., Avery, 1987.

[0064] A typical regimen for preventing, suppressing, or treating a flavivirus related pathology, comprises administration of an effective amount of a vaccine composition as described herein, administered as a single treatment, or repeated as enhancing or booster dosages, over a period up to and including between one week and about 24 months, or any range or value therein.

[0065] According to the present invention, an "effective amount" of a composition is one that is sufficient to achieve a desired biological effect. It is understood that the effective dosage will be dependent upon the age, sex, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect wanted. The ranges of effective doses provided below are not intended to limit the invention and represent suggested dose ranges. However, the dosage will be tailored to the indi-

vidual subject, as is understood and determinable by one of skill in the art. See, e.g., Avery's, 1987; and Ebadi, 1985.

[0066] The invention will be further described by the following non-limiting examples.

EXAMPLE 1

Experimental Procedures

Cells and Viruses

[0067] African Green Monkey kidney cells (Vero) and Human embryonic kidney 293 (HEK293) were obtained from ATCC (ATCC; Manassas, Va., USA) and grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS; Hyclone, Logan, Utah), 2 mM L-glutamine, 1.5 g/L sodium bicarbonate, 100 U/mL of penicillin, 100 µg/mL of streptomycin, and incubated at 37° C. in 5% CO₂. ZIKV strain H/PF/2013 (GenBank: KJ776791), was obtained from Xavier de Lamballerie (European Virus Archive, Marseille France). Virus stocks were prepared by inoculation onto a confluent monolayer of Vero cells.

Animals

[0068] Mice of the 129/Sv background deficient in alpha/beta interferon (IFN-α/β) and IFN-γ receptors (AG129 mice) were obtained from B&K Universal Limited (Hull, England) and were bred in the pathogen-free animal facilities of the University of Wisconsin-Madison School of Veterinary Medicine. Groups of mixed sex mice were used for all experiments.

Production and purification of ZIKV VLPs

[0069] The prM and E genes of ZIKV strain H/PF/2013 with nascent signal sequence were cloned into a pCMV/V expression vector under the control of a cytomegalovirus (CMV) promoter and CMV polyadenylation signal (pCMV-prM/E). Endotoxin free, transfection grade DNA was prepared using Maxiprep kit (Zymo Research, Irvine, Calif.). VLPs were expressed by transfecting 90% confluent monolayers of HEK293 cells in a T-75 flasks with 15 µg of pCMV-prM/E using Eugene HD (Promega, Madison, Wis.) transfection reagent according to manufacturer protocol. The 10 ml supernatant was harvested 72 hours after transfection, and clarified by centrifugation at 15,000 RCF for 30 minutes at 4° C. Clarified supernatants were layered onto a 20% sucrose cushion and ultra-centrifuged in a SW-28 rotor at 112,000 RCF for 3.5 hours at 4° C. Pellet (PT) and supernatant (SUP) fractions at each step were saved for analysis by SDS-PAGE and Western blot. Post sucrose cushion PT were resuspended in Phosphate Buffered Saline (PBS) pH 7.2. Total protein in VLP preparations was quantified by Bradford assay. VLP specific protein was determined by comparing Zika specific bands on SDS-PAGE gels to known concentrations of BSA using ImageJ software.

Western Blot

[0070] VLP fractions were boiled in Laemmli sample buffer (BioRad, Hercules, Calif., USA) and resolved on a 4-20% SDS-PAGE gel (Biorad) by electrophoresis using a Mini-PROTEAN 3 system (BIO-RAD, CA). Gels were electroblotted onto nitrocellulose membranes using a TurboBlot® system. Membranes were blocked in 5% (W/V) skim milk and probed with mouse hyper immune ascites

fluid primary antibody (1:5000) and goat anti-mouse HRP conjugated secondary antibody (1:5000). Membranes were developed using a solid phase 3,3',5,5'-tetramethylbenzidine (TMB) substrate system.

Transmission Electron Microscopy

[0071] Samples were negatively stained for electron microscopy using the drop method. A drop of sample was placed on a Pioloform™ (Ted Pella, Inc.) carbon-coated 300 Mesh Cu grid, allowed to adsorb for 30 seconds, and the excess removed with filter paper. Next, a drop of methylamine tungstate or uranyl acetate (Nano-W, Nanoprobes Inc.) was placed on the still wet grid, and the excess removed. The negatively stained sample was allowed to dry, and was documented in a Philips CM120 (Eindhoven, The Netherlands) transmission electron microscope at 80 kV. Images were obtained using a SIS MegaView III digital camera (Soft Imaging Systems, Lakewood, Color.).

Vaccination and Viral Challenge

[0072] For VLP formulations, 0.45 µg of sucrose cushion purified VLPs was mixed with 0.2% Imject Alum (Thermo Scientific) according to manufacturer's protocol. Groups of AG129 mice were injected intramuscularly (IM) with VLPs mixed with alum (n=5) or PBS mixed with alum (n=6) at 6 weeks of age, and again at 8 weeks of age. Sub-mandibular blood draws were performed pre boost and pre challenge to collect serum for analysis by neutralization assays and for passive transfer studies.

[0073] Vaccinated mice were challenged with 200 PFU of ZIKV strain H/PF/2013 in 25 µl volumes by intradermal (ID) injection into the right hind footpad. Following infection, mice were monitored daily for the duration of the study. Mice that were moribund or that lost greater than 20% of starting weight were humanely euthanized. Sub-mandibular blood draws were performed on day two post challenge (PC) and serum collected to measure viremia.

[0074] For passive transfer studies, 5 naive mice were injected intraperitoneally (IP) with 500 µl of pooled serum from VLP vaccinated, diluted serum (1:5 n=4, 1:10, n=4), or serum from PBS/alum (n=5) treated mice. At 12 hours post transfer, mice were challenged with 20 PFU in 25 µl as above.

Viremia Assays

[0075] Viremia was determined by TCID₅₀ assay. Briefly, serum was serially diluted ten-fold in microtiter plates 263 and added to duplicate wells of Vero cells in 96-well plates, incubated at 37° C. for 5 days, then fixed and 264 stained with 10% (W/V) crystal violet in 10% (V/V) formalin. Plates were observed under a light microscope to determine the 50% tissue culture infective doses (TCID₅₀s). Serum samples were also tested for viral RNA copies by qRT-PCR. RNA was extracted from 0.02 ml of serum using the ZR Viral 267 RNA Kit (Zymo Research, Irvine, Calif.). Viral RNA was quantified by qRT-PCR using the primers and probe designed by Lanciotti et al. (Lanciotti et al., 2008). The qRT-PCR was performed using the iTaq Universal Probes One-Step Kit (BioRad, Hercules, Calif.) on an iCycler instrument (BioRad, Hercules, Calif.). Primers and probe were used at final concentrations of 500 nM and 250 nM respectively. Cycling conditions were as follows: 50° C. for 10 minutes and 95° C. for 2 minutes, followed by 40

cycles of 95° C. for 15 seconds and 60° C. for 30 seconds. Virus concentration was determined by interpolation onto an internal standard curve made up of a 5-point dilution series of in vitro transcribed RNA.

Neutralization Assay

[0076] Serum antibody titers were determined by microneutralization assay. Briefly, serum was incubated at 56° C. for 30 minutes to inactivate complement and then serially diluted two-fold in microtiter plates. 200 PFUs of virus were added to each well and incubated at 37° C. for 1 hour. The virus-serum mixture was added to duplicate wells of Vero cells in 96-well plates, incubated at 37° C. for 5 days, then fixed and stained with 10% (W/V) crystal violet in 10% (V/V) formalin, then observed under a light microscope. The titer was determined as the serum dilution resulting in the complete neutralization of the virus.

Plaque Reduction Neutralization Test

[0077] Serum samples were serially diluted, mixed with 200 PFU of the ZIKV H/PF/2013 strain and incubated for 1 hour at 37° C. This serum/virus mixture was added to confluent layers of Vero cells in 96 well plates and incubated for 1 hour at 37° C., after which the serum/virus mixture was removed and overlay solution (3% CMC, 1×DMEM, 2% FBS and 1×Anti/Anti) was added. After 48 hours of infec-

tion, the monolayers were fixed with 4% PFA, washed twice with PBS, and then incubated with ZIKV hyperimmune mouse ascitic fluid (1:2000, UTMB) diluted in blocking solution (1×PBS, 0.01% Tween-20 and 5% Milk) and incubated overnight at 4° C. Plates were washed three times with PBS-T and then peroxidase-labeled goat anti-mouse secondary antibody (1:2000) was incubated on monolayers for 2 hours at 37° C. Following incubation, cells were washed a final three times with PBS-T and developed using 3-amino-9-ethylcarbazole (AEC)-peroxidase substrate. The amount of formed foci were counted using an 292 ELISPOT plate reader (ImmunoSPOT-Cellular Technology); quality control was performed to each scanned well to ensure accurate counting. Neutralization percentages (Nx) were calculated per sample/replicate/dilution as follows:

$$Nx = \left\{ 100 - \left[100 \left(\frac{A}{\text{Control}} \right) \right] \right\}$$

Where A corresponds to the amount of foci counted in the sample and Control is the geometric mean of foci counted from wells treated with cells and virus only. Data of corresponding transformed dilutions (Log(1/Dilution)) against neutralization percentages per sample was plotted and fitted to a sigmoidal dose-299 response curve to interpolate PRNT₅₀ and PRNT₉₀ values (GraphPad Prism software).

SEQ ID NO: 9:
 mknpkkksgg frivnmlkrg varvspfggl krlpaglllg hgpirmvlai laflrftaik
 pslglinrwg svgkkeamei ikkfkkdlaa mlriinarke kkrrgadtsv givgl11tta
 maaevtrrgs aymyldrnd ageaisfptt lgmnkcyiqi mdlghmcdat msyecpmlde
 gvepddvdcw cnttstwvvy gtchhkkgea rrsrravtlp shstrklqtr sqtwlesrey
 tkhlirvenw ifrnpgfala aaaiawllgs stsqkviylv milliapays ircigvsnrnd
 fvegmsggtw vdvvlehggc vtvmaqdkpt vdielvtttv snmaevrsyc yeasisdmas
 dsrceptgea yldkqsdtqy vckrtlvdrq wngcgflfgk gslvtcakfa cskkmtgksi
 qpenleyrim lsvhgsqhsq mivndtghet denrakveit pnspraeatl ggfgsglglc
 eprtglfdsd lyyltmnnkh wlvhkewfhd iplpwhagad tgtpwhnnke alvefkdaaha
 krqtvvvlgs qegavhtala galeaemdga kgrlssghlk crlkmcklrl kgvsyslcta
 aefttkipae tlhgtvtvve qyagtdgpck vpaqmavdmq tltpvgrlit anpviteste
 nskmmlldp pfgdsvyivig vgekkithhw hrsgstigka featvrgakr mavlgdtawd
 fgsvggalns lgkghihqifg aafkslfggm swfsqiligt llmwlglnlk ngsislmcla
 lggvlflfst avsadvscsv dfskketrcg tgvfvyndve awrdrykyhp dsprl1aaav
 kqawedgicg issvsrmeni mwsrvegel n aileengvql tvvvgsvknp mwrqgqrlpv
 pvnelpghwk awgksyfvra aktnnsfvvd gdtlkecpk hrawnsflve dhgfgvfhts
 vwlkvredys lecdpavigt avkgkeavhs dlgywiesek ndtwrlkrah liemktcewp
 kshltwdgi eesdliipks lagplshhnt regyrtqmkg pwhseleir feecpgtkvh
 veetcgtrgp slrsttasgr vieewccrec tmpplsfrak dgcwygmeir prkepesnlv
 rsmvtagstd hmdhfslgvl villmvgegl kkrmttkiii stsmavlvam ilggfmsmdl
 aklailmgat faemntggdv ahlaliaaafk vrpallvsfi franwtpres mllalasc11

-continued

qtaisaalegd lmvlingfal awlairaamvv prtdnitlai laaltplarg tllvawragl
atcggmlls lkgkgsvkkn lpfvmaigl avrlvdpinv vgl1111trsg krswwpsevl
tavglicala ggfakadiem agpmaavgl1 ivsyvvsgks vdmwieragd itwekdaevt
gnsprldval desgdfslve ddgppmreii lkvvlmticg mnpiaipfaa gawyvyvktg
krsгалwdvp apkevkkget tdgvyrvmtr rllgstqvgv gvmqegvfht mwhvtkgsal
rsgegrldpy wgdvkqdlvs ycgpwkldaa wdghsevql1 avppgerarn iqt1pgifkt
kdgdigaval dypagtsesp ildkcgrvig lynggvvikh gsyvsaitqg rreetpvec
fepsm1kkkq ltvlldhpga gktrrvlpei vreaiktrlr tvilaptrvv aaemeealrg
lpvrymttav nvthsgteiv dlmchatfts rllqp1rvpn ynlyimdeah ftdpssiaar
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tvwfvpsvrn gneiaacltk agkrviqlsr ktfetefqkt khqewdfvvt tdisemganf
kadrvidsrr clkpvildge rvilagpmpv thasaaqrrg rigrnpnkpq deyllygggca
etdedhahwl earmldniy lqdgliasly rpeadkvaai egefk1rteq rktfvelmkr
gd1pvwlayq vasagitytd rrwcfdg1tn nt1medsvpa evwtrhgekr vlkprwmdar
vcsdhaalks fkefaagkrg aafgvmealg t1pghmterf qeaidnlavl mraetasrpy
kaaaqlpet letim1lg1l gtvs1giffv lmrnk1gigkm gfgmvt1gas awlmw1seie
pariacvliv v1111lv1ip epekqrspqd nqmaiiimva vgl1glitan elglw1ertks
dlshlmgrre egatigfsm1 idlrpasawa iyaalttfit pavqhavtts ynnyslmama
tqagv1fgmg kgmpfyawdf gvpllmigcy sq1t1pt1liv ai11lvahvm ylipglqaaa
araaqkrtaa gimknpvvdg ivvtdid1tm idp1vekkmg qv1liavavs sailsrtawg
wgeaqalita atst1wegsp nkywnsstat slcn1frgsy lagas1iytv trnaglvkrr
gggtget1ge kwkarlnqms alefysykks gitevcreea rralkdgvat gghavsr1gsa
klrwlvergy lqpygkvidl gcgrgwsy aatirkvqev kgytkggpgh eepmlvqsyg
wnivrlksgv dvfhmaae1c dt11cdiges ssspeveear t1rv1smvgd wlekrp1gafc
ikvlcpytst mmetler1lqr rvggglvrvp lsrnsthemv wvsgaksnti ksvsttsq1l
lgrmdgpr1p vk1eedvnl1 sgtravvsca eapnmkiign rierir1seha etwffdenhp
yrtwayhgsy eaptqgsass lingvvr1ls kpwdvvtgvt g1amt1t1py gqqr1v1kek1v
dtrvpdpqeg trqvmsmvss w1wkelgkhk rprvctkeef inkvr1naal gaifeeekew
ktaveavndp rfwalvdker eh1lrgecqs cvynmmgkre kkggefgkak gsraiwymw1
garflefeal gflnedhwm1 rensgggveg lglqr1gyv1 eemsrip1gr myaddtagwd
tr1srfden ealitnqmek ghralalaii ktyqnkvk v1rpaekgt vmdisr1qdq
rgsgqvvt1ya 1ntftn1lvq l1rnmeaeev lem1dlw1lr rsekvt1nw1q sngwdr1lkr1m
avsgddcvk1 pid1rfahal r1f1ndmgkvr kdtqewk1pt gwdnw1e1vpf cshhfnk1hl
kdgrsivvp1 rhqdeligra rvs1pgagws1 retac1aksy aqmwq1lyf1h rrd1rlmana
icssvpvdwv ptgrttwsih gkgewmtted mlvvnrvwi eendhmedkt pvtkwtdip1y
lgkredlw1c slighr1ptt waenikntvn mvrriigdee kymdylstq1v rylgeegst1p
gv1

RESULTS

Expression and Purification of Soluble, Zika VLPs

[00778] To generate Zika VLPs (ZIKVLPs), the prM/E genes with a native signal sequence were cloned into a pCMV expression vector (pCMV-prM/E) (FIG. 1A), transfected HEK293 cells and harvested supernatants (supe) 3 days post transfection. 78 μ g total protein was recovered from post sucrose purification of which 21.6 μ g was VLP protein. Western blot analysis of this pCMV-prM/E supe. revealed expression of about 50 kDa size band (FIG. 1B, lane 2) that corresponded in size to the predicted size of the Zika virus E gene, and additionally matched positive control Zika virus stocks (FIG. 1B, lane 3). To test the hypothesis that expression of Zika prM and E genes spontaneously form extracellular particles, supernatants from pCMV-prM/E and pCMV-GFP (negative control) transfected cells were centrifuged on a sucrose cushion (SC) sufficient for pelleting of flavivirus particles from cell culture proteins (Merino-Ramos et al., 2014). pCMV-prM/E SC purified pellet (pt) appeared to contain high levels of E protein, while pCMV-GFP pt. did not, indicating that staining was specific to expression of 100 prM and E genes.

[00779] To determine if the immune reactive extracellular particles were virus like in nature, transmission electron microscopy (TEM) was performed on pCMV-prM/E SC pt. material. TEM revealed flavivirus 103 like particles with a size that ranged from 30-60 nm (data not show), and a typical size of about 50 nm (FIG. 1C). High magnification images demonstrated surface structures characteristic of flavivirus envelope proteins (FIGS. 1D, E).

Administration of ZIKVLPs is Immunogenic and Protects Highly ZIKV Susceptible $\alpha/\beta/\gamma$ Interferon Deficient Mice

[0080] Mice that received ZIKVLPs developed low levels (GMT=1:9.2) of neutralizing antibodies (nAbs) at 109 two weeks post administration, that increased two weeks after boost (GMT=1:32). Five weeks after primary vaccination, all mice were challenged with 200 PFU of ZIKV by the ID route. Mice administered ZIKVLP maintained weight, while mice that received PBS/alum experienced significant weight loss associated morbidity throughout the challenge period.

[0081] All control mice (n=6) died 9 days after ZIKV challenge. Mice administered ZIKVLP survived with no apparent morbidity. Finally, ZIKVLP vaccinated mice had significantly lower levels of viremia on day 2 post challenge than control mice detected by qRT-PCR ($p=0.0356$) and 116 TCID₅₀ assay ($p=0.0493$).

ZIKVLPs Elicit Plaque Reducing Neutralizing Antibody Titers in Mice That Can Be Passively Transferred to Naïve Mice.

[0082] The plaque reduction neutralization test (PRNT) assay is widely considered to be the “gold standard” for characterizing and quantifying circulating levels of anti-dengue and other flaviviral neutralizing antibodies (nAb) (Thomas et al., 2009). A PRNT assay was developed for rapidly measuring ZIKV specific neutralizing antibodies. Pooled serum samples collected from mice pre-challenge, as well as individual serum samples collected from mice post-challenge were tested by this PRNT assay. Pre-challenge, pooled serum from mice administered ZIKVLP had a cal-

culated 90% plaque reduction (PRNT₉₀) titer of 1:34. The PRNT₉₀ titer increased 2 weeks post challenge (GMT=126 662).

[0083] To test the role of anti-ZIKV antibodies in protection against challenge, groups of mice received ZIKVLP 128 antiserum, undiluted (n=5), diluted 1:5 (n=4), or 1:10 (n=4). As a negative control mice (n=5) were transferred serum from mice previously vaccinated with PBS alum.

[0084] Negative control mice rapidly lost weight starting after day 7 and all died day 9 post challenge. Mice that received undiluted serum maintained weight throughout the 12 day period post challenge, and showed no signs of infection. Mice that received diluted anti-ZIKV antibodies were not protected from challenge, although survival and weight loss were slightly extended relative to negative control mice 134.

DISCUSSION

[0085] Most experts and public health workers agree that a Zika vaccine is urgently needed. In February 2016, the World Health Organization declared that the recent clusters of microcephaly and other neurological disorders in Brazil constitute a public health emergency of international concern. Their recommendations included enhanced surveillance and research, as well as aggressive measures to reduce infection with Zika virus, particularly amongst pregnant women and women of childbearing age. ZIKV is now receiving considerable attention due to its rapid spread in the Americas, and its association with microcephaly (Mlakar et al., 2016) and Guillain-Barre syndrome (Pinto Junior et al., 2015). In our studies, we designed a ZIKV-virus-like particle (VLP) vaccine, demonstrated expression in vitro by western blot and transmission electron microscopy, and tested the protective efficacy and role of antibodies in protection in the AG129 mouse model.

[0086] Although the transfection and purification procedures for this ZIKV-VLP have yet to be optimized, we had an overall calculated yield of 2.2 mg/ml. Similar expression levels have been reported for other flavivirus VLP expression strategies (Pijlman, 2015). Future work will optimize VLP production and purification parameters, which should significantly increase both yield and purity. Stably transfected HEK cells that continuously express VLPs allow for scalable production to meet global demand for a ZIKV vaccine.

[0087] ZIKV-VLPs, formulated with alum, induced detectable neutralizing antibodies and protected animals against lethal challenge (>400 LD₅₀s) with no morbidity or weight loss. Pre-challenge GMT neutralizing titers were 1:32, and pooled pre-challenge serum PRNT₉₀ and PRNT₅₀ titers were 1:34 and 1:157 respectively. At a relatively low dose of 450 ng, the present results indicate that the ZIKV VLPs are highly immunogenic. Additionally, the antibody titers we obtained are consistent with those reported for other highly immunogenic flavivirus VLP vaccines (Ohtaki et al., 2010; Pijlman, 2015).

[0088] Vaccinated mice challenged with >400 LD₅₀s had low levels of viremia (mean=127, geometric mean=25.4 TCID₅₀/ml) detected after challenge. Copies of RNA ZIKV genomes in serum of mice were significantly higher than levels of viremia. However, the disparity between viral genome copies and viremia has been observed for other flaviviruses including dengue (Bae et al., 2003). Since AG129 mice are highly susceptible to viral challenge, it is

possible that the challenge dose given for the active vaccination study was artificially high. Additionally, methods for challenging mice from infected mosquito bite should be developed to most accurately mimic natural infection. Animal studies can determine if the ZIK VLP vaccine can protect female mice from contracting ZIKV during pregnancy using established models for such studies (Miner et al., 2016). ZIK-VLP vaccines may be tested in a non-human primate translational model which most accurately mimics human infection.

[0089] A VLP vaccine approach against ZIKV has significant advantages over other technologies as it will eliminate concerns of live attenuated vaccines and insufficient inactivation of killed vaccines for pregnant women and other populations at high risk of suffering the devastating effects of ZIKV infections. In recent years, recombinant virus-like particle (VLP)-based vaccine strategies have been frequently used for novel vaccine design. VLPs are known to be highly immunogenic and elicit higher titer neutralizing antibody responses than subunit vaccines based on individual proteins (Ariano et al., 2010).

[0090] The role of neutralizing antibodies in protecting against ZIKV was demonstrated by antibody passive transfer studies as naive AG129 mice receiving pooled serum from VLP vaccinated animals were fully protected. These results are consistent with previous findings that indicate the important role of antibodies in protecting against many mosquito-borne viruses, such as Japanese encephalitis, yellow fever and chikungunya. In this study, full protection was observed when animals received undiluted serum, with no weight loss or other clinical signs observed. While these studies highlight the importance of serum antibodies in ZIKV protection, upcoming studies will determine the minimum antibody titer needed for protection, whether the ZIKV-VLP can elicit CD8+ responses, and the overall role of cellular immunity in protection. It is also important to

determine whether anti-ZIKV antibodies elicited by the VLPs play any role in dengue protection or disease enhancement.

[0091] In this study, the AG129 IFN receptor-deficient mouse model was used for evaluation of the ZIKV-VLP. Recently, the suitability of mice deficient in IFN- α/β and - γ receptors as an animal model for ZIKV was demonstrated, as they are highly susceptible to ZIKV infection and disease, developing rapid viremic dissemination in visceral organs and brain and dying 7-8 days post-infection (Aliota et al., 2016). The AG129 mouse model exhibits an intact adaptive immune system, despite the lack of an IFN response, and it has been used extensively to evaluate vaccines and antivirals for DENV (Brewoo et al., 2012; Fuchs et al., 2014; Johnson and Roehrig, 1999; Sarathy et al., 2015).

[0092] In the present study, aluminum hydroxide (commonly known as alum) was used as the adjuvant for the ZIKV-VLP preparations. Since its first use in 1932, vaccines containing aluminum-based adjuvants have been successfully administered in humans demonstrating excellent safety. A variety of adjuvant formulations may, however, be employed with ZIKV VLPs to enhance immunogenic potential including adjuvants that facilitate antigen dose sparing, enhanced immunogenicity, and/or broadened pathogen protection.

[0093] Thus, a VLP based Zika vaccine is described herein that elicits protective antibodies in mice, and is safe, suitable for scalable production, and highly immunogenic. Fast-tracking development of this ZIKV vaccine is a public health priority and is crucial for restoring confidence and security to people who wish to have children or reside in, or visit areas in which ZIKV is endemic.

EXAMPLE 2

Exemplary Zika Virus Polyprotein Sequences:

[0094] Accession No. KU646827 (Which is Incorporated by Reference Herein)

(SEQ ID NO: 6)

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(SEQ ID NO: 1)

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KU955593 (full-length)

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ggagtgtgt aa

prM/E proteins include those having at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 99% or more amino acid sequence identity to the prM/E proteins encoded by SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:11, SEQ ID NO:12, or SEQ ID NO:13.

[0095] Capsid proteins include those having at least 80%, 82%, 85%, 87%, 90%, 92%, 95%, 97%, 99% or more amino acid sequence identity to the proteins encoded by one or more of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:11, SEQ ID NO:12, or SEQ ID NO:13.

[0096] An exemplary intron/enhancer sequences useful in a vector include: atgcctggagacgccatccacgtgtttgacctccata-gaagacacgggaccgatccagctccgcccgggaa cgggtgcattg-gaacgcggattccccgtgccaagagtgtacccgctccggatctcagaacag-

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gtaaacctgtccgaatattgcctctcacatctcgtaactcgcgag-
gactggggaccctgtgacgaac (SEQ ID NO:4), or a nucleotide
sequence having at least 80%, 82%, 85%, 87%, 90%, 92%,
95%, 97%, 99% or more nucleotide sequence identity to
SEQ ID NO:4.

[0097] An exemplary vector sequence useful to produce VLPs is shown in FIG. 6 (SEQ ID NO:5).

[0098] An exemplary African lineage Zika isolate has the following nucleotide sequence (SEQ ID NO:11 which encodes the protein provided at Accession No. HQ234500 which is incorporated by reference herein):

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gccgactagg tcccaactgg gagaaccacc tggtaaatcc atagaaaggg aaaatggata
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 gacacagtca acatggtgcg tagaatcata gataatgaaa aaaggtacat ggactaccta
 tccacccaag tacgctactt ggatgaggag aggtccacac ctggaatgct g

[0099] An exemplary Asian lineage Zika isolate has the following sequence (SEQ ID NO:12 which encodes the protein provided at Accession No. HQ234499 which is incorporated by reference herein):

ATGAAAAACC CAAAAAGAA ATCCGGAGGA TTCCGGATTG
 TCAATATGCT AAAACGCGGA GTAGCCCGTG TGAGCCCTTT
 TGGGGGCTTG AAGAGGCTAC CAGCTGGACT TCTGCTGGGT
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 TGAGATTACG GCAATCAAG CCATCACTGG GTCTCATCAA
 TAGATGGGGT TCCGTGGGGA AAAAAGAGGC TATGGAAATA
 ATAAAGAAGT TCAAGAAAGA TCTGGCTGCC ATGCTGAGAA
 TAATCAATGC TAGGAAGGAG AAGAAGAGAC GTGGCGCAGA
 CACCAGTGTC GGAATGTGTT GCCTCCTGCT GACCACAGCC
 ATGGCAGTGG AGGTCACCAG ACGTGGGAGT GCATACTATA
 TGTACTTAGA CAGAAGCGAT GCTGGGGAGG CCATATCTTT
 TCCAACCACA CTGGGGGTGA ATAAGTGTTA CATACAGATC
 ATGGATCTTG GACACATGTG TGATGCCACA ATGAGCTATG
 AATGCCCTAT GTTGATGAG GGGGTAGAAC CAGATGACGT
 CGATTGTGCG TGCAACACGA CATCGACTTG GGTGTGTAC
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 GCAAACGCGG TCGCAGACCT GGTGGAATC AAGAGAATAC
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 TTTGGGAAGC TCAACGAGCC AAAAAGTCAT ATACTTGGTC
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[0100] An exemplary Spodweni virus lineage has the following nucleotide sequence (SEQ ID NO:13 which encodes the protein provided at Accession No. DQ859064, which is incorporated by reference herein:

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EXAMPLE 3

[0101] Exemplary vectors expressing GFP were transfected into HEK293 cells and expression was assessed (FIGS. 7-8). prM/E sequences were also expressed from the two vectors in HEK cells and supernatants and cells analyzed 48 hours later (FIG. 9). Supernatants were concentrated by centrifugation at 100,000 g for 60 minutes. Western blots were analyzed using University of Texas Medical Branch (UTMB) mouse ascites. More VLPs were secreted from pCMV-FP transfected cells (lane 11 in FIG. 9) than pTriex transfected cells (lane 13). Sucrose purified fractions were subjected to Western blot (FIGS. 10-11). pCMV-prM/E SC purified pellet (pt) appeared to contain high levels of E protein while pCMV-GFP pt did not, indicating that staining was specific to expression of prM and E genes. In summary, a pCMVvector expressed more protein than a pTriex vector. VLPs collected at days 3-10 provided for about 60 µg total protein from about 100 mL. On day 3 the productivity of the cells was about 50 µg per 15 mL (3.3 µg per mL, or 3.3 mg/L). For stably transfected cells, a marker, e.g., a Zeocin resistance gene, may be introduced into the vector that expresses prM/E.

[0102] ZIKV VLPS (ZIKVLPs) formulated with alum were injected into 6-8-week-old interferon deficient A129 and AG129 mice. Control mice received PBS/alum. Animals were challenged with 200 PFU (>400 LD₅₀s) of ZIKV strain H/PF/2013. All vaccinated mice survived with no morbidity or weight loss while control animals either died at 9 days post challenge (AG129) or had increased viremia (A129). Neutralizing antibodies were observed in all ZIKVLP vaccinated mice.

EXAMPLE 4

Materials and Methods

Cells and Viruses

[0103] African Green Monkey kidney cells (Vero) and Human embryonic kidney 293 (HEK293) were obtained from ATCC (ATCC; Manassas, Va., USA) and grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS; Hyclone, Logan, Utah), 2 mM L-glutamine, 1.5 g/l sodium bicarbonate, 100 U/ml of penicillin, 100 µg/ml of streptomycin, and incubated at 37° C. in 5% CO₂. ZIKV strain H/PF/2013 (GenBank: KJ776791), was obtained from Xavier de Lamballerie (European Virus Archive, Marseille France). Virus stocks were prepared by inoculation onto a confluent monolayer of Vero cells.

Animals

[0104] Mice of the 129/Sv background deficient in alpha/beta interferon alpha/beta/gamma (IFN-α/β/IFN-γ) receptors (AG129 mice) were obtained from B&K Universal Limited (Hull, England) and were bred in the pathogen-free animal facilities of the University of Wisconsin-Madison School of Veterinary Medicine. 5-week-old BALB/c mice (The Jackson Laboratory, Maine, USA) were used for wild-type vaccination studies. Groups of mixed sex mice were used for all experiments.

Production and Purification of ZIKV VLPs

[0105] The prM and E genes of ZIKV strain H/PF/2013 with nascent signal sequence were cloned into a pCMV expression vector under the control of a cytomegalovirus (CMV) promoter and CMV polyadenylation signal (pCMV-prM/E, FIG. 1). Endotoxin free, transfection grade DNA was prepared using Maxiprep kit (Zymo Research, Irvine, Calif.). VLPs were expressed by transfecting 90% confluent monolayers of HEK293 cells in a T-75 flasks with 15 µg of pCMV-prM/E using Fugene HD (Promega, Madison, Wis.) transfection reagent according to manufacturer protocol. The 10 ml supernatant was harvested 72 hr after transfection, and clarified by centrifugation at 15,000 RCF for 30 min at 4° C. Clarified supernatants were layered onto a 20% sucrose cushion and ultra-centrifuged in a SW-28 rotor at 112,000 RCF for 3.5 hours at 4° C. Pellet (PT) and supernatant (SUP.) fractions at each step were saved for analysis by SDS-PAGE and Western blot. Post sucrose cushion PT were resuspended in Phosphate Buffered Saline (PBS) pH 7.2. Total protein in VLP preparations was quantified by Bradford assay. VLP specific protein was determined by comparing Zika specific bands on SDS-PAGE gels to known concentrations of BSA using ImageJ software.

Western Blot

[0106] VLP fractions were boiled in Laemmli sample buffer (BioRad, Hercules, Calif., USA) and resolved on a 4-20% SDS-PAGE gel (Biorad) by electrophoresis using a Mini-PROTEAN 3 system (BIO-RAD, CA). Gels were electroblotted onto nitrocellulose membranes using a TurboBlot® system. Membranes were blocked in 5% (W/V) skim milk and probed with mouse hyper immune ascites fluid primary antibody (1:5000) and goat anti-mouse HRP conjugated secondary antibody (1:5000). Membranes were developed using a solid phase 3,3',5,5'-tetramethylbenzidine (TMB) substrate system.

Transmission Electron Microscopy

[0107] Samples were negatively stained for electron microscopy using the drop method. A drop of sample was placed on a Pioloform™ (Ted Pella, Inc.) carbon-coated 300 Mesh Cu grid, allowed to adsorb for 30 seconds, and the excess removed with filter paper. Next, a drop of methylamine tungstate or uranyl acetate (Nano-W, Nanoprobes Inc.) was placed on the still wet grid, and the excess removed. The negatively stained sample was allowed to dry, and was documented in a Philips CM120 (Eindhoven, The Netherlands) transmission electron microscope at 80 kV. Images were obtained using a SIS MegaView III digital camera (Soft Imaging Systems, Lakewood, Colo.).

Vaccination and Viral Challenge

[0108] Each of the following animal studies was performed as one biological replicate. For VLP formulations, the indicated dose of sucrose cushion purified VLPs was mixed with 0.2% Inject Alum (Thermo Scientific) according to manufacturer's protocol. Groups of AG129 mice were injected intramuscularly (IM) with VLPs mixed with alum (n=5) or PBS mixed with alum (n=6) at 6 weeks of age, and again at 8 weeks of age. Sub-mandibular blood draws were

performed pre boost and pre challenge to collect serum for analysis by neutralization assays and for passive transfer studies.

AG129 mice were challenged with 200 PFU of ZIKV strain H/PF/2013 in 25 μ L volumes by intradermal (ID) injection into the right hind footpad at 11 weeks of age. Balb/c mice were vaccinated once at 5 weeks of age as above, and challenged at 13 weeks of age with 200 PFU of H/PF/2013 in 50 μ L by retro orbital injection (IV route).

[0109] Following infection, mice were monitored daily for the duration of the study. Mice that were moribund or that lost greater than 20% of starting weight were humanely euthanized. Sub-mandibular blood draws were performed on day two post challenge (PC) and serum collected to measure viremia.

[0110] Eight week old AG129 mice were used for passive transfer studies. Five naive mice were injected intraperitoneally (IP) with 500 μ L of pooled serum from VLP vaccinated, diluted serum (1:5 n=4, 1:10, n=4), or serum from PBS/alum (n=5) treated mice. At 12 h post transfer, mice were challenged with 20 PFU in 25 μ L as above.

Viremia Assays

[0111] Viremia was determined by TCID₅₀ assay. Briefly, serum was serially diluted ten-fold in microtiter plates and added to duplicate wells of Vero cells in 96-well plates, incubated at 37° C. for 5 days, then fixed and stained with 10% (W/V) crystal violet in 10% (V/V) formalin. Plates were observed under a light microscope to determine the 50% tissue culture infective doses (TCID₅₀s). Serum samples were also tested for viral RNA copies by qRT-PCR. RNA was extracted from 0.02ml of serum using the ZR Viral RNA Kit (Zymo Research, Irvine, Calif.). Viral RNA was quantified by qRT-PCR using the primers and probe designed by Lanciotti et al (Lanciotti et al., 2008). The qRT-PCR was performed using the iTaq Universal Probes One-Step Kit (BioRad, Hercules, Calif.) on an iCycler instrument (BioRad, Hercules, Calif.). Primers and probe were used at final concentrations of 500 nM and 250 nM respectively. Cycling conditions were as follows: 50° C. for 10 min and 95° C. for 2 min, followed by 40 cycles of 95° C. for 15 sec and 60° C. for 30 sec. Virus concentration was determined by interpolation onto an internal standard curve made up of a 5-point dilution series of in vitro transcribed RNA, with the lowest copies per reaction being 100.

Neutralization Assay

[0112] Serum antibody titers were determined by microneutralization assay. Briefly, serum was incubated at 56° C. for 30 min to inactivate complement and then serially diluted two-fold in microtiter plates. 200 PFUs of virus were added to each well and incubated at 37° C. for 1 h. The virus-serum mixture was added to duplicate wells of Vero cells in 96-well plates, incubated at 37° C. for 5 days, then fixed and stained with 10% (W/V) crystal violet in 10% (V/V) formalin, then observed under a light microscope. The titer was determined as the serum dilution resulting in the complete neutralization of the virus.

Plaque Reduction Neutralization Test

[0113] Serum samples were serially diluted, mixed with 200 PFU of the ZIKV H/PF/2013 strain and incubated for 1 hr at 37° C. This serum/virus mixture was added to confluent

layers of Vero cells in 96 well plates and incubated for 1 hr at 37° C., after which the serum/virus mixture was removed and overlay solution (3% CMC, 1xDMEM, 2% FBS and 1xAnti/Anti) was added. After 48 hrs of infection, the monolayers were fixed with 4% PFA, washed twice with PBS, and then incubated with ZIKV hyperimmune mouse ascitic fluid (1:2000, UTMB) diluted in blocking solution (1xPBS, 0.01% Tween-20 and 5% Milk) and incubated overnight at 4° C. Plates were washed three times with PBS-T and then peroxidase-labeled goat anti-mouse secondary antibody (1:2000) was incubated on monolayers for 2 hours at 37° C. Following incubation, cells were washed a final three times with PBS-T and developed using 3-amino-9-ethylcarbazole (AEC)-peroxidase substrate. The amount of formed foci were counted using an ELISPOT plate reader (ImmunoSPOT-Cellular Technology); quality control was performed to each scanned well to ensure accurate counting. Neutralization percentages (Nx) were calculated per sample/replicate/dilution as follows:

$$Nx = \left\{ 100 - \left[100 \left(\frac{A}{\text{Control}} \right) \right] \right\}$$

Where A corresponds to the amount of foci counted in the sample and Control is the geometric mean of foci counted from wells treated with cells and virus only. Data of corresponding transformed dilutions (Log(1/Dilution)) against neutralization percentages per sample was plotted and fitted to a sigmoidal dose-response curve to interpolate PRNT₅₀ and PRNT₉₀ values (GraphPad Prism software).

RESULTS

[0114] Expression and Purification of Soluble, Zika VLPs To generate Zika VLPs (ZIKVLPs), we cloned the prM/E genes with native signal sequence into a pCMV expression vector (pCMV-prM/E) (FIG. 1A), transfected HEK293 cells and harvested supernatants (supe.) 3 days post transfection. 78 μ g total protein was recovered from post sucrose purification of which 21.6 μ g was ZIKVLP protein. Western blot analysis of this pCMV-prM/E supe. revealed expression of an about 50 kDa size band (FIG. 1B, lane 2) that corresponded in size to the predicted size of the Zika virus E gene, and additionally matched positive control Zika virus stocks (FIG. 1B, lane 3). To test the hypothesis that expression of Zika prM and E genes spontaneously form extracellular particles, supernatants from pCMV-prM/E and pCMV-GFP (negative control) transfected cells were centrifuged on a sucrose cushion (SC) sufficient for pelleting of flavivirus particles from cell culture proteins (Merino-Ramos et al., 2014). pCMV-prM/E SC purified pellet (pt.) appeared to contain high levels of E protein, indicating that staining was specific to expression of prM and E genes. To determine if the immune reactive extracellular particles were virus like in nature, we performed transmission electron microscopy (TEM) on pCMV-prM/E SC pt. material. TEM revealed virus like particles with a size that ranged from 30-60 nm, and a typical size of about 50 nm (FIGS. 1C-E).

Administration of ZIKVLPs is Immunogenic and Protects Highly ZIKV Susceptible $\alpha/\beta/\gamma$ Interferon Deficient (AG129) Mice

[0115] First, the LD50 of the H/PF/2013 strain in 12 week-old mixed sex AG129 mice was determined. Groups

of mice (n=5) were infected with 5-fold serial dilutions from 2 PFU to 0.02PFU of ZIKV and monitored for 4 weeks following the last mortality. All mice infected with 2 or 0.4 PFU died within the first week of challenge (FIG. 4), while lower doses killed only 1 to 2 mice within the first two weeks. Interestingly, 2 mice infected with 0.2 PFU ZIKV became ill and were euthanized due to weight loss and paralysis 4.5 weeks following challenge. The resultant LD₅₀ value in PFUs was calculated to be 0.19 PFU by the Reed-Muench (REED and MUENCH, 1938) method.

[0116] To determine if ZIKVLPs are immunogenic and protective in highly susceptible AG129 mice, groups of mice received a prime and boost of 450ng ZIKVLPs. AG129 mice that received ZIKVLPs developed low levels (GMT=1:9.2) of neutralizing antibodies (nAbs) at two weeks post administration (FIG. 2A), that increased two weeks after boost (GMT=1:32). Five weeks after primary vaccination, all mice were challenged with 200 PFU (>1000 LD₅₀s) of ZIKV by the ID route. Mice administered ZIKVLPs maintained weight, while mice that received PBS/alum experienced significant morbidity throughout the challenge period (FIG. 20B). All control mice (survival 0/6) died 9 days after ZIKV challenge and had significantly lower survival (p=0.0016) than mice administered ZIKVLPs (survival 5/5, FIGS. 2B and C). Finally, ZIKVLPs vaccinated mice had significantly lower levels of viremia on day 2 post challenge than control mice detected by qRT-PCR (ZIKVLP=1.3×10⁴ RNA copies, PBS/alum 9.6×10⁷ RNA copies, p=0.0356, FIG. 2D) and TCID₅₀ assay (ZIKVLP=1.3×10² TCID₅₀s, PBS/alum 2.8×10⁵ TCID₅₀s p=0.0493, FIG. 2E).

ZIKVLPs Elicit Plaque Reducing Neutralizing Antibody Titers in Mice That Can Be Passively Transferred to Naïve Mice.

[0117] The plaque reduction neutralization test (PRNT) assay is widely considered to be the “gold standard” for characterizing and quantifying circulating levels of anti-dengue and other flaviviral neutralizing antibodies (nAb) (Thomas et al., 2009). A PRNT assay was developed for rapidly measuring ZIKV specific neutralizing antibodies. Pooled serum samples collected from mice pre-challenge, as well as individual serum samples collected from mice post-challenge were tested by this PRNT assay. Pre challenge, pooled serum from mice administered ZIKVLPs had a calculated 50% plaque reduction (PRNT₅₀) titer of 1:157. The PRNT₅₀ titer increased 2 weeks post challenge (GMT=5122) (FIG. 2F).

[0118] To test the role of anti-ZIKV antibodies in protection against challenge, groups of mice received ZIKVLP antiserum (pooled pre challenge serum, titer in FIG. 2F), undiluted (n=5), diluted 1:5 (n=4), or 1:10 (n=4). As a negative control, mice (n=5) were transferred serum from mice previously vaccinated with PBS alum. Negative control mice rapidly lost weight starting after day 7 and all died day 9 post challenge (FIGS. 3A-B). Mice that received undiluted serum maintained weight throughout the 14 day period post challenge, and showed no signs of infection. Mice that received diluted anti-ZIKV antibodies were not protected from challenge, although survival and weight loss were slightly extended relative to negative control mice (FIGS. 3A-B).

A Single Dose of ZIKVLPs Can Protect Highly Susceptible AG129 Mice

[0119] To determine if a single dose could protect AG129 mice, groups of 6-week old AG129 mice were vaccinated with 3 µg ZIKVLPs adjuvanted with alum. An additional group of mice (n=5) was vaccinated with a prime and boost of 0.45 µg adjuvanted with alum for comparison. Negative control mice (n=5) received a prime and boost of PBS/alum. Vaccinated mice developed neutralizing antibodies measured by PRNT assay prior to challenge (FIG. 17A). Eight weeks following primary vaccination mice were challenged with 200 PFU (>1000LD₅₀s) of ZIKV by the ID route. All mice administered a prime of 3 µg or a prime and boost of 0.45 µg ZIKVLPs survived throughout the 6 week challenge period (FIG. 17C) and maintained weight throughout the challenge period. Pre challenge neutralizing antibody titers in both single (GMT PRNT₅₀=288, PRNT₉₀=81) and double dose (GMT PRNT₅₀=235, PRNT₉₀=50) groups increased significantly (p<0.005) in all animals measured at 3 weeks post challenge (FIGS. 17A-B).

ZIKVLPs Protect Wildtype BALB/c Mice

[0120] To determine if ZIKVLPs can protect wildtype BALB/c mice against non-lethal ZIKV challenge, a group (n=6) was vaccinated with a single dose of 3 ZIKVLPs adjuvanted with alum. Negative control mice (n=5) were administered PBS/alum. Eight weeks after vaccination mice were challenged with 200 PFU ZIKV by the IV route. A single dose of ZIKVLPs elicited high titers of neutralizing antibodies (PRNT₅₀=381, PRNT₉₀=75) detected immediately prior to challenge (FIG. 22A). Mice vaccinated with ZIKVLPs were completely protected from viremia on day 2 post challenge (FIG. 18B), and maintained weight throughout the challenge period (FIG. 18C). Negative control animals lost minor amounts of weight beginning at day 2 post challenge, had high levels of viremia and recovered by 2 weeks post challenge. Neutralizing antibodies were undetectable in negative control mice prior to challenge, but increased significantly after challenge (FIG. 18A). Antibody titers in vaccinated mice decreased, but were not significantly different than before ZIKV challenge (FIG. 18A).

DISCUSSION

[0121] Most experts and public health workers agree that a Zika vaccine is urgently needed. In February 2016, the World Health Organization declared that the recent clusters of microcephaly and other neurological disorders in Brazil constitute a public health emergency of international concern. Their recommendations included enhanced surveillance and research, as well as aggressive measures to reduce infection with Zika virus, particularly amongst pregnant women and women of childbearing age. ZIKV is now receiving considerable attention due to its rapid spread in the Americas, and its association with microcephaly (Mlakar et al., 2016) and Guillain-Barre syndrome (Pinto Junior et al., 2015). In these studies, a ZIKV-virus-like particle (VLP) vaccine was designed and it was expressed in vitro as shown by western blot and transmission electron microscopy, and its protective efficacy and role of antibodies in protection in the AG129 mouse model tested. An overall yield of 2.2 mg/L was calculated for the VLP tested. Similar expression levels have been reported for other flavivirus VLP expression strategies (Pijlman, 2015). Future work will optimize VLP

production and purification parameters, which should significantly increase both yield and purity. Stably transfected HEK cells that continuously express VLPs allow for scalable production to help meet global demand for a ZIKV vaccine, which is estimated to be 100 million doses a year.

[0122] ZIKV-VLPs, formulated with alum, induced detectable neutralizing antibodies and protected animals against lethal challenge (>400 LD₅₀s) with no morbidity or mortality. Pre-challenge GMT neutralizing titers were 1:32, and pooled pre-challenge serum PRNT₉₀ and PRNT₅₀ titers were 1:34 and 1:157 respectively. At a relatively low dose of 450 ng, our results indicate that our ZIKVLPs are highly immunogenic. The antibody titers obtained are consistent with those reported for other highly immunogenic flavivirus VLP vaccines (Ohtaki et al., 2010; Pijlman, 2015). Previous work has shown a direct correlation between dose of VLPs and neutralizing antibody titers. For ZIKV, questions remain about the quantitative relationship between dose of VLPs and their effect on neutralizing antibody titers and protection from ZIKV challenge in vivo.

[0123] In the above-described studies, mice were vaccinated with ZIKVLPs and challenged with a homologous strain of ZIKV (H/PE/2013), which raises the question of ZIKVLP specific antibody cross reactivity to heterologous viruses currently circulating in the Americas. Although the H/PE/2013 virus was isolated well before the current outbreak from a patient infected in French Polynesia, there is a high degree of amino acid similarity (about 99%) to endemic South American strains of ZIKV (Faria et al., 2016; Zanluca et al., 2015). Some experts agree that the high serological cross-reactivity among ZIKV strains would allow for a monovalent vaccine (Lazear and Diamond, 2016). Nevertheless, care must be taken to empirically determine if antibody responses elicited by ZIKV LPs cross-react and protect against South American strains. Finally, any future ZIKV vaccination programs should incorporate careful surveillance of circulating strains to help suppress immunological escape, and ensure efficacy of vaccines in human populations.

[0124] Vaccinated AG129 mice challenged with >1000 LD₅₀s had low levels of viremia (1.3×10^2 TCID₅₀s, FIG. 2E) detected after challenge. Copies of RNA ZIKV genomes in serum of mice were significantly higher than levels of viremia. However, the disparity between viral genome copies and viremia has been observed for other flaviviruses including dengue (Bae et al., 2003). Since AG129 mice are highly susceptible to viral challenge, it is possible that the challenge dose given for the active vaccination study was artificially high. Methods for challenging mice from infected mosquito bite should be developed to most accurately mimic natural infection. The most important criteria for any ZIKV vaccine is its ability to prevent placental and fetal pathology in ZIKV infected pregnant women. Recently developed IFN deficient pregnant mouse models can provide an opportunity to assess if vaccination of pregnant animals can protect the fetus from ZIKV-induced pathology. (Miner et al., 2016). Although models for ZIKV infection in pregnant non-human primates (NHP) are still being developed, ZIKV vaccines should be tested in NHP translational models which most accurately mimics human immune responses to vaccination.

[0125] A VLP vaccine approach against ZIKV has significant advantages over other technologies as it will eliminate concerns of live attenuated vaccines and insufficient inactivation of killed vaccines for pregnant women and other

populations at high risk of suffering the devastating effects of ZIKV infections. Production of inactivated vaccines requires high titer growth of infectious virus which may pose a safety concern for workers. Additionally, the production of both attenuated and inactivated ZIKV vaccines is limited to "batch" production, whereas flavivirus VLPs can continuously expressed from stable cell lines. In recent years, recombinant virus-like particle (VLP)-based vaccine strategies have been frequently used for vaccine design. VLPs are known to be highly immunogenic and elicit higher titer neutralizing antibody responses than subunit vaccines based on individual proteins (Ariano et al., 2010).

[0126] The role of neutralizing antibodies in protecting against ZIKV was demonstrated by antibody passive transfer studies as naive AG129 mice receiving pooled serum from VLP vaccinated animals were fully protected. These results are consistent with previous findings that indicate the important role of antibodies in protecting against many insect-borne flaviviruses, such as Japanese encephalitis, west Nile virus, and tick borne encephalitis (Chiba et al., 1999; Kimura-Kuroda and Yasui, 1988; Tesh et al., 2002), even at low levels of circulating antibodies. In this study, full protection was observed when animals received undiluted serum (PRNT₅₀ 1:157), with no weight loss or other clinical signs observed. While these studies highlight the importance of serum antibodies in ZIKV protection, there are still many important questions related to ZIKV immunology. What is the minimum antibody titer needed for protection, do ZIKVLPs elicit CD8+ responses and are these responses involved in protection, and what is the overall role of cellular immunity in protection? It is also important to determine if anti-ZIKV antibodies, particularly those elicited by ZIKVLPs, play any role in dengue protection or disease enhancement.

[0127] In this study AG129 IFN receptor-deficient mice were used. This mouse models are commonly used for the evaluation of arboviral vaccines, including dengue, chikungunya and yellow fever virus (Meier et al., 2009; Partidos et al., 2011; Prestwood et al., 2012). We recently documented the suitability of mice deficient in IFN- α/β and - γ receptors as an animal model for ZIKV, as they are highly susceptible to ZIKV infection and disease, developing rapid viremic dissemination in visceral organs and brain and dying 7-8 days post-infection (Aliota et al., 2016), and evaluated doses as low as 1 PFU. In our current studies we observed consistent lethality at doses below 1 PFU, indicating that there are viral subpopulations refractory for the formation of CPE in cell culture, but still capable of establishing a lethal infection in highly susceptible mice. It is of great interest is that at a very low dose (0.2PFU) two of five mice became ill more than 1 month after infection, as infection with ZIKV typically produces rapid lethality in AG129 mice.

[0128] The current studies challenged mice with 200 PFU at 11 weeks of age. All control mice lost 20% weight, were moribund, and succumbed to by challenge by day 9. ZIKV challenge therefore appears to be completely lethal in both juvenile and adult AG129 mice. The AG129 mouse model exhibits an intact adaptive immune system, despite the lack of an IFN response, and it has been used extensively to evaluate vaccines and antivirals for DENV (Brewer et al., 2012; Fuchs et al., 2014; Johnson and Roehrig, 1999; Sarathy et al., 2015). In our studies WT BALB/c mice did not succumb to infection with ZIKV consistent with previous studies where BALB/c mice were experimentally inocu-

lated with 200 PFU of ZIKV (Larocca et al., 2016). Mice also developed high levels of viremia following IV inoculation. A single dose of VLPs prevented detection of viral RNA copies in serum of vaccinated mice at 2 days post infection—when viremia levels typically peak in the BALB/c model. It is possible that viral replication was completely inhibited, as there was no “boost” response in neutralizing antibodies observed following challenge. Finally, in repeat AG129, and Balb/c mice mouse studies, animals were protected from ZIKV challenge 8 weeks after vaccination. ZIKVLP therefore appear to elicit a potent “memory” response.

[0129] In the present study, aluminum hydroxide (commonly known as alum) was used as the adjuvant for ZIKV-VLP preparations. Since its first use in 1932, vaccines containing aluminum-based adjuvants have been successfully administered in humans demonstrating excellent safety. Adjuvant formulations of ZIKV-VLP may facilitate antigen dose sparing, enhanced immunogenicity, and broadened pathogen protection.

[0130] In summary, a vaccine against ZIKV is currently unavailable, nor is there any specific prophylactic treatment. A VLP based Zika vaccine that elicits protective antibodies in mice, and is safe, suitable for scalable production, and highly immunogenic, is disclosed herein. Fast-tracking development of this ZIKV vaccine is a public health priority and is crucial for restoring confidence and security to people who wish to have children or reside in, or visit areas in which ZIKV is endemic.

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<210> SEQ ID NO 4
<211> LENGTH: 392
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<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: A synthetic oligonucleotide

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<400> SEQUENCE: 4

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<210> SEQ ID NO 6

<211> LENGTH: 867

<212> TYPE: PRT

<213> ORGANISM: Zika virus

<400> SEQUENCE: 6

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Gln	Ile	Leu	Ile	Gly	Thr	Leu	Leu	Met	Trp	Leu	Gly	Leu	Asn	Thr	Lys
465					470					475					480
Asn	Gly	Ser	Ile	Ser	Leu	Met	Cys	Leu	Ala	Leu	Gly	Gly	Val	Leu	Ile
			485					490						495	
Phe	Leu	Ser	Thr	Ala	Val	Ser	Ala	Asp	Val	Gly	Cys	Ser	Val	Asp	Phe
			500					505					510		
Ser	Lys	Lys	Glu	Thr	Arg	Cys	Gly	Thr	Gly	Val	Phe	Val	Tyr	Asn	Asp
		515					520					525			
Val	Glu	Ala	Trp	Arg	Asp	Arg	Tyr	Lys	Tyr	His	Pro	Asp	Ser	Pro	Arg
	530					535					540				
Arg	Leu	Ala	Ala	Ala	Val	Lys	Gln	Ala	Trp	Glu	Asp	Gly	Ile	Cys	Gly
545					550					555					560
Ile	Ser	Ser	Val	Ser	Arg	Met	Glu	Asn	Ile	Met	Trp	Arg	Ser	Val	Glu
			565					570						575	
Gly	Glu	Leu	Asn	Ala	Ile	Leu	Glu	Glu	Asn	Gly	Val	Gln	Leu	Thr	Val
			580					585					590		
Val	Val	Gly	Ser	Val	Lys	Asn	Pro	Met	Trp	Arg	Gly	Pro	Gln	Arg	Leu
		595					600					605			
Pro	Val	Pro	Val	Asn	Glu	Leu	Pro	His	Gly	Trp	Lys	Ala	Trp	Gly	Lys
	610					615					620				
Ser	Tyr	Phe	Val	Arg	Ala	Ala	Lys	Thr	Asn	Asn	Ser	Phe	Val	Val	Asp
625					630					635					640
Gly	Asp	Thr	Leu	Lys	Glu	Cys	Pro	Leu	Lys	His	Arg	Ala	Trp	Asn	Ser
			645					650						655	
Phe	Leu	Val	Glu	Asp	His	Gly	Phe	Gly	Val	Phe	His	Thr	Ser	Val	Trp
			660					665					670		
Leu	Lys	Val	Arg	Glu	Asp	Tyr	Ser	Leu	Glu	Cys	Asp	Pro	Ala	Val	Ile
		675					680					685			
Gly	Thr	Ala	Val	Lys	Gly	Lys	Glu	Ala	Val	His	Ser	Asp	Leu	Gly	Tyr
	690					695					700				
Trp	Ile	Glu	Ser	Glu	Lys	Asn	Asp	Thr	Trp	Arg	Leu	Lys	Arg	Ala	His
705					710					715					720
Leu	Ile	Glu	Met	Lys	Thr	Cys	Glu	Trp	Pro	Lys	Ser	His	Thr	Leu	Trp
			725					730						735	
Thr	Asp	Gly	Ile	Glu	Glu	Ser	Asp	Leu	Ile	Ile	Pro	Lys	Ser	Leu	Ala
			740					745					750		
Gly	Pro	Leu	Ser	His	His	Asn	Thr	Arg	Glu	Gly	Tyr	Arg	Thr	Gln	Met
		755					760					765			
Lys	Gly	Pro	Trp	His	Ser	Glu	Glu	Leu	Glu	Ile	Arg	Phe	Glu	Glu	Cys
	770					775					780				
Pro	Gly	Thr	Lys	Val	His	Val	Glu	Glu	Thr	Cys	Gly	Thr	Arg	Gly	Pro
785					790					795					800
Ser	Leu	Arg	Ser	Thr	Thr	Ala	Ser	Gly	Arg	Val	Ile	Glu	Glu	Trp	Cys
			805					810						815	
Cys	Arg	Glu	Cys	Thr	Met	Pro	Pro	Leu	Ser	Phe	Trp	Ala	Lys	Asp	Gly

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820					825					830					
Cys	Trp	Tyr	Gly	Met	Glu	Ile	Arg	Pro	Arg	Lys	Glu	Pro	Glu	Ser	Asn
	835						840					845			
Leu	Val	Arg	Ser	Met	Val	Thr	Ala	Gly	Ser	Thr	Asp	His	Met	Asp	His
	850					855					860				
Phe	Ser	Leu													
865															
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Met	Lys	Asn	Pro	Lys	Lys	Lys	Ser	Gly	Gly	Phe	Arg	Ile	Val	Asn	Met
1				5				10					15		
Leu	Lys	Arg	Gly	Val	Ala	Arg	Val	Ser	Pro	Phe	Gly	Gly	Leu	Lys	Arg
			20				25						30		
Leu	Pro	Ala	Gly	Leu	Leu	Leu	Gly	His	Gly	Pro	Ile	Arg	Met	Val	Leu
		35					40					45			
Ala	Ile	Leu	Ala	Phe	Leu	Arg	Phe	Thr	Ala	Ile	Lys	Pro	Ser	Leu	Gly
	50					55					60				
Leu	Ile	Asn	Arg	Trp	Gly	Ser	Val	Gly	Lys	Lys	Glu	Ala	Met	Glu	Ile
65				70					75					80	
Ile	Lys	Lys	Phe	Lys	Lys	Asp	Leu	Ala	Ala	Met	Leu	Arg	Ile	Ile	Asn
			85					90					95		
Ala	Arg	Lys	Glu	Lys	Lys	Arg	Arg	Gly	Thr	Asp	Thr	Ser	Val	Gly	Ile
			100					105					110		
Val	Gly	Leu	Leu	Leu	Thr	Thr	Ala	Met	Ala	Val	Glu	Val	Thr	Arg	Arg
		115					120					125			
Gly	Asn	Ala	Tyr	Tyr	Met	Tyr	Leu	Asp	Arg	Ser	Asp	Ala	Gly	Glu	Ala
		130				135					140				
Ile	Ser	Phe	Pro	Thr	Thr	Met	Gly	Met	Asn	Lys	Cys	Tyr	Ile	Gln	Ile
145				150					155					160	
Met	Asp	Leu	Gly	His	Met	Cys	Asp	Ala	Thr	Met	Ser	Tyr	Glu	Cys	Pro
			165					170					175		
Met	Leu	Asp	Glu	Gly	Val	Glu	Pro	Asp	Asp	Val	Asp	Cys	Trp	Cys	Asn
		180					185					190			
Thr	Thr	Ser	Thr	Trp	Val	Val	Tyr	Gly	Thr	Cys	His	His	Lys	Lys	Gly
		195					200					205			
Glu	Ala	Arg	Arg	Ser	Arg	Arg	Ala	Val	Thr	Leu	Pro	Ser	His	Ser	Thr
		210				215					220				
Arg	Lys	Leu	Gln	Thr	Arg	Ser	Gln	Thr	Trp	Leu	Glu	Ser	Arg	Glu	Tyr
225				230					235					240	
Thr	Lys	His	Leu	Ile	Arg	Val	Glu	Asn	Trp	Ile	Phe	Arg	Asn	Pro	Gly
			245					250					255		
Phe	Ala	Leu	Ala	Ala	Ala	Ala	Ile	Ala	Trp	Leu	Leu	Gly	Ser	Ser	Thr
		260					265					270			
Ser	Gln	Lys	Val	Ile	Tyr	Leu	Val	Met	Ile	Leu	Leu	Ile	Ala	Pro	Ala
		275					280					285			
Tyr	Ser	Ile	Arg	Cys	Ile	Gly	Val	Ser	Asn	Arg	Asp	Phe	Val	Glu	Gly
		290				295					300				

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Met	Ser	Gly	Gly	Thr	Trp	Val	Asp	Val	Val	Leu	Glu	His	Gly	Gly	Cys	305	310	315	320
Val	Thr	Val	Met	Ala	Gln	Asp	Lys	Pro	Thr	Val	Asp	Ile	Glu	Leu	Val	325	330	335	
Thr	Thr	Thr	Val	Ser	Asn	Met	Ala	Glu	Val	Arg	Ser	Tyr	Cys	Tyr	Glu	340	345	350	
Ala	Ser	Ile	Ser	Asp	Met	Ala	Ser	Asp	Ser	Arg	Cys	Pro	Thr	Gln	Gly	355	360	365	
Glu	Ala	Tyr	Leu	Asp	Lys	Gln	Ser	Asp	Thr	Gln	Tyr	Val	Cys	Lys	Arg	370	375	380	
Thr	Leu	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys	385	390	395	400
Gly	Ser	Leu	Val	Thr	Cys	Ala	Lys	Phe	Ala	Cys	Ser	Lys	Lys	Met	Thr	405	410	415	
Gly	Lys	Ser	Ile	Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Arg	Ile	Met	Leu	Ser	420	425	430	
Val	His	Gly	Ser	Gln	His	Ser	Gly	Met	Ile	Val	Asn	Asp	Thr	Gly	His	435	440	445	
Glu	Thr	Asp	Glu	Asn	Arg	Ala	Lys	Val	Glu	Ile	Thr	Pro	Asn	Ser	Pro	450	455	460	
Arg	Ala	Glu	Ala	Thr	Leu	Gly	Gly	Phe	Gly	Ser	Leu	Gly	Leu	Asp	Cys	465	470	475	480
Glu	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Ser	Asp	Leu	Tyr	Tyr	Leu	Thr	Met	485	490	495	
Asn	Asn	Lys	His	Trp	Leu	Val	His	Lys	Glu	Trp	Phe	His	Asp	Ile	Pro	500	505	510	
Leu	Pro	Trp	His	Ala	Gly	Ala	Asp	Thr	Gly	Thr	Pro	His	Trp	Asn	Asn	515	520	525	
Lys	Glu	Ala	Leu	Val	Glu	Phe	Lys	Asp	Ala	His	Ala	Lys	Arg	Gln	Thr	530	535	540	
Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Val	His	Thr	Ala	Leu	Ala	545	550	555	560
Gly	Ala	Leu	Glu	Ala	Glu	Met	Asp	Gly	Ala	Lys	Gly	Arg	Leu	Ser	Ser	565	570	575	
Gly	His	Leu	Lys	Cys	Arg	Leu	Lys	Met	Asp	Lys	Leu	Arg	Leu	Lys	Gly	580	585	590	
Val	Ser	Tyr	Ser	Leu	Cys	Thr	Ala	Ala	Phe	Thr	Phe	Thr	Lys	Ile	Pro	595	600	605	
Ala	Glu	Thr	Leu	His	Gly	Thr	Val	Thr	Val	Glu	Val	Gln	Tyr	Ala	Gly	610	615	620	
Thr	Asp	Gly	Pro	Cys	Lys	Val	Pro	Ala	Gln	Met	Ala	Val	Asp	Met	Gln	625	630	635	640
Thr	Leu	Thr	Pro	Val	Gly	Arg	Leu	Ile	Thr	Ala	Asn	Pro	Val	Ile	Thr	645	650	655	
Glu	Ser	Thr	Glu	Asn	Ser	Lys	Met	Met	Leu	Glu	Leu	Asp	Pro	Pro	Phe	660	665	670	
Gly	Asp	Ser	Tyr	Ile	Val	Ile	Gly	Val	Gly	Glu	Lys	Lys	Ile	Thr	His	675	680	685	
His	Trp	His	Arg	Ser	Gly	Ser	Thr	Ile	Gly	Lys	Ala	Phe	Glu	Ala	Thr	690	695	700	
Val	Arg	Gly	Ala	Lys	Arg	Met	Ala	Val	Leu	Gly	Asp	Thr	Ala	Trp	Asp				

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705	710	715	720
Phe Gly Ser Val Gly Gly Ala Leu Asn Ser Leu Gly Lys Gly Ile His	725	730	735
Gln Ile Phe Gly Ala Ala Phe Lys Ser Leu Phe Gly Gly Met Ser Trp	740	745	750
Phe Ser Gln Ile Leu Ile Gly Thr Leu Leu Val Trp Leu Gly Leu Asn	755	760	765
Thr Lys Asn Gly Ser Ile Ser Leu Met Cys Leu Ala Leu Gly Gly Val	770	775	780
Leu Ile Phe Leu Ser Thr Ala Val Ser Ala Asp Val Gly Cys Ser Val	785	790	795
Asp Phe Ser Lys Lys Glu Thr Arg Cys Gly Thr Gly Val Phe Val Tyr	805	810	815
Asn Asp Val Glu Ala Trp Arg Asp Arg Tyr Lys Tyr His Pro Asp Ser	820	825	830
Pro Arg Arg Leu Ala Ala Ala Val Lys Gln Ala Trp Glu Asp Gly Ile	835	840	845
Cys Gly Ile Ser Ser Val Ser Arg Met Glu Asn Ile Met Trp Arg Ser	850	855	860
Val Glu Gly Glu Leu Asn Ala Ile Leu Glu Glu Asn Gly Val Gln Leu	865	870	875
Thr Val Val Val Gly Ser Val Lys Asn Pro Met Trp Arg Gly Pro Gln	885	890	895
Arg Leu Pro Val Pro Val Asn Glu Leu Pro His Gly Trp Lys Ala Trp	900	905	910
Gly Lys Ser Tyr Phe Val Arg Ala Ala Lys Thr Asn Asn Ser Phe Val	915	920	925
Val Asp Gly Asp Thr Leu Lys Glu Cys Pro Leu Lys His Arg Ala Trp	930	935	940
Asn Ser Phe Leu Val Glu Asp His Gly Phe Gly Val Phe His Thr Ser	945	950	955
Val Trp Leu Lys Val Arg Glu Asp Tyr Ser Leu Glu Cys Asp Pro Ala	965	970	975
Val Ile Gly Thr Ala Ala Lys Gly Lys Glu Ala Val His Ser Asp Leu	980	985	990
Gly Tyr Trp Ile Glu Ser Glu Lys Asn Asp Thr Trp Arg Leu Lys Arg	995	1000	1005
Ala His Leu Ile Glu Met Lys Thr Cys Glu Trp Pro Lys Ser His Thr	1010	1015	1020
Leu Trp Thr Asp Gly Ile Glu Glu Ser Asp Leu Ile Ile Pro Lys Ser	1025	1030	1035
Leu Ala Gly Pro Leu Ser His His Asn Thr Arg Glu Gly Tyr Arg Thr	1045	1050	1055
Gln Met Lys Gly Pro Trp His Ser Glu Glu Leu Glu Ile Arg Phe Glu	1060	1065	1070
Glu Cys Pro Gly Thr Lys Val His Val Glu Glu Thr Cys Gly Thr Arg	1075	1080	1085
Gly Pro Ser Leu Arg Ser Thr Thr Ala Ser Gly Arg Val Ile Glu Glu	1090	1095	1100
Trp Cys Cys Arg Glu Cys Thr Met Pro Pro Leu Ser Phe Arg Ala Lys	1105	1110	1115
			1120

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Asp Gly Cys Trp Tyr Gly Met Glu Ile Arg Pro Arg Lys Glu Pro Glu	1125	1130	1135
Ser Asn Leu Val Arg Ser Met Val Thr Ala Gly Ser Thr Asp His Met	1140	1145	1150
Asp His Phe Ser Leu Gly Val Leu Val Ile Leu Leu Met Val Gln Glu	1155	1160	1165
Gly Leu Lys Lys Arg Met Thr Thr Lys Ile Ile Ile Ser Thr Ser Met	1170	1175	1180
Ala Val Leu Val Ala Met Ile Leu Gly Gly Phe Ser Met Ser Asp Leu	1185	1190	1195
Ala Lys Leu Ala Ile Leu Met Gly Ala Thr Phe Ala Glu Met Asn Thr	1205	1210	1215
Gly Gly Asp Val Ala His Leu Ala Leu Ile Ala Ala Phe Lys Val Arg	1220	1225	1230
Pro Ala Leu Leu Val Ser Phe Ile Phe Arg Ala Asn Trp Thr Pro Arg	1235	1240	1245
Glu Ser Met Leu Leu Ala Leu Ala Ser Cys Leu Leu Gln Thr Ala Ile	1250	1255	1260
Ser Ala Leu Glu Gly Asp Leu Met Val Pro Ile Asn Gly Phe Ala Leu	1265	1270	1275
Ala Trp Leu Ala Ile Arg Ala Met Val Val Pro Arg Thr Asp Asn Ile	1285	1290	1295
Thr Leu Ala Ile Leu Ala Ala Leu Thr Pro Leu Ala Arg Gly Thr Leu	1300	1305	1310
Leu Val Ala Trp Arg Ala Gly Leu Ala Thr Cys Gly Gly Phe Met Leu	1315	1320	1325
Leu Ser Leu Lys Gly Lys Gly Ser Val Lys Lys Asn Leu Pro Phe Val	1330	1335	1340
Met Ala Leu Gly Leu Thr Ala Val Arg Leu Val Asp Pro Ile Asn Val	1345	1350	1355
Val Gly Leu Leu Leu Leu Thr Arg Ser Gly Lys Arg Ser Trp Pro Pro	1365	1370	1375
Ser Glu Val Leu Thr Ala Val Gly Leu Ile Cys Ala Leu Ala Gly Gly	1380	1385	1390
Phe Ala Lys Ala Asp Ile Glu Met Ala Gly Pro Met Ala Ala Val Gly	1395	1400	1405
Leu Leu Ile Val Ser Tyr Val Val Ser Gly Lys Ser Val Asp Met Tyr	1410	1415	1420
Ile Glu Arg Ala Gly Asp Ile Thr Trp Glu Lys Asp Ala Glu Val Thr	1425	1430	1435
Gly Asn Ser Pro Arg Leu Asp Val Ala Leu Asp Glu Ser Gly Asp Phe	1445	1450	1455
Ser Leu Val Glu Asp Asp Gly Pro Pro Met Arg Glu Ile Ile Leu Lys	1460	1465	1470
Val Val Leu Met Ala Ile Cys Gly Met Asn Pro Ile Ala Ile Pro Phe	1475	1480	1485
Ala Ala Gly Ala Trp Tyr Val Tyr Val Lys Thr Gly Lys Arg Ser Gly	1490	1495	1500
Ala Leu Trp Asp Val Pro Ala Pro Lys Glu Val Lys Lys Gly Glu Thr	1505	1510	1515
			1520

Thr	Asp	Gly	Val	Tyr	Arg	Val	Met	Thr	Arg	Arg	Leu	Leu	Gly	Ser	Thr			
				1525							1530							
Gln	Val	Gly	Val	Gly	Val	Met	Gln	Glu	Gly	Val	Phe	His	Thr	Met	Trp			
				1540							1545			1550				
His	Val	Thr	Lys	Gly	Ser	Ala	Leu	Arg	Ser	Gly	Glu	Gly	Arg	Leu	Asp			
				1555			1560				1565							
Pro	Tyr	Trp	Gly	Asp	Val	Lys	Gln	Asp	Leu	Val	Ser	Tyr	Cys	Gly	Pro			
				1570			1575				1580							
Trp	Lys	Leu	Asp	Ala	Ala	Trp	Asp	Gly	His	Ser	Glu	Val	Gln	Leu	Leu			
1585				1590			1595								1600			
Ala	Val	Pro	Pro	Gly	Glu	Arg	Ala	Arg	Asn	Ile	Gln	Thr	Leu	Pro	Gly			
				1605			1610								1615			
Ile	Phe	Lys	Thr	Lys	Asp	Gly	Asp	Ile	Gly	Ala	Val	Ala	Leu	Asp	Tyr			
				1620			1625				1630							
Pro	Ala	Gly	Thr	Ser	Gly	Ser	Pro	Ile	Leu	Asp	Lys	Cys	Gly	Arg	Val			
1635				1640			1645											
Ile	Gly	Leu	Tyr	Gly	Asn	Gly	Val	Val	Ile	Lys	Asn	Gly	Ser	Tyr	Val			
1650				1655			1660											
Ser	Ala	Ile	Thr	Gln	Gly	Arg	Arg	Glu	Glu	Glu	Thr	Pro	Val	Glu	Cys			
1665				1670			1675								1680			
Phe	Glu	Pro	Ser	Met	Leu	Lys	Lys	Lys	Gln	Leu	Thr	Val	Leu	Asp	Leu			
				1685			1690								1695			
His	Pro	Gly	Ala	Gly	Lys	Thr	Arg	Arg	Val	Leu	Pro	Glu	Ile	Val	Arg			
1700				1705			1710											
Glu	Ala	Ile	Lys	Thr	Arg	Leu	Arg	Thr	Val	Ile	Leu	Ala	Pro	Thr	Arg			
1715				1720			1725											
Val	Val	Ala	Ala	Glu	Met	Glu	Glu	Ala	Leu	Arg	Gly	Leu	Pro	Val	Arg			
1730				1735			1740											
Tyr	Met	Thr	Thr	Ala	Val	Asn	Val	Thr	His	Ser	Gly	Thr	Glu	Ile	Val			
1745				1750			1755								1760			
Asp	Leu	Met	Cys	His	Ala	Thr	Phe	Thr	Ser	Arg	Leu	Leu	Gln	Pro	Ile			
				1765			1770								1775			
Arg	Val	Pro	Asn	Tyr	Asn	Leu	Tyr	Ile	Met	Asp	Glu	Ala	His	Phe	Thr			
1780				1785			1790											
Asp	Pro	Ser	Ser	Ile	Ala	Ala	Arg	Gly	Tyr	Ile	Ser	Thr	Arg	Val	Glu			
1795				1800			1805											
Met	Gly	Glu	Ala	Ala	Ala	Ile	Phe	Met	Thr	Ala	Thr	Pro	Pro	Gly	Thr			
1810				1815			1820											
Arg	Asp	Ala	Phe	Pro	Asp	Ser	Asn	Ser	Pro	Ile	Met	Asp	Thr	Glu	Val			
1825				1830			1835								1840			
Glu	Val	Pro	Glu	Arg	Ala	Trp	Ser	Ser	Gly	Phe	Asp	Trp	Val	Thr	Asp			
				1845			1850								1855			
His	Ser	Gly	Lys	Thr	Val	Trp	Phe	Val	Pro	Ser	Val	Arg	Asn	Gly	Asn			
				1860			1865				1870							
Glu	Ile	Ala	Ala	Cys	Leu	Thr	Lys	Ala	Gly	Lys	Arg	Val	Ile	Gln	Leu			
1875				1880			1885											
Ser	Arg	Lys	Thr	Phe	Glu	Thr	Glu	Phe	Gln	Lys	Thr	Lys	His	Gln	Glu			
1890				1895			1900											
Trp	Asp	Phe	Val	Val	Thr	Thr	Asp	Ile	Ser	Glu	Met	Gly	Ala	Asn	Phe			
1905				1910			1915								1920			
Lys	Ala	Asp	Arg															

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1925						1930						1935					
Leu	Asp	Gly	Glu	Arg	Val	Ile	Leu	Ala	Gly	Pro	Met	Pro	Val	Thr	His		
1940						1945						1950					
Ala	Ser	Ala	Ala	Gln	Arg	Arg	Gly	Arg	Ile	Gly	Arg	Asn	Pro	Asn	Lys		
1955						1960						1965					
Pro	Gly	Asp	Glu	Tyr	Leu	Tyr	Gly	Gly	Gly	Cys	Ala	Glu	Thr	Asp	Glu		
1970						1975						1980					
Asp	His	Ala	His	Trp	Leu	Glu	Ala	Arg	Met	Leu	Leu	Asp	Asn	Ile	Tyr		
1985						1990						1995					
Leu	Gln	Asp	Gly	Leu	Ile	Ala	Ser	Leu	Tyr	Arg	Pro	Glu	Ala	Asp	Lys		
2005						2010						2015					
Val	Ala	Ala	Ile	Glu	Gly	Glu	Phe	Lys	Leu	Arg	Thr	Glu	Gln	Arg	Lys		
2020						2025						2030					
Thr	Phe	Val	Glu	Leu	Met	Lys	Arg	Gly	Asp	Leu	Pro	Val	Trp	Leu	Ala		
2035						2040						2045					
Tyr	Gln	Val	Ala	Ser	Ala	Gly	Ile	Thr	Tyr	Thr	Asp	Arg	Arg	Trp	Cys		
2050						2055						2060					
Phe	Asp	Gly	Thr	Thr	Asn	Asn	Thr	Ile	Met	Glu	Asp	Ser	Val	Pro	Ala		
2065						2070						2075					
Glu	Val	Trp	Thr	Arg	Tyr	Gly	Glu	Lys	Arg	Val	Leu	Lys	Pro	Arg	Trp		
2085						2090						2095					
Met	Asp	Ala	Arg	Val	Cys	Ser	Asp	His	Ala	Ala	Leu	Lys	Ser	Phe	Lys		
2100						2105						2110					
Glu	Phe	Ala	Ala	Gly	Lys	Arg	Gly	Ala	Ala	Phe	Gly	Val	Met	Glu	Ala		
2115						2120						2125					
Leu	Gly	Thr	Leu	Pro	Gly	His	Met	Thr	Glu	Arg	Phe	Gln	Glu	Ala	Ile		
2130						2135						2140					
Asp	Asn	Leu	Ala	Val	Leu	Met	Arg	Ala	Glu	Thr	Gly	Ser	Arg	Pro	Tyr		
2145						2150						2155					
Lys	Ala	Ala	Ala	Ala	Gln	Leu	Pro	Glu	Thr	Leu	Glu	Thr	Ile	Met	Leu		
2165						2170						2175					
Leu	Gly	Leu	Leu	Gly	Thr	Val	Ser	Leu	Gly	Ile	Phe	Phe	Val	Leu	Met		
2180						2185						2190					
Arg	Asn	Lys	Gly	Ile	Gly	Lys	Met	Gly	Phe	Gly	Met	Val	Thr	Leu	Gly		
2195						2200						2205					
Ala	Ser	Ala	Trp	Leu	Met	Trp	Leu	Ser	Glu	Ile	Glu	Pro	Ala	Arg	Ile		
2210						2215						2220					
Ala	Cys	Val	Leu	Ile	Val	Val	Phe	Leu	Leu	Leu	Val	Val	Leu	Ile	Pro		
2225						2230						2235					
Glu	Pro	Glu	Lys	Gln	Arg	Ser	Pro	Gln	Asp	Asn	Gln	Met	Ala	Ile	Ile		
2245						2250						2255					
Ile	Met	Val	Ala	Val	Gly	Leu	Leu	Gly	Leu	Ile	Thr	Ala	Asn	Glu	Leu		
2260						2265						2270					
Gly	Trp	Leu	Glu	Arg	Thr	Lys	Ser	Asp	Leu	Ser	His	Leu	Met	Gly	Arg		
2275						2280						2285					
Arg	Glu	Glu	Gly	Ala	Thr	Ile	Gly	Phe	Ser	Met	Asp	Ile	Asp	Leu	Arg		
2290						2295						2300					
Pro	Ala	Ser	Ala	Trp	Ala	Ile	Tyr	Ala	Ala	Leu	Thr	Thr	Phe	Ile	Thr		
2305						2310						2315					
Pro	Ala	Val	Gln	His	Ala	Val	Thr	Thr	Ser	Tyr	Asn	Asn	Tyr	Ser	Leu		
2325						2330						2335					

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Met Ala Met Ala Thr Gln Ala Gly Val Leu Phe Gly Met Gly Lys Gly
2340 2345 2350

Met Pro Phe Tyr Ala Trp Asp Phe Gly Val Pro Leu Leu Met Ile Gly
2355 2360 2365

Cys Tyr Ser Gln Leu Thr Pro Leu Thr Leu Ile Val Ala Ile Ile Leu
2370 2375 2380

Leu Val Ala His Tyr Met Tyr Leu Ile Pro Gly Leu Gln Ala Ala Ala
2385 2390 2395 2400

Ala Arg Ala Ala Gln Lys Arg Thr Ala Ala Gly Ile Met Lys Asn Pro
2405 2410 2415

Val Val Asp Gly Ile Val Val Thr Asp Ile Asp Thr Met Thr Ile Asp
2420 2425 2430

Pro Gln Val Glu Lys Lys Met Gly Gln Val Leu Leu Ile Ala Val Ala
2435 2440 2445

Val Ser Ser Ala Ile Leu Ser Arg Thr Ala Trp Gly Trp Gly Glu Ala
2450 2455 2460

Gly Ala Leu Ile Thr Ala Ala Thr Ser Thr Leu Trp Glu Gly Ser Pro
2465 2470 2475 2480

Asn Lys Tyr Trp Asn Ser Ser Thr Ala Thr Ser Leu Cys Asn Ile Phe
2485 2490 2495

Arg Gly Ser Tyr Leu Ala Gly Ala Ser Leu Ile Tyr Thr Val Thr Arg
2500 2505 2510

Asn Ala Gly Leu Val Lys Arg Arg Gly Gly Gly Thr Gly Glu Thr Leu
2515 2520 2525

Gly Glu Lys Trp Lys Ala Arg Leu Asn Gln Met Ser Ala Leu Glu Phe
2530 2535 2540

Tyr Ser Tyr Lys Lys Ser Gly Ile Thr Glu Val Cys Arg Glu Glu Ala
2545 2550 2555 2560

Arg Arg Ala Leu Lys Asp Gly Val Ala Thr Gly Gly His Ala Val Ser
2565 2570 2575

Arg Gly Ser Ala Lys Leu Arg Trp Leu Val Glu Arg Gly Tyr Leu Gln
2580 2585 2590

Pro Tyr Gly Lys Val Ile Asp Leu Gly Cys Gly Arg Gly Gly Trp Ser
2595 2600 2605

Tyr Tyr Ala Ala Thr Ile Arg Lys Val Gln Glu Val Lys Gly Tyr Thr
2610 2615 2620

Lys Gly Gly Pro Gly His Glu Glu Pro Met Leu Val Gln Ser Tyr Gly
2625 2630 2635 2640

Trp Asn Ile Val Arg Leu Lys Ser Gly Val Asp Val Phe His Met Ala
2645 2650 2655

Ala Glu Pro Cys Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser Ser Ser
2660 2665 2670

Ser Pro Glu Val Glu Glu Ala Arg Thr Leu Arg Val Leu Ser Met Val
2675 2680 2685

Gly Asp Trp Leu Glu Lys Arg Pro Gly Ala Phe Cys Ile Lys Val Leu
2690 2695 2700

Cys Pro Tyr Thr Ser Thr Met Met Glu Thr Leu Glu Arg Leu Gln Arg
2705 2710 2715 2720

Arg Tyr Gly Gly Gly Leu Val Arg Val Pro Leu Ser Arg Asn Ser Thr
2725 2730 2735

His	Glu	Met	Tyr	Trp	Val	Ser	Gly	Ala	Lys	Ser	Asn	Thr	Ile	Lys	Ser		
			2740					2745					2750				
Val	Ser	Thr	Thr	Ser	Gln	Leu	Leu	Leu	Gly	Arg	Met	Asp	Gly	Pro	Arg		
			2755					2760					2765				
Arg	Pro	Val	Lys	Tyr	Glu	Glu	Asp	Val	Asn	Leu	Gly	Ser	Gly	Thr	Arg		
			2770					2775					2780				
Ala	Val	Val	Ser	Cys	Ala	Glu	Ala	Pro	Asn	Met	Lys	Ile	Ile	Gly	Asn		
			2785					2790					2795		2800		
Arg	Ile	Glu	Arg	Ile	Arg	Ser	Glu	His	Ala	Glu	Thr	Trp	Phe	Phe	Asp		
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Glu	Asn	His	Pro	Tyr	Arg	Thr	Trp	Ala	Tyr	His	Gly	Ser	Tyr	Glu	Ala		
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Pro	Thr	Gln	Gly	Ser	Ala	Ser	Ser	Leu	Ile	Asn	Gly	Val	Val	Arg	Leu		
			2835					2840					2845				
Leu	Ser	Lys	Pro	Trp	Asp	Val	Val	Thr	Gly	Val	Thr	Gly	Ile	Ala	Met		
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Thr	Asp	Thr	Thr	Pro	Tyr	Gly	Gln	Gln	Arg	Val	Phe	Lys	Glu	Lys	Val		
			2865					2870					2875		2880		
Asp	Thr	Arg	Val	Pro	Asp	Pro	Gln	Glu	Gly	Thr	Arg	Gln	Val	Met	Ser		
			2885					2890					2895				
Met	Val	Ser	Ser	Trp	Leu	Trp	Lys	Glu	Leu	Gly	Lys	His	Lys	Arg	Pro		
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Glu	Ala	Val	Asn	Asp	Pro	Arg	Phe	Trp	Ala	Leu	Val	Asp	Lys	Glu	Arg		
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Glu	His	His	Leu	Arg	Gly	Glu	Cys	Gln	Ser	Cys	Val	Tyr	Asn	Met	Met		
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Gly	Lys	Arg	Glu	Lys	Lys	Gln	Gly	Glu	Phe	Gly	Lys	Ala	Lys	Gly	Ser		
			2980					2985					2990				
Arg	Ala	Ile	Trp	Tyr	Met	Trp	Leu	Gly	Ala	Arg	Phe	Leu	Glu	Phe	Glu		
			2995					3000					3005				
Ala	Leu	Gly	Phe	Leu	Asn	Glu	Asp	His	Trp	Met	Gly	Arg	Glu	Asn	Ser		
			3010					3015					3020				
Gly	Gly	Gly	Val	Glu	Gly	Leu	Gly	Leu	Gln	Arg	Leu	Gly	Tyr	Val	Leu		
			3025					3030					3035		3040		
Glu	Glu	Met	Ser	Arg	Ile	Pro	Gly	Gly	Arg	Met	Tyr	Ala	Asp	Asp	Thr		
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Ala	Gly	Trp	Asp	Thr	Arg	Ile	Ser	Arg	Phe	Asp	Leu	Glu	Asn	Glu	Ala		
			3060					3065					3070				
Leu	Ile	Thr	Asn	Gln	Met	Glu	Lys	Gly	His	Arg	Ala	Leu	Ala	Leu	Ala		
			3075					3080					3085				
Ile	Ile	Lys	Tyr	Thr	Tyr	Gln	Asn	Lys	Val	Val	Lys	Val	Leu	Arg	Pro		
			3090					3095					3100				
Ala	Glu	Lys	Gly														

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3140					3145					3150						
Met	Gln	Asp	Leu	Trp	Leu	Leu	Arg	Arg	Ser	Glu	Lys	Val	Thr	Asn	Trp	
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Leu	Gln	Ser	Asn	Gly	Trp	Asp	Arg	Leu	Lys	Arg	Met	Ala	Val	Ser	Gly	
3170					3175					3180						
Asp	Asp	Cys	Val	Val	Lys	Pro	Ile	Asp	Asp	Arg	Phe	Ala	His	Ala	Leu	
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Arg	Phe	Leu	Asn	Asp	Met	Gly	Lys	Val	Arg	Lys	Asp	Thr	Gln	Glu	Trp	
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Lys	Pro	Ser	Thr	Gly	Trp	Asp	Asn	Trp	Glu	Glu	Val	Pro	Phe	Cys	Ser	
3220					3225					3230						
His	His	Phe	Asn	Lys	Leu	His	Leu	Lys	Asp	Gly	Arg	Ser	Ile	Val	Val	
3235					3240					3245						
Pro	Cys	Arg	His	Gln	Asp	Glu	Leu	Ile	Gly	Arg	Ala	Arg	Val	Ser	Pro	
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Gly	Ala	Gly	Trp	Ser	Ile	Arg	Glu	Thr	Ala	Cys	Leu	Ala	Lys	Ser	Tyr	
3265					3270					3275					3280	
Ala	Gln	Met	Trp	Gln	Leu	Leu	Tyr	Phe	His	Arg	Arg	Asp	Leu	Arg	Leu	
3285					3290					3295						
Met	Ala	Asn	Ala	Ile	Cys	Ser	Ser	Val	Pro	Val	Asp	Trp	Val	Pro	Thr	
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Gly	Arg	Thr	Thr	Trp	Ser	Ile	His	Gly	Lys	Gly	Glu	Trp	Met	Thr	Thr	
3315					3320					3325						
Glu	Asp	Met	Leu	Val	Val	Trp	Asn	Arg	Val	Trp	Ile	Glu	Glu	Asn	Asp	
3330					3335					3340						
His	Met	Glu	Asp	Lys	Thr	Pro	Val	Thr	Lys	Trp	Thr	Asp	Ile	Pro	Tyr	
3345					3350					3355					3360	
Leu	Gly	Lys	Arg	Glu	Asp	Leu	Trp	Cys	Gly	Ser	Leu	Ile	Gly	His	Arg	
3365					3370					3375						
Pro	Arg	Thr	Thr	Trp	Ala	Glu	Asn	Ile	Lys	Asn	Thr	Val	Asn	Met	Met	
3380					3385					3390						
Arg	Arg	Ile	Ile	Gly	Asp	Glu	Glu	Lys	Tyr	Val	Asp	Tyr	Leu	Ser	Thr	
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Gln	Val	Arg	Tyr	Leu	Gly	Glu	Glu	Gly	Ser	Thr	Pro	Gly	Val	Leu		
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<210> SEQ ID NO 8

<211> LENGTH: 3423

<212> TYPE: PRT

<213> ORGANISM: Zika virus

<400> SEQUENCE: 8

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Leu	Lys	Arg	Gly	Val	Ala	Arg	Val	Ser	Pro	Phe	Gly	Gly	Leu	Lys	Arg
			20					25					30		
Leu	Pro	Ala	Gly	Leu	Leu	Leu	Gly	His	Gly	Pro	Ile	Arg	Met	Val	Leu
		35					40					45			
Ala	Ile	Leu	Ala	Phe	Leu	Arg	Phe	Thr	Ala	Ile	Lys	Pro	Ser	Leu	Gly
		50				55					60				
Leu	Ile	Asn	Arg	Trp	Gly	Ser	Val	Gly	Lys	Lys	Glu	Ala	Met	Glu	Ile
65					70				75					80	

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Ile	Lys	Lys	Phe	Lys	Lys	Asp	Leu	Ala	Ala	Met	Leu	Arg	Ile	Ile	Asn
			85						90					95	
Ala	Arg	Lys	Glu	Lys	Lys	Arg	Arg	Gly	Ala	Asp	Thr	Asn	Val	Gly	Ile
			100					105					110		
Val	Gly	Leu	Leu	Leu	Thr	Thr	Ala	Met	Ala	Ala	Glu	Val	Thr	Arg	Arg
		115					120					125			
Gly	Ser	Ala	Tyr	Tyr	Met	Tyr	Leu	Asp	Arg	Asn	Asp	Ala	Gly	Glu	Ala
	130					135					140				
Ile	Ser	Phe	Pro	Thr	Thr	Leu	Gly	Met	Asn	Lys	Cys	Tyr	Ile	Gln	Ile
145					150					155					160
Met	Asp	Leu	Gly	His	Met	Cys	Asp	Ala	Thr	Met	Ser	Tyr	Glu	Cys	Pro
				165					170					175	
Met	Leu	Asp	Glu	Gly	Val	Glu	Pro	Asp	Asp	Val	Asp	Cys	Trp	Cys	Asn
			180					185					190		
Thr	Thr	Ser	Thr	Trp	Val	Val	Tyr	Gly	Thr	Cys	His	His	Lys	Lys	Gly
		195					200					205			
Glu	Ala	Arg	Arg	Ser	Arg	Arg	Ala	Val	Thr	Leu	Pro	Ser	His	Ser	Thr
	210					215					220				
Arg	Lys	Leu	Gln	Thr	Arg	Ser	Gln	Thr	Trp	Leu	Glu	Ser	Arg	Glu	Tyr
225					230					235					240
Thr	Lys	His	Leu	Ile	Arg	Val	Glu	Asn	Trp	Ile	Phe	Arg	Asn	Pro	Gly
			245					250					255		
Phe	Ala	Leu	Ala	Ala	Ala	Ile	Ala	Trp	Leu	Leu	Gly	Ser	Ser	Thr	
		260					265					270			
Ser	Gln	Lys	Val	Ile	Tyr	Leu	Val	Met	Ile	Leu	Leu	Ile	Ala	Pro	Ala
	275						280					285			
Tyr	Ser	Ile	Arg	Cys	Ile	Gly	Val	Ser	Asn	Arg	Asp	Phe	Val	Glu	Gly
	290				295						300				
Met	Ser	Gly	Gly	Thr	Trp	Val	Asp	Val	Val	Leu	Glu	His	Gly	Gly	Cys
305				310						315					320
Val	Thr	Val	Met	Ala	Gln	Asp	Lys	Pro	Thr	Val	Asp	Ile	Glu	Leu	Val
			325					330					335		
Thr	Thr	Thr	Val	Ser	Asn	Met	Ala	Glu	Val	Arg	Ser	Tyr	Cys	Tyr	Glu
		340						345					350		
Ala	Ser	Ile	Ser	Asp	Met	Ala	Ser	Asp	Ser	Arg	Cys	Pro	Thr	Gln	Gly
	355					360					365				
Glu	Ala	Tyr	Leu	Asp	Lys	Gln	Ser	Asp	Thr	Gln	Tyr	Val	Cys	Lys	Arg
	370				375						380				
Thr	Leu	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys
385				390						395					400
Gly	Ser	Leu	Val	Thr	Cys	Ala	Lys	Phe	Ala	Cys	Ser	Lys	Lys	Met	Thr
			405					410					415		
Gly	Lys	Ser	Ile	Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Arg	Ile	Met	Leu	Ser
			420				425					430			
Val	His	Gly	Ser	Gln	His	Ser	Gly	Met	Ile	Val	Asn	Asp	Thr	Gly	His
	435						440				445				
Glu	Thr	Asp	Glu	Asn	Arg	Ala	Lys	Val	Glu	Ile	Thr	Pro	Asn	Ser	Pro
	450				455					460					
Arg	Ala	Glu	Ala	Thr	Leu	Gly	Gly	Phe	Gly	Ser	Leu	Gly	Leu	Asp	Cys
465				470					475						480
Glu	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Ser	Asp	Leu	Tyr	Tyr	Leu	Thr	Met

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485								490					495				
Asn	Asn	Lys	His	Trp	Leu	Val	His	Lys	Glu	Trp	Phe	His	Asp	Ile	Pro		
			500					505					510				
Leu	Pro	Trp	His	Ala	Gly	Ala	Asp	Thr	Gly	Thr	Pro	His	Trp	Asn	Asn		
		515					520					525					
Lys	Glu	Ala	Leu	Val	Glu	Phe	Lys	Asp	Ala	His	Ala	Lys	Arg	Gln	Thr		
	530					535					540						
Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Val	His	Thr	Ala	Leu	Ala		
545					550					555					560		
Gly	Ala	Leu	Glu	Ala	Glu	Met	Asp	Gly	Ala	Lys	Gly	Arg	Leu	Ser	Ser		
			565					570					575				
Gly	His	Leu	Lys	Cys	Arg	Leu	Lys	Met	Asp	Lys	Leu	Arg	Leu	Lys	Gly		
			580					585					590				
Val	Ser	Tyr	Ser	Leu	Cys	Thr	Ala	Ala	Phe	Thr	Phe	Thr	Lys	Ile	Pro		
	595					600						605					
Ala	Glu	Thr	Leu	His	Gly	Thr	Val	Thr	Val	Glu	Val	Gln	Tyr	Ala	Gly		
	610					615					620						
Thr	Asp	Gly	Pro	Cys	Lys	Val	Pro	Ala	Gln	Met	Ala	Val	Asp	Met	Gln		
625					630					635					640		
Thr	Leu	Thr	Pro	Val	Gly	Arg	Leu	Ile	Thr	Ala	Asn	Pro	Val	Ile	Thr		
			645					650						655			
Glu	Ser	Thr	Glu	Asn	Ser	Lys	Met	Met	Leu	Glu	Leu	Asp	Pro	Pro	Phe		
		660						665					670				
Gly	Asp	Ser	Tyr	Ile	Val	Ile	Gly	Val	Gly	Glu	Lys	Lys	Ile	Thr	His		
	675					680						685					
His	Trp	His	Arg	Ser	Gly	Ser	Thr	Ile	Gly	Lys	Ala	Phe	Glu	Ala	Thr		
	690					695					700						
Val	Arg	Gly	Ala	Arg	Arg	Met	Ala	Val	Leu	Gly	Asp	Thr	Ala	Trp	Asp		
705					710					715					720		
Phe	Gly	Ser	Val	Gly	Gly	Ala	Leu	Asn	Ser	Leu	Gly	Lys	Gly	Ile	His		
			725					730						735			
Gln	Ile	Phe	Gly	Ala	Ala	Phe	Lys	Ser	Leu	Phe	Gly	Gly	Met	Ser	Trp		
		740						745					750				
Phe	Ser	Gln	Ile	Leu	Ile	Gly	Thr	Leu	Leu	Met	Trp	Leu	Gly	Leu	Asn		
	755					760						765					
Thr	Lys	Asn	Gly	Ser	Ile	Ser	Leu	Met	Cys	Leu	Ala	Leu	Gly	Gly	Val		
	770					775					780						
Leu	Ile	Phe	Leu	Ser	Thr	Ala	Val	Ser	Ala	Asp	Val	Gly	Cys	Ser	Val		
785					790					795					800		
Asp	Phe	Ser	Lys	Lys	Glu	Thr	Arg	Cys	Gly	Thr	Gly	Val	Phe	Val	Tyr		
			805						810					815			
Asn	Asp	Val	Glu	Ala	Trp	Arg	Asp	Arg	Tyr	Lys	Tyr	His	Pro	Asp	Ser		
		820						825					830				
Pro	Arg	Arg	Leu	Ala	Ala	Ala	Val	Lys	Gln	Ala	Trp	Glu	Asp	Gly	Ile		
	835						840					845					
Cys	Gly	Ile	Ser	Ser	Val	Ser	Arg	Met	Glu	Asn	Ile	Met	Trp	Arg	Ser		
	850					855					860						
Val	Glu	Gly	Glu	Leu	Asn	Ala	Ile	Leu	Glu	Glu	Asn	Gly	Val	Gln	Leu		
865					870					875					880		
Thr	Val	Val	Val	Gly	Ser	Val	Lys	Asn	Pro	Met	Trp	Arg	Gly	Pro	Gln		
				885					890					895			

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Arg	Leu	Pro	Val	Pro	Val	Asn	Glu	Leu	Pro	His	Gly	Trp	Lys	Ala	Trp	900	905	910
Gly	Lys	Ser	Tyr	Phe	Val	Arg	Ala	Ala	Lys	Thr	Asn	Asn	Ser	Phe	Val	915	920	925
Val	Asp	Gly	Asp	Thr	Leu	Lys	Glu	Cys	Pro	Leu	Lys	His	Arg	Ala	Trp	930	935	940
Asn	Ser	Phe	Leu	Val	Glu	Asp	His	Gly	Phe	Gly	Val	Phe	His	Thr	Ser	945	950	955
Val	Trp	Leu	Lys	Val	Arg	Glu	Asp	Tyr	Ser	Leu	Glu	Cys	Asp	Pro	Ala	965	970	975
Val	Ile	Gly	Thr	Ala	Val	Lys	Gly	Lys	Glu	Ala	Val	His	Ser	Asp	Leu	980	985	990
Gly	Tyr	Trp	Ile	Glu	Ser	Glu	Lys	Asn	Asp	Thr	Trp	Arg	Leu	Lys	Arg	995	1000	1005
Ala	His	Leu	Ile	Glu	Met	Lys	Thr	Cys	Glu	Trp	Pro	Lys	Ser	His	Thr	1010	1015	1020
Leu	Trp	Thr	Asp	Gly	Ile	Glu	Glu	Ser	Asp	Leu	Ile	Ile	Pro	Lys	Ser	1025	1030	1035
Leu	Ala	Gly	Pro	Leu	Ser	His	His	Asn	Thr	Arg	Glu	Gly	Tyr	Arg	Thr	1045	1050	1055
Gln	Met	Lys	Gly	Pro	Trp	His	Ser	Glu	Glu	Leu	Glu	Ile	Arg	Phe	Glu	1060	1065	1070
Glu	Cys	Pro	Gly	Thr	Lys	Val	His	Val	Glu	Glu	Thr	Cys	Gly	Thr	Arg	1075	1080	1085
Gly	Pro	Ser	Leu	Arg	Ser	Thr	Thr	Ala	Ser	Gly	Arg	Val	Ile	Glu	Glu	1090	1095	1100
Trp	Cys	Cys	Arg	Glu	Cys	Thr	Met	Pro	Pro	Leu	Ser	Phe	Gln	Ala	Lys	1105	1110	1115
Asp	Gly	Cys	Trp	Tyr	Gly	Met	Glu	Ile	Arg	Pro	Arg	Lys	Glu	Pro	Glu	1125	1130	1135
Ser	Asn	Leu	Val	Arg	Ser	Met	Val	Thr	Ala	Gly	Ser	Thr	Asp	His	Met	1140	1145	1150
Asp	His	Phe	Ser	Leu	Gly	Val	Leu	Val	Ile	Leu	Leu	Met	Val	Gln	Glu	1155	1160	1165
Gly	Leu	Lys	Lys	Arg	Met	Thr	Thr	Lys	Ile	Ile	Ile	Ser	Thr	Ser	Met	1170	1175	1180
Ala	Val	Leu	Val	Ala	Met	Ile	Leu	Gly	Gly	Phe	Ser	Met	Ser	Asp	Leu	1185	1190	1195
Ala	Lys	Leu	Ala	Ile	Leu	Met	Gly	Ala	Thr	Phe	Ala	Glu	Met	Asn	Thr	1205	1210	1215
Gly	Gly	Asp	Val	Ala	His	Leu	Ala	Leu	Ile	Ala	Ala	Phe	Lys	Val	Arg	1220	1225	1230
Pro	Ala	Leu	Leu	Val	Ser	Phe	Ile	Phe	Arg	Ala	Asn	Trp	Thr	Pro	Arg	1235	1240	1245
Glu	Ser	Met	Leu	Leu	Ala	Leu	Ala	Ser	Cys	Leu	Leu	Gln	Thr	Ala	Ile	1250	1255	1260
Ser	Ala	Leu	Glu	Gly	Asp	Leu	Met	Val	Leu	Ile	Asn	Gly	Phe	Ala	Leu	1265	1270	1275
Ala	Trp	Leu	Ala	Ile	Arg	Ala	Met	Val	Val	Pro	Arg	Thr	Asp	Asn	Ile	1285	1290	1295

Thr	Leu	Ala	Ile	Leu	Ala	Ala	Leu	Thr	Pro	Leu	Ala	Arg	Gly	Thr	Leu
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Leu	Val	Ala	Trp	Arg	Ala	Gly	Leu	Ala	Thr	Cys	Gly	Gly	Phe	Met	Leu
		1315					1320					1325			
Leu	Ser	Leu	Lys	Gly	Lys	Gly	Ser	Val	Lys	Lys	Asn	Leu	Pro	Phe	Val
		1330				1335					1340				
Met	Ala	Leu	Gly	Leu	Thr	Ala	Val	Arg	Leu	Val	Asp	Pro	Ile	Asn	Val
1345						1350					1355				1360
Val	Gly	Leu	Leu	Leu	Leu	Thr	Arg	Ser	Gly	Lys	Arg	Ser	Trp	Pro	Pro
				1365					1370					1375	
Ser	Glu	Val	Leu	Thr	Ala	Val	Gly	Leu	Ile	Cys	Ala	Leu	Ala	Gly	Gly
			1380					1385					1390		
Phe	Ala	Lys	Ala	Asp	Ile	Glu	Met	Ala	Gly	Pro	Met	Ala	Ala	Val	Gly
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Leu	Leu	Ile	Val	Ser	Tyr	Val	Val	Ser	Gly	Lys	Ser	Val	Asp	Met	Tyr
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Ile	Glu	Arg	Ala	Gly	Asp	Ile	Thr	Trp	Glu	Lys	Asp	Ala	Glu	Val	Thr
1425					1430						1435				1440
Gly	Asn	Ser	Pro	Arg	Leu	Asp	Val	Ala	Leu	Asp	Glu	Ser	Gly	Asp	Phe
				1445					1450					1455	
Ser	Leu	Val	Glu	Asp	Asp	Gly	Pro	Pro	Met	Arg	Glu	Ile	Ile	Leu	Lys
			1460					1465					1470		
Val	Val	Leu	Met	Thr	Ile	Cys	Gly	Met	Asn	Pro	Ile	Ala	Ile	Pro	Phe
		1475					1480					1485			
Ala	Ala	Gly	Ala	Trp	Tyr	Val	Tyr	Val	Lys	Thr	Gly	Lys	Arg	Ser	Gly
		1490				1495						1500			
Ala	Leu	Trp	Asp	Val	Pro	Ala	Pro	Lys	Glu	Val	Lys	Lys	Gly	Glu	Thr
1505					1510					1515					1520
Thr	Asp	Gly	Val	Tyr	Arg	Val	Met	Thr	Arg	Arg	Leu	Leu	Gly	Ser	Thr
			1525						1530					1535	
Gln	Val	Gly	Val	Gly	Val	Met	Gln	Glu	Gly	Val	Phe	His	Thr	Met	Trp
			1540						1545				1550		
His	Val	Thr	Lys	Gly	Ser	Ala	Leu	Arg	Ser	Gly	Glu	Gly	Arg	Leu	Asp
		1555					1560					1565			
Pro	Tyr	Trp	Gly	Asp	Val	Lys	Gln	Asp	Leu	Val	Ser	Tyr	Cys	Gly	Pro
		1570				1575					1580				
Trp	Lys	Leu	Asp	Ala	Ala	Trp	Asp	Gly	His	Ser	Glu	Val	Gln	Leu	Leu
1585					1590					1595					1600
Ala	Val	Pro	Pro	Gly	Glu	Arg	Ala	Arg	Asn	Ile	Gln	Thr	Leu	Pro	Gly
			1605						1610				1615		
Ile	Phe	Lys	Thr	Lys	Asp	Gly	Asp	Ile	Gly	Ala	Val	Ala	Leu	Asp	Tyr
		1620						1625					1630		
Pro	Ala	Gly	Thr	Ser	Gly	Ser	Pro	Ile	Leu	Asp	Lys	Cys	Gly	Arg	Val
		1635													

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1700						1705						1710					
Glu	Ala	Ile	Lys	Thr	Arg	Leu	Arg	Thr	Val	Ile	Leu	Ala	Pro	Thr	Arg		
1715						1720						1725					
Val	Val	Ala	Ala	Glu	Met	Glu	Glu	Ala	Leu	Arg	Gly	Leu	Pro	Val	Arg		
1730						1735						1740					
Tyr	Met	Thr	Thr	Ala	Val	Asn	Val	Thr	His	Ser	Gly	Thr	Glu	Ile	Val		
1745						1750						1755					
Asp	Leu	Met	Cys	His	Ala	Thr	Phe	Thr	Ser	Arg	Leu	Leu	Gln	Pro	Ile		
1765						1770						1775					
Arg	Val	Pro	Asn	Tyr	Asn	Leu	Tyr	Ile	Met	Asp	Glu	Ala	His	Phe	Thr		
1780						1785						1790					
Asp	Pro	Ser	Ser	Ile	Ala	Ala	Arg	Gly	Tyr	Ile	Ser	Thr	Arg	Val	Glu		
1795						1800						1805					
Met	Gly	Glu	Ala	Ala	Ala	Ile	Phe	Met	Thr	Ala	Thr	Pro	Pro	Gly	Thr		
1810						1815						1820					
Arg	Asp	Ala	Phe	Pro	Asp	Ser	Asn	Ser	Pro	Ile	Met	Asp	Thr	Glu	Val		
1825						1830						1835					
Glu	Val	Pro	Glu	Arg	Ala	Trp	Ser	Ser	Gly	Phe	Asp	Trp	Val	Thr	Asp		
1845						1850						1855					
His	Ser	Gly	Lys	Thr	Val	Trp	Phe	Val	Pro	Ser	Val	Arg	Asn	Gly	Asn		
1860						1865						1870					
Glu	Ile	Ala	Ala	Cys	Leu	Thr	Lys	Ala	Gly	Lys	Arg	Val	Ile	Gln	Leu		
1875						1880						1885					
Ser	Arg	Lys	Thr	Phe	Glu	Thr	Glu	Phe	Gln	Lys	Thr	Lys	His	Gln	Glu		
1890						1895						1900					
Trp	Asp	Phe	Val	Val	Thr	Thr	Asp	Ile	Ser	Glu	Met	Gly	Ala	Asn	Phe		
1905						1910						1915					
Lys	Ala	Asp	Arg	Val	Ile	Asp	Ser	Arg	Arg	Cys	Leu	Lys	Pro	Val	Ile		
1925						1930						1935					
Leu	Asp	Gly	Glu	Arg	Val	Ile	Leu	Ala	Gly	Pro	Met	Pro	Val	Thr	His		
1940						1945						1950					
Ala	Ser	Ala	Ala	Gln	Arg	Arg	Gly	Arg	Ile	Gly	Arg	Asn	Pro	Asn	Lys		
1955						1960						1965					
Pro	Gly	Asp	Glu	Tyr	Leu	Tyr	Gly	Gly	Gly	Cys	Ala	Glu	Thr	Asp	Glu		
1970						1975						1980					
Asp	His	Ala	His	Trp	Leu	Glu	Ala	Arg	Met	Leu	Leu	Asp	Asn	Ile	Tyr		
1985						1990						1995					
Leu	Gln	Asp	Gly	Leu	Ile	Ala	Ser	Leu	Tyr	Arg	Pro	Glu	Ala	Asp	Lys		
2005						2010						2015					
Val	Ala	Ala	Ile	Glu	Gly	Glu	Phe	Lys	Leu	Arg	Thr	Glu	Gln	Arg	Lys		
2020						2025						2030					
Thr	Phe	Val	Glu	Leu	Met	Lys	Arg	Gly	Asp	Leu	Pro	Val	Trp	Leu	Ala		
2035						2040						2045					
Tyr	Gln	Val	Ala	Ser	Ala	Gly	Ile	Thr	Tyr	Thr	Asp	Arg	Arg	Trp	Cys		
2050						2055						2060					
Phe	Asp	Gly	Thr	Thr	Asn	Asn	Thr	Ile	Met	Glu	Asp	Ser	Val	Pro	Ala		
2065						2070						2075					
Glu	Val	Trp	Thr	Arg	His	Gly	Glu	Lys	Arg	Val	Leu	Lys	Pro	Arg	Trp		
2085						2090						2095					
Met	Asp	Ala	Arg	Val	Cys	Ser	Asp	His	Ala	Ala	Leu	Lys	Ser	Phe	Lys		
2100						2105						2110					

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Glu Phe Ala Ala Gly Lys Arg Gly Ala Ala Phe Gly Val Met Glu Ala
 2115 2120 2125
 Leu Gly Thr Leu Pro Gly His Met Thr Glu Arg Phe Gln Glu Ala Ile
 2130 2135 2140
 Asp Asn Leu Ala Val Leu Met Arg Ala Glu Thr Gly Ser Arg Pro Tyr
 2145 2150 2155 2160
 Lys Ala Ala Ala Ala Gln Leu Pro Glu Thr Leu Glu Thr Ile Met Leu
 2165 2170 2175
 Leu Gly Leu Leu Gly Thr Val Ser Leu Gly Ile Phe Phe Val Leu Met
 2180 2185 2190
 Arg Asn Lys Gly Ile Gly Lys Met Gly Phe Gly Met Val Thr Leu Gly
 2195 2200 2205
 Ala Ser Ala Trp Leu Met Trp Leu Ser Glu Ile Glu Pro Ala Arg Ile
 2210 2215 2220
 Ala Cys Val Leu Ile Val Val Phe Leu Leu Leu Val Val Leu Ile Pro
 2225 2230 2235 2240
 Glu Pro Glu Lys Gln Arg Ser Pro Gln Asp Asn Gln Met Ala Ile Ile
 2245 2250 2255
 Ile Met Val Ala Val Gly Leu Leu Gly Leu Ile Thr Ala Asn Glu Leu
 2260 2265 2270
 Gly Trp Leu Glu Arg Thr Lys Ser Asp Leu Ser His Leu Met Gly Arg
 2275 2280 2285
 Arg Glu Glu Gly Ala Thr Ile Gly Phe Ser Met Asp Ile Asp Leu Arg
 2290 2295 2300
 Pro Ala Ser Ala Trp Ala Ile Tyr Ala Ala Leu Thr Thr Phe Ile Thr
 2305 2310 2315 2320
 Pro Ala Val Gln His Ala Val Thr Thr Ser Tyr Asn Asn Tyr Ser Leu
 2325 2330 2335
 Met Ala Met Ala Thr Gln Ala Gly Val Leu Phe Gly Met Gly Lys Gly
 2340 2345 2350
 Met Pro Phe Tyr Ala Trp Asp Phe Gly Val Pro Leu Leu Met Ile Gly
 2355 2360 2365
 Cys Tyr Ser Gln Leu Thr Pro Leu Thr Leu Ile Val Ala Ile Ile Leu
 2370 2375 2380
 Leu Val Ala His Tyr Met Tyr Leu Ile Pro Gly Leu Gln Ala Ala Ala
 2385 2390 2395 2400
 Ala Arg Ala Ala Gln Lys Arg Thr Ala Ala Gly Ile Met Lys Asn Pro
 2405 2410 2415
 Val Val Asp Gly Ile Val Val Thr Asp Ile Asp Thr Met Thr Ile Asp
 2420 2425 2430
 Pro Gln Val Glu Lys Lys Met Gly Gln Val Leu Leu Ile Ala Val Ala
 2435 2440 2445
 Val Ser Ser Ala Ile Leu Ser Arg Thr Ala Trp Gly Trp Gly Glu Ala
 2450 2455 2460
 Gly Ala Leu Ile Thr Ala Ala Thr Ser Thr Leu Trp Glu Gly Ser Pro
 2465 2470 2475 2480
 Asn Lys Tyr Trp Asn Ser Ser Thr Ala Thr Ser Leu Cys Asn Ile Phe
 2485 2490 2495
 Arg Gly Ser Tyr Leu Ala Gly Ala Ser Leu Ile Tyr Thr Val Thr Arg
 2500 2505 2510

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Asn	Ala	Gly	Leu	Val	Lys	Arg	Arg	Gly	Gly	Gly	Thr	Gly	Glu	Thr	Leu
2515							2520					2525			
Gly	Glu	Lys	Trp	Lys	Ala	Arg	Leu	Asn	Gln	Met	Ser	Ala	Leu	Glu	Phe
2530						2535					2540				
Tyr	Ser	Tyr	Lys	Lys	Ser	Gly	Ile	Thr	Glu	Val	Cys	Arg	Glu	Glu	Ala
2545					2550					2555					2560
Arg	Arg	Ala	Leu	Lys	Asp	Gly	Val	Ala	Thr	Gly	Gly	His	Ala	Val	Ser
			2565						2570					2575	
Arg	Gly	Ser	Ala	Lys	Leu	Arg	Trp	Leu	Val	Glu	Arg	Gly	Tyr	Leu	Gln
			2580					2585					2590		
Pro	Tyr	Gly	Lys	Val	Ile	Asp	Leu	Gly	Cys	Gly	Arg	Gly	Gly	Trp	Ser
	2595						2600					2605			
Tyr	Tyr	Ala	Ala	Thr	Ile	Arg	Lys	Val	Gln	Glu	Val	Lys	Gly	Tyr	Thr
2610						2615					2620				
Lys	Gly	Gly	Pro	Gly	His	Glu	Glu	Pro	Met	Leu	Val	Gln	Ser	Tyr	Gly
2625					2630					2635					2640
Trp	Asn	Ile	Val	Arg	Leu	Lys	Ser	Gly	Val	Asp	Val	Phe	His	Met	Ala
			2645						2650					2655	
Ala	Glu	Pro	Cys	Asp	Thr	Leu	Leu	Cys	Asp	Ile	Gly	Glu	Ser	Ser	Ser
			2660					2665					2670		
Ser	Pro	Glu	Val	Glu	Glu	Ala	Arg	Thr	Leu	Arg	Val	Leu	Ser	Met	Val
	2675						2680					2685			
Gly	Asp	Trp	Leu	Glu	Lys	Arg	Pro	Gly	Ala	Phe	Cys	Ile	Lys	Val	Leu
2690					2695						2700				
Cys	Pro	Tyr	Thr	Ser	Thr	Met	Met	Glu	Thr	Leu	Glu	Arg	Leu	Gln	Arg
2705					2710					2715					2720
Arg	Tyr	Gly	Gly	Gly	Leu	Val	Arg	Val	Pro	Leu	Ser	Arg	Asn	Ser	Thr
			2725						2730					2735	
His	Glu	Met	Tyr	Trp	Val	Ser	Gly	Ala	Lys	Ser	Asn	Thr	Ile	Lys	Ser
		2740						2745					2750		
Val	Ser	Thr	Thr	Ser	Gln	Leu	Leu	Leu	Gly	Arg	Met	Asp	Gly	Pro	Arg
	2755						2760					2765			
Arg	Pro	Val	Lys	Tyr	Glu	Glu	Asp	Val	Asn	Leu	Gly	Ser	Gly	Thr	Arg
	2770					2775					2780				
Ala	Val	Val	Ser	Cys	Ala	Glu	Ala	Pro	Asn	Met	Lys	Ile	Ile	Gly	Asn
2785					2790					2795					2800
Arg	Ile	Glu	Arg	Ile	Arg	Ser	Glu	His	Ala	Glu	Thr	Trp	Phe	Phe	Asp
			2805						2810					2815	
Glu	Asn	His	Pro	Tyr	Arg	Thr	Trp	Ala	Tyr	His	Gly	Ser	Tyr	Glu	Ala
		2820						2825					2830		
Pro	Thr	Gln	Gly	Ser	Ala	Ser	Ser	Leu	Ile	Asn	Gly	Val	Val	Arg	Leu
	2835						2840					2845			
Leu	Ser	Lys	Pro	Trp	Asp	Val	Val	Thr	Gly	Val	Thr	Gly	Ile	Ala	Met
	2850					2855					2860				
Thr	Asp	Thr	Thr	Pro	Tyr	Gly	Gln	Gln	Arg	Val	Phe	Lys	Glu	Lys	Val
2865					2870					2875					2880
Asp	Thr	Arg	Val	Pro	Asp	Pro	Gln	Glu	Gly	Thr	Arg	Gln	Val	Met	Ser
			2885					2890					2895		
Met	Val	Ser	Ser	Trp	Leu	Trp	Lys	Glu	Leu	Gly	Lys	His	Lys	Arg	Pro
		2900						2905					2910		
Arg	Val	Cys	Thr	Lys	Glu	Glu	Phe	Ile	Asn	Lys	Val	Arg	Ser	Asn	Ala

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2915	2920	2925
Ala Leu Gly	Ala Ile Phe Glu Glu Glu Lys Glu Trp	Lys Thr Ala Val
2930	2935	2940
Glu Ala Val Asn Asp Pro Arg Phe Trp Ala Leu Val Asp Lys Glu Arg		
2945	2950	2955 2960
Glu His His Leu Arg Gly Glu Cys Gln Ser Cys Val Tyr Asn Met Met		
	2965	2970 2975
Gly Lys Arg Glu Lys Lys Gln Gly Glu Phe Gly Lys Ala Lys Gly Ser		
	2980	2985 2990
Arg Ala Ile Trp Tyr Met Trp Leu Gly Ala Arg Phe Leu Glu Phe Glu		
	2995	3000 3005
Ala Leu Gly Phe Leu Asn Glu Asp His Trp Met Gly Arg Glu Asn Ser		
3010	3015	3020
Gly Gly Gly Val Glu Gly Leu Gly Leu Gln Arg Leu Gly Tyr Val Leu		
3025	3030	3035 3040
Glu Glu Met Ser Arg Ile Pro Gly Gly Arg Met Tyr Ala Asp Asp Thr		
	3045	3050 3055
Ala Gly Trp Asp Thr Arg Ile Ser Arg Phe Asp Leu Glu Asn Glu Ala		
	3060	3065 3070
Leu Ile Thr Asn Gln Met Glu Lys Gly His Arg Ala Leu Ala Leu Ala		
	3075	3080 3085
Ile Ile Lys Tyr Thr Tyr Gln Asn Lys Val Val Lys Val Leu Arg Pro		
3090	3095	3100
Ala Glu Lys Gly Lys Thr Val Met Asp Ile Ile Ser Arg Gln Asp Gln		
3105	3110	3115 3120
Arg Gly Ser Gly Gln Val Val Thr Tyr Ala Leu Asn Thr Phe Thr Asn		
	3125	3130 3135
Leu Val Val Gln Leu Ile Arg Ser Met Glu Ala Glu Glu Val Leu Glu		
	3140	3145 3150
Met Gln Asp Leu Trp Leu Leu Arg Arg Ser Glu Lys Val Thr Asn Trp		
3155	3160	3165
Leu Gln Ser Asn Gly Trp Asp Arg Leu Lys Arg Met Ala Val Ser Gly		
3170	3175	3180
Asp Asp Cys Val Val Arg Pro Ile Asp Asp Arg Phe Ala His Ala Leu		
3185	3190	3195 3200
Arg Phe Leu Asn Asp Met Gly Lys Val Arg Lys Asp Thr Gln Glu Trp		
	3205	3210 3215
Lys Pro Ser Thr Gly Trp Asp Asn Trp Glu Glu Val Pro Phe Cys Ser		
	3220	3225 3230
His His Phe Asn Lys Leu His Leu Lys Asp Gly Arg Ser Ile Val Val		
	3235	3240 3245
Pro Cys Arg His Gln Asp Glu Leu Ile Gly Arg Ala Arg Val Ser Pro		
3250	3255	3260
Gly Ala Gly Trp Ser Ile Arg Glu Thr Ala Cys Leu Ala Lys Ser Tyr		
3265	3270	3275 3280
Ala Gln Met Trp Gln Leu Leu Tyr Phe His Arg Arg Asp Leu Arg Leu		
	3285	3290 3295
Met Ala Asn Ala Ile Cys Ser Ser Val Pro Val Asp Trp Val Pro Thr		
	3300	3305 3310
Gly Arg Thr Thr Trp Ser Ile His Gly Lys Gly Glu Trp Met Thr Thr		
	3315	3320 3325

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Glu Asp Met Leu Val Val Trp Asn Arg Val Trp Ile Glu Glu Asn Asp
 3330 3335 3340

His Met Glu Asp Lys Thr Pro Val Thr Lys Trp Thr Asp Ile Pro Tyr
 3345 3350 3355 3360

Leu Gly Lys Arg Glu Asp Leu Trp Cys Gly Ser Leu Ile Gly His Arg
 3365 3370 3375

Pro Arg Thr Thr Trp Ala Glu Asn Ile Lys Asn Thr Val Asn Met Val
 3380 3385 3390

Arg Arg Ile Ile Gly Asp Glu Glu Lys Tyr Met Asp Tyr Leu Ser Thr
 3395 3400 3405

Gln Val Arg Tyr Leu Gly Glu Glu Gly Ser Thr Pro Gly Val Leu
 3410 3415 3420

<210> SEQ ID NO 9
 <211> LENGTH: 3423
 <212> TYPE: PRT
 <213> ORGANISM: Zika virus

<400> SEQUENCE: 9

Met Lys Asn Pro Lys Lys Lys Ser Gly Gly Phe Arg Ile Val Asn Met
 1 5 10 15

Leu Lys Arg Gly Val Ala Arg Val Ser Pro Phe Gly Gly Leu Lys Arg
 20 25 30

Leu Pro Ala Gly Leu Leu Leu Gly His Gly Pro Ile Arg Met Val Leu
 35 40 45

Ala Ile Leu Ala Phe Leu Arg Phe Thr Ala Ile Lys Pro Ser Leu Gly
 50 55 60

Leu Ile Asn Arg Trp Gly Ser Val Gly Lys Lys Glu Ala Met Glu Ile
 65 70 75 80

Ile Lys Lys Phe Lys Lys Asp Leu Ala Ala Met Leu Arg Ile Ile Asn
 85 90 95

Ala Arg Lys Glu Lys Lys Arg Arg Gly Ala Asp Thr Ser Val Gly Ile
 100 105 110

Val Gly Leu Leu Leu Thr Thr Ala Met Ala Ala Glu Val Thr Arg Arg
 115 120 125

Gly Ser Ala Tyr Tyr Met Tyr Leu Asp Arg Asn Asp Ala Gly Glu Ala
 130 135 140

Ile Ser Phe Pro Thr Thr Leu Gly Met Asn Lys Cys Tyr Ile Gln Ile
 145 150 155 160

Met Asp Leu Gly His Met Cys Asp Ala Thr Met Ser Tyr Glu Cys Pro
 165 170 175

Met Leu Asp Glu Gly Val Glu Pro Asp Asp Val Asp Cys Trp Cys Asn
 180 185 190

Thr Thr Ser Thr Trp Val Val Tyr Gly Thr Cys His His Lys Lys Gly
 195 200 205

Glu Ala Arg Arg Ser Arg Arg Ala Val Thr Leu Pro Ser His Ser Thr
 210 215 220

Arg Lys Leu Gln Thr Arg Ser Gln Thr Trp Leu Glu Ser Arg Glu Tyr
 225 230 235 240

Thr Lys His Leu Ile Arg Val Glu Asn Trp Ile Phe Arg Asn Pro Gly
 245 250 255

Phe Ala Leu Ala Ala Ala Ala Ile Ala Trp Leu Leu Gly Ser Ser Thr

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260						265						270					
Ser	Gln	Lys	Val	Ile	Tyr	Leu	Val	Met	Ile	Leu	Leu	Ile	Ala	Pro	Ala		
275						280						285					
Tyr	Ser	Ile	Arg	Cys	Ile	Gly	Val	Ser	Asn	Arg	Asp	Phe	Val	Glu	Gly		
290						295						300					
Met	Ser	Gly	Gly	Thr	Trp	Val	Asp	Val	Val	Leu	Glu	His	Gly	Gly	Cys		
305						310						315					
Val	Thr	Val	Met	Ala	Gln	Asp	Lys	Pro	Thr	Val	Asp	Ile	Glu	Leu	Val		
325						330						335					
Thr	Thr	Thr	Val	Ser	Asn	Met	Ala	Glu	Val	Arg	Ser	Tyr	Cys	Tyr	Glu		
340						345						350					
Ala	Ser	Ile	Ser	Asp	Met	Ala	Ser	Asp	Ser	Arg	Cys	Pro	Thr	Gln	Gly		
355						360						365					
Glu	Ala	Tyr	Leu	Asp	Lys	Gln	Ser	Asp	Thr	Gln	Tyr	Val	Cys	Lys	Arg		
370						375						380					
Thr	Leu	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys	Gly	Leu	Phe	Gly	Lys		
385						390						395					
Gly	Ser	Leu	Val	Thr	Cys	Ala	Lys	Phe	Ala	Cys	Ser	Lys	Lys	Met	Thr		
405						410						415					
Gly	Lys	Ser	Ile	Gln	Pro	Glu	Asn	Leu	Glu	Tyr	Arg	Ile	Met	Leu	Ser		
420						425						430					
Val	His	Gly	Ser	Gln	His	Ser	Gly	Met	Ile	Val	Asn	Asp	Thr	Gly	His		
435						440						445					
Glu	Thr	Asp	Glu	Asn	Arg	Ala	Lys	Val	Glu	Ile	Thr	Pro	Asn	Ser	Pro		
450						455						460					
Arg	Ala	Glu	Ala	Thr	Leu	Gly	Gly	Phe	Gly	Ser	Leu	Gly	Leu	Asp	Cys		
465						470						475					
Glu	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Ser	Asp	Leu	Tyr	Tyr	Leu	Thr	Met		
485						490						495					
Asn	Asn	Lys	His	Trp	Leu	Val	His	Lys	Glu	Trp	Phe	His	Asp	Ile	Pro		
500						505						510					
Leu	Pro	Trp	His	Ala	Gly	Ala	Asp	Thr	Gly	Thr	Pro	His	Trp	Asn	Asn		
515						520						525					
Lys	Glu	Ala	Leu	Val	Glu	Phe	Lys	Asp	Ala	His	Ala	Lys	Arg	Gln	Thr		
530						535						540					
Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Val	His	Thr	Ala	Leu	Ala		
545						550						555					
Gly	Ala	Leu	Glu	Ala	Glu	Met	Asp	Gly	Ala	Lys	Gly	Arg	Leu	Ser	Ser		
565						570						575					
Gly	His	Leu	Lys	Cys	Arg	Leu	Lys	Met	Asp	Lys	Leu	Arg	Leu	Lys	Gly		
580						585						590					
Val	Ser	Tyr	Ser	Leu	Cys	Thr	Ala	Ala	Phe	Thr	Phe	Thr	Lys	Ile	Pro		
595						600						605					
Ala	Glu	Thr	Leu	His	Gly	Thr	Val	Thr	Val	Glu	Val	Gln	Tyr	Ala	Gly		
610						615						620					
Thr	Asp	Gly	Pro	Cys	Lys	Val	Pro	Ala	Gln	Met	Ala	Val	Asp	Met	Gln		
625						630						635					
Thr	Leu	Thr	Pro	Val	Gly	Arg	Leu	Ile	Thr	Ala	Asn	Pro	Val	Ile	Thr		
645						650						655					
Glu	Ser	Thr	Glu	Asn	Ser	Lys	Met	Met	Leu	Glu	Leu	Asp	Pro	Pro	Phe		
660						665						670					

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Gly	Asp	Ser	Tyr	Ile	Val	Ile	Gly	Val	Gly	Glu	Lys	Lys	Ile	Thr	His
	675						680					685			
His	Trp	His	Arg	Ser	Gly	Ser	Thr	Ile	Gly	Lys	Ala	Phe	Glu	Ala	Thr
	690					695					700				
Val	Arg	Gly	Ala	Lys	Arg	Met	Ala	Val	Leu	Gly	Asp	Thr	Ala	Trp	Asp
705					710					715					720
Phe	Gly	Ser	Val	Gly	Gly	Ala	Leu	Asn	Ser	Leu	Gly	Lys	Gly	Ile	His
				725				730						735	
Gln	Ile	Phe	Gly	Ala	Ala	Phe	Lys	Ser	Leu	Phe	Gly	Gly	Met	Ser	Trp
			740					745					750		
Phe	Ser	Gln	Ile	Leu	Ile	Gly	Thr	Leu	Leu	Met	Trp	Leu	Gly	Leu	Asn
		755					760					765			
Thr	Lys	Asn	Gly	Ser	Ile	Ser	Leu	Met	Cys	Leu	Ala	Leu	Gly	Gly	Val
	770					775					780				
Leu	Ile	Phe	Leu	Ser	Thr	Ala	Val	Ser	Ala	Asp	Val	Gly	Cys	Ser	Val
785					790					795					800
Asp	Phe	Ser	Lys	Lys	Glu	Thr	Arg	Cys	Gly	Thr	Gly	Val	Phe	Val	Tyr
			805						810					815	
Asn	Asp	Val	Glu	Ala	Trp	Arg	Asp	Arg	Tyr	Lys	Tyr	His	Pro	Asp	Ser
			820					825					830		
Pro	Arg	Arg	Leu	Ala	Ala	Ala	Val	Lys	Gln	Ala	Trp	Glu	Asp	Gly	Ile
		835					840					845			
Cys	Gly	Ile	Ser	Ser	Val	Ser	Arg	Met	Glu	Asn	Ile	Met	Trp	Arg	Ser
	850					855					860				
Val	Glu	Gly	Glu	Leu	Asn	Ala	Ile	Leu	Glu	Glu	Asn	Gly	Val	Gln	Leu
865					870					875					880
Thr	Val	Val	Val	Gly	Ser	Val	Lys	Asn	Pro	Met	Trp	Arg	Gly	Pro	Gln
				885					890					895	
Arg	Leu	Pro	Val	Pro	Val	Asn	Glu	Leu	Pro	His	Gly	Trp	Lys	Ala	Trp
			900					905					910		
Gly	Lys	Ser	Tyr	Phe	Val	Arg	Ala	Ala	Lys	Thr	Asn	Asn	Ser	Phe	Val
		915					920					925			
Val	Asp	Gly	Asp	Thr	Leu	Lys	Glu	Cys	Pro	Leu	Lys	His	Arg	Ala	Trp
	930					935					940				
Asn	Ser	Phe	Leu	Val	Glu	Asp	His	Gly	Phe	Gly	Val	Phe	His	Thr	Ser
945				950						955					960
Val	Trp	Leu	Lys	Val	Arg	Glu	Asp	Tyr	Ser	Leu	Glu	Cys	Asp	Pro	Ala
			965					970						975	
Val	Ile	Gly	Thr	Ala	Val	Lys	Gly	Lys	Glu	Ala	Val	His	Ser	Asp	Leu
		980						985					990		
Gly	Tyr	Trp	Ile	Glu	Ser	Glu	Lys	Asn	Asp	Thr	Trp	Arg	Leu	Lys	Arg
	995						1000					1005			
Ala	His	Leu	Ile	Glu	Met	Lys	Thr	Cys	Glu	Trp	Pro	Lys	Ser	His	Thr
	1010					1015					1020				
Leu	Trp	Thr	Asp	Gly	Ile	Glu	Glu	Ser	Asp	Leu	Ile	Ile	Pro	Lys	Ser
1025					1030					1035					1040
Leu	Ala	Gly	Pro	Leu	Ser	His	His	Asn	Thr	Arg	Glu	Gly	Tyr	Arg	Thr
			1045						1050					1055	
Gln	Met	Lys	Gly	Pro	Trp	His	Ser	Glu	Glu	Leu	Glu	Ile	Arg	Phe	Glu
		1060						1065					1070		

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Glu Cys Pro Gly Thr Lys Val His Val Glu Glu Thr Cys Gly Thr Arg	1075	1080	1085
Gly Pro Ser Leu Arg Ser Thr Thr Ala Ser Gly Arg Val Ile Glu Glu	1090	1095	1100
Trp Cys Cys Arg Glu Cys Thr Met Pro Pro Leu Ser Phe Arg Ala Lys	1105	1110	1115
Asp Gly Cys Trp Tyr Gly Met Glu Ile Arg Pro Arg Lys Glu Pro Glu	1125	1130	1135
Ser Asn Leu Val Arg Ser Met Val Thr Ala Gly Ser Thr Asp His Met	1140	1145	1150
Asp His Phe Ser Leu Gly Val Leu Val Ile Leu Leu Met Val Gln Glu	1155	1160	1165
Gly Leu Lys Lys Arg Met Thr Thr Lys Ile Ile Ile Ser Thr Ser Met	1170	1175	1180
Ala Val Leu Val Ala Met Ile Leu Gly Gly Phe Ser Met Ser Asp Leu	1185	1190	1195
Ala Lys Leu Ala Ile Leu Met Gly Ala Thr Phe Ala Glu Met Asn Thr	1205	1210	1215
Gly Gly Asp Val Ala His Leu Ala Leu Ile Ala Ala Phe Lys Val Arg	1220	1225	1230
Pro Ala Leu Leu Val Ser Phe Ile Phe Arg Ala Asn Trp Thr Pro Arg	1235	1240	1245
Glu Ser Met Leu Leu Ala Leu Ala Ser Cys Leu Leu Gln Thr Ala Ile	1250	1255	1260
Ser Ala Leu Glu Gly Asp Leu Met Val Leu Ile Asn Gly Phe Ala Leu	1265	1270	1275
Ala Trp Leu Ala Ile Arg Ala Met Val Val Pro Arg Thr Asp Asn Ile	1285	1290	1295
Thr Leu Ala Ile Leu Ala Ala Leu Thr Pro Leu Ala Arg Gly Thr Leu	1300	1305	1310
Leu Val Ala Trp Arg Ala Gly Leu Ala Thr Cys Gly Gly Phe Met Leu	1315	1320	1325
Leu Ser Leu Lys Gly Lys Gly Ser Val Lys Lys Asn Leu Pro Phe Val	1330	1335	1340
Met Ala Leu Gly Leu Thr Ala Val Arg Leu Val Asp Pro Ile Asn Val	1345	1350	1355
Val Gly Leu Leu Leu Leu Thr Arg Ser Gly Lys Arg Ser Trp Pro Pro	1365	1370	1375
Ser Glu Val Leu Thr Ala Val Gly Leu Ile Cys Ala Leu Ala Gly Gly	1380	1385	1390
Phe Ala Lys Ala Asp Ile Glu Met Ala Gly Pro Met Ala Ala Val Gly	1395	1400	1405
Leu Leu Ile Val Ser Tyr Val Val Ser Gly Lys Ser Val Asp Met Tyr	1410	1415	1420
Ile Glu Arg Ala Gly Asp Ile Thr Trp Glu Lys Asp Ala Glu Val Thr	1425	1430	1435
Gly Asn Ser Pro Arg Leu Asp Val Ala Leu Asp Glu Ser Gly Asp Phe	1445	1450	1455
Ser Leu Val Glu Asp Asp Gly Pro Pro Met Arg Glu Ile Ile Leu Lys	1460	1465	1470
Val Val Leu Met Thr Ile Cys Gly Met Asn Pro Ile Ala Ile Pro Phe			

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1475	1480	1485
Ala Ala Gly Ala Trp Tyr Val Tyr Val Lys Thr Gly Lys Arg Ser Gly 1490 1495 1500		
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Thr Asp Gly Val Tyr Arg Val Met Thr Arg Arg Leu Leu Gly Ser Thr 1525 1530 1535		
Gln Val Gly Val Gly Val Met Gln Glu Gly Val Phe His Thr Met Trp 1540 1545 1550		
His Val Thr Lys Gly Ser Ala Leu Arg Ser Gly Glu Gly Arg Leu Asp 1555 1560 1565		
Pro Tyr Trp Gly Asp Val Lys Gln Asp Leu Val Ser Tyr Cys Gly Pro 1570 1575 1580		
Trp Lys Leu Asp Ala Ala Trp Asp Gly His Ser Glu Val Gln Leu Leu 1585 1590 1595 1600		
Ala Val Pro Pro Gly Glu Arg Ala Arg Asn Ile Gln Thr Leu Pro Gly 1605 1610 1615		
Ile Phe Lys Thr Lys Asp Gly Asp Ile Gly Ala Val Ala Leu Asp Tyr 1620 1625 1630		
Pro Ala Gly Thr Ser Gly Ser Pro Ile Leu Asp Lys Cys Gly Arg Val 1635 1640 1645		
Ile Gly Leu Tyr Gly Asn Gly Val Val Ile Lys Asn Gly Ser Tyr Val 1650 1655 1660		
Ser Ala Ile Thr Gln Gly Arg Arg Glu Glu Glu Thr Pro Val Glu Cys 1665 1670 1675 1680		
Phe Glu Pro Ser Met Leu Lys Lys Lys Gln Leu Thr Val Leu Asp Leu 1685 1690 1695		
His Pro Gly Ala Gly Lys Thr Arg Arg Val Leu Pro Glu Ile Val Arg 1700 1705 1710		
Glu Ala Ile Lys Thr Arg Leu Arg Thr Val Ile Leu Ala Pro Thr Arg 1715 1720 1725		
Val Val Ala Ala Glu Met Glu Glu Ala Leu Arg Gly Leu Pro Val Arg 1730 1735 1740		
Tyr Met Thr Thr Ala Val Asn Val Thr His Ser Gly Thr Glu Ile Val 1745 1750 1755 1760		
Asp Leu Met Cys His Ala Thr Phe Thr Ser Arg Leu Leu Gln Pro Ile 1765 1770 1775		
Arg Val Pro Asn Tyr Asn Leu Tyr Ile Met Asp Glu Ala His Phe Thr 1780 1785 1790		
Asp Pro Ser Ser Ile Ala Ala Arg Gly Tyr Ile Ser Thr Arg Val Glu 1795 1800 1805		
Met Gly Glu Ala Ala Ala Ile Phe Met Thr Ala Thr Pro Pro Gly Thr 1810 1815 1820		
Arg Asp Ala Phe Pro Asp Ser Asn Ser Pro Ile Met Asp Thr Glu Val 1825 1830 1835 1840		
Glu Val Pro Glu Arg Ala Trp Ser Ser Gly Phe Asp Trp Val Thr Asp 1845 1850 1855		
His Ser Gly Lys Thr Val Trp Phe Val Pro Ser Val Arg Asn Gly Asn 1860 1865 1870		
Glu Ile Ala Ala Cys Leu Thr Lys Ala Gly Lys Arg Val Ile Gln Leu 1875 1880 1885		

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Ser Arg Lys Thr Phe Glu Thr Glu Phe Gln Lys Thr Lys His Gln Glu			
1890	1895	1900	
Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly Ala Asn Phe			
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Lys Ala Asp Arg Val Ile Asp Ser Arg Arg Cys Leu Lys Pro Val Ile			
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Leu Asp Gly Glu Arg Val Ile Leu Ala Gly Pro Met Pro Val Thr His			
	1940	1945	1950
Ala Ser Ala Ala Gln Arg Arg Gly Arg Ile Gly Arg Asn Pro Asn Lys			
	1955	1960	1965
Pro Gly Asp Glu Tyr Leu Tyr Gly Gly Gly Cys Ala Glu Thr Asp Glu			
1970	1975	1980	
Asp His Ala His Trp Leu Glu Ala Arg Met Leu Leu Asp Asn Ile Tyr			
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Leu Gln Asp Gly Leu Ile Ala Ser Leu Tyr Arg Pro Glu Ala Asp Lys			
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Val Ala Ala Ile Glu Gly Glu Phe Lys Leu Arg Thr Glu Gln Arg Lys			
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Thr Phe Val Glu Leu Met Lys Arg Gly Asp Leu Pro Val Trp Leu Ala			
	2035	2040	2045
Tyr Gln Val Ala Ser Ala Gly Ile Thr Tyr Thr Asp Arg Arg Trp Cys			
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Phe Asp Gly Thr Thr Asn Asn Thr Ile Met Glu Asp Ser Val Pro Ala			
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Glu Val Trp Thr Arg His Gly Glu Lys Arg Val Leu Lys Pro Arg Trp			
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Met Asp Ala Arg Val Cys Ser Asp His Ala Ala Leu Lys Ser Phe Lys			
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Glu Phe Ala Ala Gly Lys Arg Gly Ala Ala Phe Gly Val Met Glu Ala			
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Leu Gly Thr Leu Pro Gly His Met Thr Glu Arg Phe Gln Glu Ala Ile			
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Asp Asn Leu Ala Val Leu Met Arg Ala Glu Thr Gly Ser Arg Pro Tyr			
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Lys Ala Ala Ala Ala Gln Leu Pro Glu Thr Leu Glu Thr Ile Met Leu			
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Leu Gly Leu Leu Gly Thr Val Ser Leu Gly Ile Phe Phe Val Leu Met			
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Arg Asn Lys Gly Ile Gly Lys Met Gly Phe Gly Met Val Thr Leu Gly			
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Ala Ser Ala Trp Leu Met Trp Leu Ser Glu Ile Glu Pro Ala Arg Ile			
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Ala Cys Val Leu Ile Val Val Phe Leu Leu Leu Val Val Leu Ile Pro			
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Glu Pro Glu Lys Gln Arg Ser Pro Gln Asp Asn Gln Met Ala Ile Ile			
	2245	2250	2255
Ile Met Val Ala Val Gly Leu Leu Gly Leu Ile Thr Ala Asn Glu Leu			
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Gly Trp Leu Glu Arg Thr Lys Ser Asp Leu Ser His Leu Met Gly Arg			
	2275	2280	2285

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2370					2375						2380				
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2385					2390					2395					2400
Ala	Arg	Ala	Ala	Gln	Lys	Arg	Thr	Ala	Ala	Gly	Ile	Met	Lys	Asn	Pro
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Val	Ser	Ser	Ala	Ile	Leu	Ser	Arg	Thr	Ala	Trp	Gly	Trp	Gly	Glu	Ala
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Gly	Ala	Leu	Ile	Thr	Ala	Ala	Thr	Ser	Thr	Leu	Trp	Glu	Gly	Ser	Pro
2465				2470						2475					2480
Asn	Lys	Tyr	Trp	Asn	Ser	Ser	Thr	Ala	Thr	Ser	Leu	Cys	Asn	Ile	Phe
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Pro	Tyr	Gly	Lys	Val	Ile	Asp	Leu	Gly	Cys	Gly	Arg	Gly	Gly	Trp	Ser
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Tyr	Tyr	Ala	Ala	Thr	Ile	Arg	Lys	Val	Gln	Glu	Val	Lys	Gly	Tyr	Thr
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Lys	Gly	Gly	Pro	Gly	His	Glu	Glu	Pro	Met	Leu	Val	Gln	Ser	Tyr	Gly
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Trp	Asn	Ile	Val	Arg	Leu	Lys	Ser	Gly	Val	Asp	Val	Phe	His	Met	Ala
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Ala	Glu	Pro	Cys	Asp	Thr	Leu	Leu	Cys	Asp	Ile	Gly	Glu	Ser	Ser	Ser
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Ser	Pro	Glu	Val	Glu	Glu	Ala	Arg	Thr	Leu	Arg	Val	Leu	Ser	Met	Val
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Gly	Asp	Trp	Leu	Glu	Lys	Arg	Pro	Gly	Ala	Phe	Cys	Ile	Lys	Val	Leu

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His Glu Met Tyr Trp Val Ser Gly Ala Lys Ser Asn Thr Ile Lys Ser		
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Val Ser Thr Thr Ser Gln Leu Leu Leu Gly Arg Met Asp Gly Pro Arg		
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Ala Val Val Ser Cys Ala Glu Ala Pro Asn Met Lys Ile Ile Gly Asn		
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Glu Asn His Pro Tyr Arg Thr Trp Ala Tyr His Gly Ser Tyr Glu Ala		
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Pro Thr Gln Gly Ser Ala Ser Ser Leu Ile Asn Gly Val Val Arg Leu		
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Ala Gly Trp Asp Thr Arg Ile Ser Arg Phe Asp Leu Glu Asn Glu Ala		
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Leu Ile Thr Asn Gln Met Glu Lys Gly His Arg Ala Leu Ala Leu Ala		
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Ile Ile Lys Tyr Thr Tyr Gln Asn Lys Val Val Lys Val Leu Arg Pro		
	3090	3095 3100

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 Leu Val Val Gln Leu Ile Arg Asn Met Glu Ala Glu Glu Val Leu Glu
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 Met Gln Asp Leu Trp Leu Leu Arg Arg Ser Glu Lys Val Thr Asn Trp
 3155 3160 3165
 Leu Gln Ser Asn Gly Trp Asp Arg Leu Lys Arg Met Ala Val Ser Gly
 3170 3175 3180
 Asp Asp Cys Val Val Lys Pro Ile Asp Asp Arg Phe Ala His Ala Leu
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 3205 3210 3215
 Lys Pro Ser Thr Gly Trp Asp Asn Trp Glu Glu Val Pro Phe Cys Ser
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 His His Phe Asn Lys Leu His Leu Lys Asp Gly Arg Ser Ile Val Val
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 3315 3320 3325
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 3330 3335 3340
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 3345 3350 3355 3360
 Leu Gly Lys Arg Glu Asp Leu Trp Cys Gly Ser Leu Ile Gly His Arg
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 <213> ORGANISM: Zika virus

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 <212> TYPE: DNA

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<213> ORGANISM: Zika virus

<400> SEQUENCE: 11

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What is claimed is:

1. A recombinant nucleic acid vector comprising a heterologous promoter operably linked to a nucleotide sequence encoding flavivirus prM/E, which vector lacks nucleic acid sequences encoding one or more of flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks nucleic acid sequences encoding functional flavivirus capsid.

2. The recombinant vector of claim 1 wherein the heterologous promoter is a heterologous viral promoter. The recombinant vector of claim 1 which includes a portion of flavivirus capsid sequences.

4. The recombinant vector of claim 1 wherein the capsid sequence includes amino acids 98 to 112 of the capsid protein encoded by SEQ ID NO:1 or a protein having at least 80% amino acid sequence identity thereto.

5. The recombinant vector of claim 1 wherein the flavivirus is a Zika virus.

6. The recombinant vector of claim 1 wherein the prM/E sequences have at least 80% amino acid sequence identity to the prM/E sequences encoded by any one of SEQ ID Nos. 1-3 or 5.

7. The recombinant vector of claim 1 wherein the prWE sequences are operably linked to a heterologous secretion signal.

8. The recombinant vector of claim 7 wherein the heterologous secretion signal is a TPA, IL-2, IgG kappa light chain, CD33, or Oikosin secretion signal.

9. A vaccine comprising an effective amount of a flavivirus like particle comprising a lipid bilayer comprising flavivirus prM/E but which particle lacks one or more of flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks functional flavivirus capsid.

10. The vaccine of claim 9 further comprising one or more adjuvants.

11. The vaccine of claim 10 wherein the adjuvant comprises alum, monophosphoryl lipid A (MPLA), squalene, aluminum hydroxide adsorbed TLR4 agonist, dimethyldioctadecylammonium, tripalmitoyl-S-glyceryl cysteine, trehalose dibehenate, saponin, MF59, AS03, virosomes, AS04, CpG, imidazoquinoline, poly I:C, flagellin, or any combination thereof

tadecylammonium, tripalmitoyl-S-glyceryl cysteine, trehalose dibehenate, saponin, MF59, AS03, virosomes, AS04, CpG, imidazoquinoline, poly I:C, flagellin, or any combination thereof

12. The vaccine of claim 9 wherein the flavivirus is a Zika virus.

13. The vaccine of claim 9 wherein the prM/E sequences have at least 80% amino acid sequence identity to the prM/E sequences encoded by any one of SEQ ID Nos. 1-3 or 5.

14. A method to prevent, inhibit or treat flavivirus infection in a mammal, comprising: administering to the mammal a composition comprising an effective amount of a flavivirus like particle comprising a lipid bilayer comprising flavivirus prM/E but which particle lacks one or more of flavivirus NS1, NS2A, NS2B, NS3, NS4A, NS4B or NS5 and optionally lacks functional flavivirus capsid, or a composition comprising an effective amount of anti-flavivirus antibodies.

13. The method of claim 14 wherein the mammal is a female mammal.

14. The method of claim 14 wherein the mammal is a human.

15. The method of claim 14 wherein the flavivirus is a Zika virus.

16. The method of claim 17 wherein the prM/E sequences have at least 80% amino acid sequence identity to the prM/E sequences encoded by any one of SEQ ID Nos. 1-3 or 5.

17. The method of claim 14 wherein the composition comprising the flavivirus like particle is administered intramuscularly, subcutaneously or intranasally.

18. The method of claim 14 wherein the composition inhibits flavivirus infection.

19. The method of claim 14 wherein the composition treats flavivirus infection.

20. The method of claim 14 wherein the composition comprising antibodies comprises antibodies pooled from multiple donors that were infected with the flavivirus.

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