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Brunkard

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(54) **METHOD FOR IMPROVING PLANT GROWTH WITH A TRNA SYNTHETASE GENE THAT ACTIVATES TOR**

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(72) Inventor: **Jacob Brunkard, Madison, WI (US)**

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(57) **ABSTRACT**

The present invention provides compositions and methods for increasing the growth, growth rate, and/or yield of a plant that lacks root nodules by engineering it to overexpress the protein asparaginyl-tRNA synthetase 1 (NARS1). Specifically, the present invention provides plants that are engineered to overexpress NARS1, seeds produced by said plants, and methods of generating and growing said plants.

Specification includes a Sequence Listing.

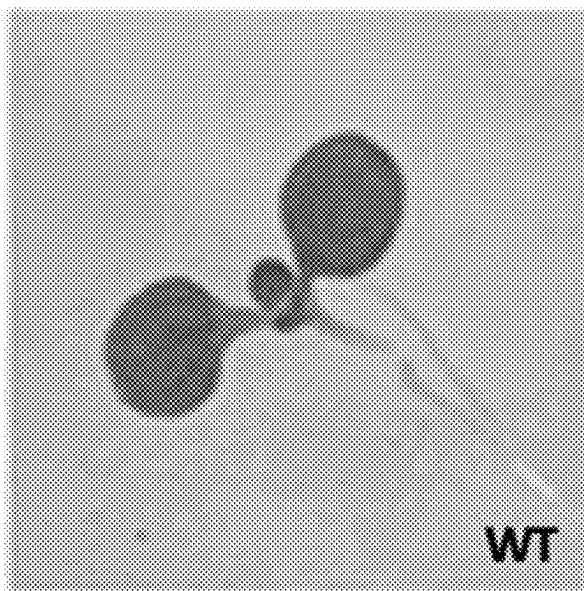
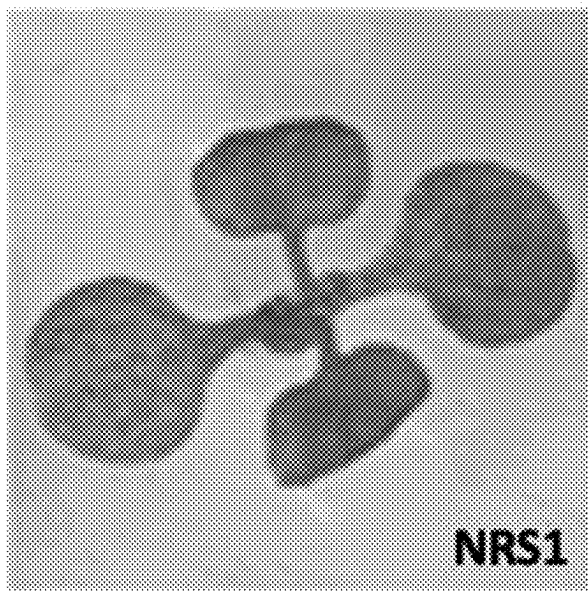


FIG. 1A

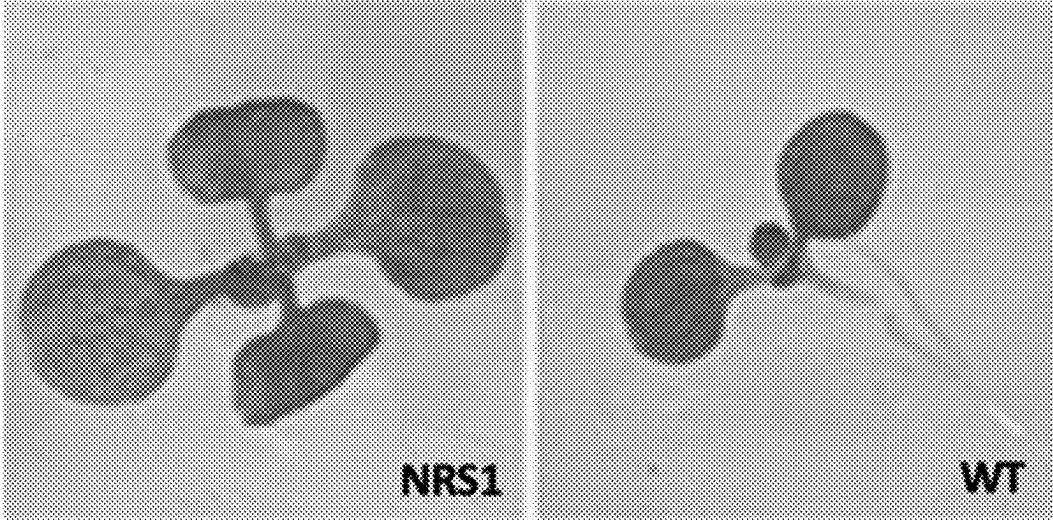


FIG. 1B

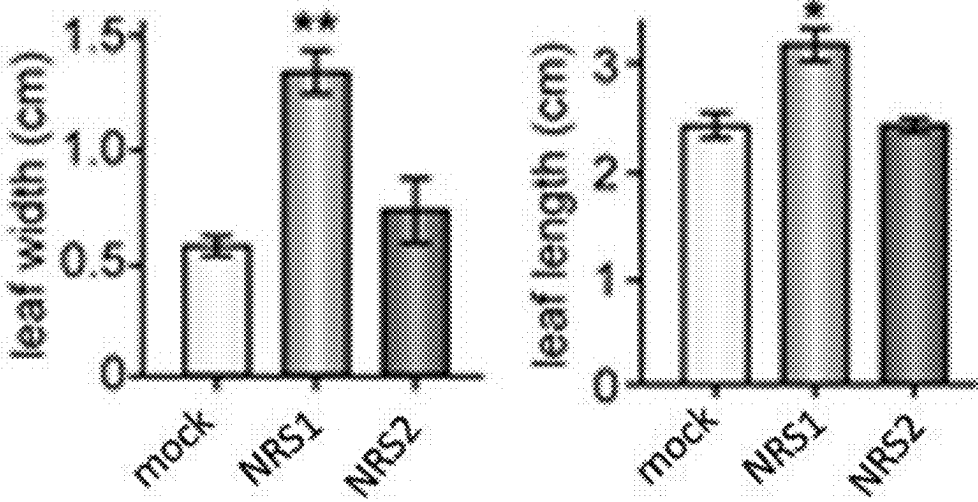


FIG. 2

Col-0



35S:PRO: NRS1: pEG1



FIG. 3

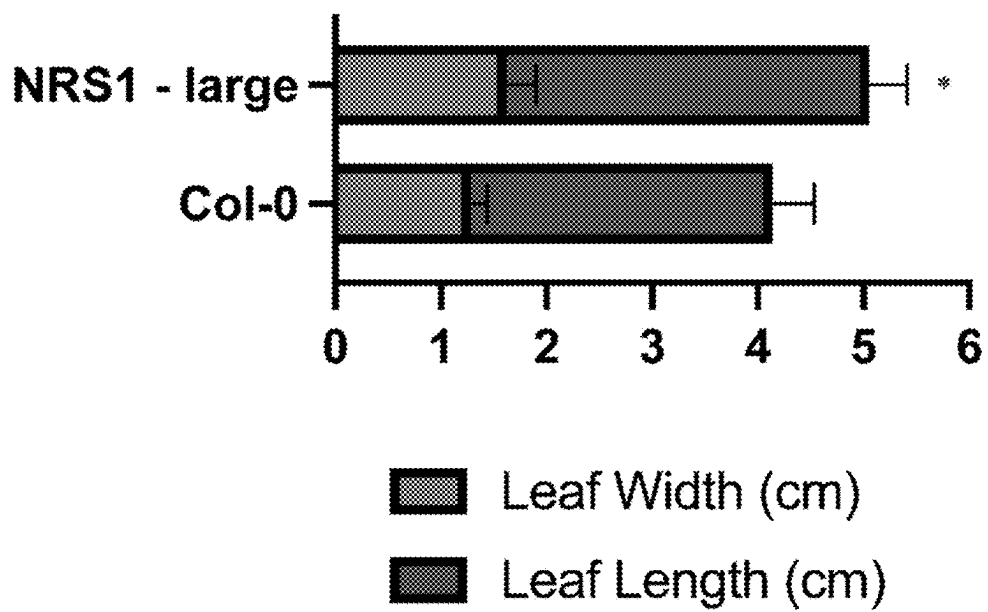


FIG. 4 (continued)

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FIG. 4 (continued)

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FIG. 5



ZmUBQ1pro:NARS1



Wild-type siblings

METHOD FOR IMPROVING PLANT GROWTH WITH A TRNA SYNTHETASE GENE THAT ACTIVATES TOR

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to U.S. Provisional Application No. 63/510,280, filed Jun. 26, 2023, the contents of which are incorporated by reference in their entireties.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH

[0002] This invention was made with government support under grant numbers OD023072 and GM145814 awarded by the National Institutes of Health. The government has certain rights in the invention.

SEQUENCE LISTING STATEMENT

[0003] This application includes a sequence listing in XML format titled “960296.04519_ST26.xml”, which is 43,231 bytes in size and was created on Jun. 24, 2024. The sequence listing is electronically submitted with this application via Patent Center and is incorporated herein by reference in its entirety.

BACKGROUND

[0004] Plants are an essential resource. We rely on them for food, water, medicine, clean air, habitat, shelter, and fuel. Climate change is expected to have a serious negative impact on our ability to grow plants. Genetic methods to improve crop plant growth rates and yields are urgently needed as demands for agricultural products (bioenergy/biomass, food, feedstock, etc.) increase while fertilizer resources and available arable land decrease. Accordingly, there is a need in the art for more efficient methods of growing plants.

SUMMARY

[0005] The present invention provides compositions and methods for increasing the growth, growth rate, and/or yield of a plant that lacks root nodules by engineering it to overexpress the protein asparaginyl-tRNA synthetase 1 (NARS1).

[0006] In a first aspect, the present invention provides plants that lack root nodules and are engineered to overexpress a NARS1 protein.

[0007] In a second aspect, the present invention provides seeds produced by the plants described herein.

[0008] In a third aspect, the present invention provides methods of generating a plant that overexpresses NARS1. The methods comprise: (a) introducing a construct comprising a heterologous promoter operably linked to a polynucleotide encoding a NARS1 protein into a plant cell; and (b) growing the plant cell into a plant.

[0009] In a fourth aspect, the present invention provides methods of growing a plant that overexpresses NARS1. The methods comprise (a) planting a seed described herein; and (b) growing the seed into a plant.

BRIEF DESCRIPTION OF THE DRAWINGS

[0010] FIGS. 1A-1B show a comparison of transgenic *Arabidopsis thaliana* plants that overexpress NARS1 to wild-type control plants after 7 days of growth. FIG. 1A shows representative images of the transgenic *Arabidopsis thaliana* seedlings (left) and the wild-type seedlings (right). FIG. 1B is two bar graphs showing the leaf width (left) and leaf length (right) of leaf 6 of wild-type control plants (“mock”), plants that overexpress NARS1 (“NRS1”), and plants that overexpress NARS2 (“NRS2”) after 7 days of growth. Leaf width and length were compared via t test (** p<10-5; * p<10-2; n≥24).

[0011] FIG. 2 shows a comparison of transgenic *Arabidopsis thaliana* plants that overexpress NARS1 to wild-type control plants after 4 weeks of growth. Representative images of Col-0 wild-type plants (top) and plants transformed with 35S:PRO:NRS1:pEG103 (bottom) are shown.

[0012] FIG. 3 is a bar graph comparing the leaf width and length of transgenic *Arabidopsis thaliana* plants that overexpress NARS1 to that of wild-type (Col-0) control plants. The length and width of leaf 6 was measured and compared via t test (** p<10-5; * p<10-2; n≥24).

[0013] FIG. 4 is an alignment of NARS1 protein sequences from 18 different plant species. The identity of each sequence is detailed in Table 1, below.

TABLE 1

NARS1 protein sequences aligned in FIG. 4			
Row in Alignment	SEQ ID NO:	Organism	Reference Genome
1	2	Query sequence	NR
2	2	<i>Arabidopsis thaliana</i>	NP_200479.1
3	3	<i>Brassica rapa</i>	XP_009120180.1
4	4	<i>Gossypium hirsutum</i>	XP_016682509.1
5	5	<i>Vitis vinifera</i>	XP_003633115.1
6	6	<i>Malus domestica</i>	XP_008368472.2
7	7	<i>Citrus sinensis</i>	XP_006475374.1
8	8	<i>Glycine max</i>	XP_003525302.1
9	9	<i>Arachis hypogaea</i>	XP_025700817.1
10	10	<i>Prunus dulcis</i>	XP_034211989.1
11	11	<i>Lactuca sativa</i>	XP_023754320.1
12	12	<i>Fragaria vesca</i> subsp. <i>vesca</i>	XP_004294177.1
13	13	<i>Solanum tuberosum</i>	XP_006351366.1
14	14	<i>Solanum lycopersicum</i>	NP_001304912.1
15	15	<i>Oryza sativa</i> Japonica Group	XP_015621402.1
16	16	<i>Ananas comosus</i>	XP_020114036.1
17	17	<i>Triticum aestivum</i>	XP_044318137.1
18	18	<i>Zea mays</i>	NP_001398552.1
19	19	<i>Beta vulgaris</i> subsp. <i>vulgaris</i>	XP_010672177.2

[0014] FIG. 5 shows representative images of T1 generation (family A) transgenic maize plants that overexpress NARS1 (left) to wild-type control plants (right) two weeks after germination.

DETAILED DESCRIPTION

[0015] The present invention provides plants that lack root nodules and are engineered to overexpress an asparaginyl-tRNA synthetase 1 (NARS1) protein, seeds produced by said plants, and methods of generating and growing said plants.

[0016] NARS1 is a cytosolic asparaginyl-tRNA synthetase, i.e., an enzyme that catalyzes the attachment of asparagine to its cognate tRNA for use in protein translation. In the Examples, the inventor demonstrates that both trans-

genic *Arabidopsis thaliana* plants (Example 1) and transgenic maize plants (Example 2) that overexpress the *Arabidopsis thaliana* NARS1 protein grow larger and faster than wild-type controls. These results suggest that NARS1 overexpression may be used to increase plant growth, plant growth rate, and/or yield.

[0017] In a previous study, researchers showed that overexpressing NARS1 in the legume *Lotus corniculatus* increased plant biomass and height in a single transgenic line (Yano et al., *Plant Root* 9:6-14, 2014). However, the researchers concluded that the increased biomass observed in this legume was due to increased nitrogen fixation due to an increased number of root nodules in these plants. Additionally, the same group later showed that overexpressing NARS1 in soybean increased plant height and bushiness in a single transgenic line (Arifin et al., *Plant Biotechnol* (Tokyo) 36 (4): 233-240, 2019). Thus, NARS1 overexpression was not expected to have a positive effect on growth or yield in plants lacking root nodules, and the inventor's discovery that increased NARS1 expression results in increased growth in *Arabidopsis thaliana* and maize was surprising.

Plants:

[0018] In a first aspect, the present invention provides plants that lack root nodules and overexpress an asparaginyl-tRNA synthetase 1 (NARS1) protein. The plants may be engineered to overexpress NARS1 relative to a control plant using any method known to those of skill in the art.

[0019] The terms "protein," "polypeptide," and "peptide" are used interchangeably herein to refer to a polymer of amino acids, i.e., a series of amino acid residues connected by peptide bonds between the alpha-amino and carboxy groups of adjacent residues. Proteins may be modified (e.g., via acetylation, glycosylation, etc.) and may include amino acid analogs.

[0020] A used herein, a protein is "overexpressed" in a plant if it is expressed at higher levels than it is expressed in a control plant. For example, a plant that overexpresses a protein may express that protein at levels that are at least 25%, at least 50%, at least 2-times, at least 3-times, at least 4-times, at least 5-times, at least 6-times, at least 7-times, at least 8-times, at least 9-times, at least 10-times, at least 20-times, at least 30-times, at least 40-times, or at least 100-times higher than levels found in a control plant.

[0021] As used herein, a "control plant" is a comparable plant (i.e., a plant of the same species, variety, age, etc.) that was grown under substantially similar conditions but that was not engineered to overexpress a NARS1 protein. Plants that are grown in "substantially similar conditions" are grown in similar locations and soil conditions, are planted with similar timing, are subjected to similar treatments and abiotic stresses, and the like.

[0022] In some embodiments, the plant comprises a construct comprising a heterologous promoter operably linked to a polynucleotide encoding the NARS1 protein. As used herein, the term "construct" refers to a recombinant polynucleotide, i.e., a synthetic polynucleotide that was formed by combining at least two polynucleotide components from different sources. For example, a construct may comprise the coding region of one gene operably linked to a promoter that is (1) associated with another gene found within the same genome, (2) from the genome of a different species, or (3) synthetic. Constructs include plasmids, viral constructs, and

transposon-based constructs. Constructs can be generated using conventional recombinant DNA methods. In some embodiments, the construct comprises multiple different NARS1-encoding polynucleotides and/or multiple copies of a single NARS1-encoding polynucleotide.

[0023] As used herein, the term "promoter" refers to a DNA sequence that defines where transcription of a polynucleotide begins. RNA polymerase and the necessary transcription factors bind to the promoter to initiate transcription. Promoters are typically located directly upstream (i.e., at the 5' end) of the transcription start site. However, a promoter may also be located at the 3' end, within a coding region, or within an intron of a gene that it regulates. Promoters may be derived in their entirety from an endogenous or heterologous gene, may be composed of elements derived from multiple regulatory sequences found in nature, or may comprise synthetic DNA. A promoter is "operably linked" to a polynucleotide if the promoter is positioned such that it can affect transcription of said polynucleotide. The promoters used in the constructs of the present invention are "heterologous", meaning that they are not naturally associated with the NARS1-encoding polynucleotide to which they are operably linked.

[0024] In preferred embodiments, the promoter is a constitutive promoter. A "constitutive promoter" is a promoter that drives transcription of a polynucleotide in most cell types of an organism at most times. In Example 1, the inventor generates transgenic *Arabidopsis thaliana* plants that comprise a construct comprising a cauliflower mosaic virus (CaMV) 35S promoter/enhancer sequence (SEQ ID NO: 26), which comprises the CaMV 35S promoter of SEQ ID NO: 27, operably linked to a NARS1-encoding polynucleotide. In Example 2, the inventor generates transgenic maize plants that comprise a construct comprising the *Zea mays* ubiquitin 1 (UBQ1) promoter (SEQ ID NO: 24) operably linked to a NARS1-encoding polynucleotide. Both the CaMV 35S promoter and the maize UBQ1 promoter are considered strong constitutive promoters for transgene expression in plants. Thus, in certain embodiments, the promoter is a CaMV 35S promoter or a maize UBQ1 promoter.

[0025] In some embodiments, the promoter is an "inducible promoter," i.e., a promoter that allows for controlled expression of a gene under particular conditions or in the presence of a particular molecule (e.g., tetracycline, dexamethasone). For example, the use of a starvation response promoter would allow for NARS1 overexpression only when nutrients are limiting, and the use of a drought-repressive promoter would allow for NARS1 overexpression only in the presence of a drought response.

[0026] Examples of other suitable promoters that may be used in the constructs of the present invention include, without limitation, other ubiquitin promoters, RuBisCO small subunit 1 (RbcS1) promoters, actin promoters, *Agrobacterium* octopine synthase (OCS) promoters, mannopine synthase (MAS) promoters, and Cestrum yellow leaf curling virus (CmYLCV) promoters.

[0027] The terms "polynucleotide," "nucleic acid," and "oligonucleotide" are used interchangeably to refer to a polymer of DNA or RNA. A polynucleotide may be single-stranded or double-stranded and may represent the sense or the antisense strand. A polynucleotide may be synthesized or obtained from a natural source. A polynucleotide may contain natural, non-natural, or altered nucleotides, as well as

natural, non-natural, or altered internucleotide linkages. The constructs of the present invention comprise a polynucleotide that encodes a NARS1 protein (i.e., a “NARS1-encoding polynucleotide”).

[0028] In the Examples, the inventor generated both transgenic *Arabidopsis thaliana* plants that comprise a construct encoding an endogenous NARS1 protein and transgenic maize plants that comprise a construct encoding a heterologous NARS1 protein (i.e., the same *Arabidopsis thaliana* NARS1 protein). Thus, the overexpressed NARS1 protein may be either an endogenous NARS1 protein (i.e., a NARS1 protein that is natively expressed by the plant) or a heterologous NARS1 protein (i.e., a NARS1 protein that is not natively expressed by the plant).

[0029] The plants of the present invention are engineered to overexpress a NARS1 protein. The term “engineered” is used herein to refer to plants that have been altered by the hand of man. Those of skill in the art are aware of multiple methods for engineering a plant to overexpress a particular protein. For example, a plant may be engineered to overexpress a NARS1 protein by introducing a NARS1-encoding polynucleotide into the plant using well-known recombinant or molecular biology techniques. In embodiments in which the NARS1 protein is an endogenous protein, the NARS1-encoding polynucleotide may comprise one or more extra copy of an endogenous polynucleotide (i.e., a polynucleotide that is natively found in the plant) that encodes NARS1. The endogenous NARS1-encoding polypeptide may optionally be altered to include synonymous mutations and/or additional sequences (e.g., adaptor sequences, a sequence encoding a reporter molecule or a protein tag), e.g., to increase protein expression or to increase the ease of cloning or protein detection. In embodiments in which the NARS1 protein is a heterologous protein, the NARS1-encoding polynucleotide comprises a heterologous polynucleotide (i.e., a polynucleotide that is not natively found in the plant) that encodes NARS1. In some embodiments, the engineered plants are genetically modified. For example, in some embodiments, the NARS1-encoding polynucleotide is integrated into the genome of the plant (e.g., using an *Agrobacterium* vector, viral vector, nuclease, or transposase). In these embodiments, the NARS1-encoding polynucleotide may be either inserted randomly into the genome or inserted into a specific location (e.g., via homologous recombination). In other embodiments, the NARS1-encoding polynucleotide remains extrachromosomal (i.e., as part of an extrachromosomal plasmid).

[0030] Alternatively, a plant may be engineered to overexpress a NARS1 protein by upregulating the expression of an endogenous NARS1 gene. As used herein, a gene is “upregulated” if it has manipulated such that it is transcribed at higher levels than it would be in the absence of said manipulation. For example, an upregulated gene may be transcribed at levels that are at least 25%, at least 50%, at least 2-times, at least 3-times, at least 4-times, at least 5-times, at least 6-times, at least 7-times, at least 8-times, at least 9-times, at least 10-times, at least 20-times, at least 30-times, at least 40-times, or at least 100-times higher than the levels at which it would be transcribed in the absence of manipulation. In another alternative, the transcript may be made more stable such that additional NARS1 is generated as compared to a control plant. Upregulation can be accomplished by inserting a regulatory element into the genome such that it is operably linked to a target gene. Examples of

suitable regulatory elements that can be used to upregulate a target gene include, without limitation, promoters, enhancers, and insulators. Alternatively, upregulation can be accomplished using CRISPR-mediated transcriptional activation (CRISPRa). In CRISPRa, a modified nuclease-dead form of Cas9 (dCas9) to which an activator domain has been attached is targeted to a target gene using guide RNAs and used to transcriptionally activate the target gene.

[0031] The plants of the present invention overexpress NARS1 in at least one tissue. The plants may express NARS1 in one, two, three, four, five, six, or more different tissues. For example, the plants may overexpress NARS1 in developing seeds to enhance seed size or yield, overexpress NARS1 in developing fruits to enhance fruit size or yield, overexpress NARS1 in leaves to potentially enhance photosynthesis, carbon sequestration, and/or leaf growth, or overexpress NARS1 in roots to enhance root growth and/or nutrient uptake. Alternatively, the plants may overexpress NARS1 in every tissue or substantially every tissue. Further, the plants may either overexpress NARS1 at one or more particular stages of development or may overexpress NARS1 throughout all developmental stages.

[0032] The term “plant” can refer to a plant at any stage of development or to any part of a plant, including a plant cutting, a plant cell, a plant cell culture, a plant organ, a plant tissue, a plant seed, or a plantlet. In some embodiments, the plant is selected from the group consisting of maize, tobacco, hemp, rice, canola, potato, wheat, cotton, and sugar beet. As is noted in the Examples, the inventor has generated both *Arabidopsis thaliana* and maize plants that overexpress the *Arabidopsis thaliana* NARS1 protein (SEQ ID NO: 2). Thus, in some embodiments, the plants are *Arabidopsis thaliana* or maize.

[0033] The plants of the present invention lack root nodules. A “root nodule” is a swelling on the root of a plant that allows the plant to house nitrogen-fixing *rhizobia* bacteria. (Note: Root nodules are primarily found in legumes, but are also found in Actinorhizal plants (e.g., alder and bayberry) and plants of the genus *Parasponia*.) Thus, the plants of the present invention are not capable of forming a symbiotic relationship with *rhizobia* that results in independent nitrogen fixation. In at least some embodiments, the plants are non-leguminous. The term “non-leguminous” refers to plants that do not belong to the Fabaceae family (i.e., plants that are not a legume).

[0034] The plants of the present invention may overexpress any plant-derived NARS1 protein. NARS1 is well conserved in plants, as is demonstrated in FIG. 4, which is an alignment of NARS1 protein sequences from 18 different plants (i.e., SEQ ID NOs: 2-19). Within the “superrosids” group of eudicots (which includes canola, cotton, grape, apple, citrus, soybean, peanut, almond, lettuce, and strawberry), NARS1 is about 70% identical at the amino acid level based on global alignments of the full-length protein (100% of NARS1 aligned). Within monocots (which include rice, wheat, maize, and banana), NARS1 is 70% identical at the amino acid level with local alignments (>90% of NARS1 aligned) or ~63% identical based on an end-to-end alignment. The N-terminal 50 amino acids, which have no assigned or predicted function, do not align entirely. Within the “superasterids” group of eudicots (which includes sugar beets, potato, and tomato), NARS1 is >63% identical at the amino acid level based on global alignments of full-length NARS1 (100% of NARS1 aligned). As an outgroup, human

cytosolic NARS1 is only 36% identical over 46% of the protein and 16.6% identical across the whole protein. Thus, in some embodiments, the NARS1 protein is at least 90% identical to an amino acid sequence selected from SEQ ID NOs: 2-19 (see Table 1). In the Examples, the inventor generated transgenic plants that overexpress the *Arabidopsis thaliana* NARS1 protein of SEQ ID NO: 2. Thus, in certain embodiments, the NARS1 protein is at least 90% identical to SEQ ID NO: 2. In some embodiments, the NARS1 protein comprises or consists of SEQ ID NO: 2. Seeds:

[0035] In a second aspect, the present invention provides seeds produced by the plants described herein. A “seed” is an embryonic plant enclosed in a protective outer covering. The seeds provided herein are engineered such that they will develop into plants that overexpress NARS1 in at least one tissue.

Methods of Generating Plants that Overexpress NARS1:

[0036] In a third aspect, the present invention provides methods of generating a plant that lacks root nodules and overexpresses NARS1. The methods comprise: (a) introducing a construct comprising a heterologous promoter operably linked to a polynucleotide encoding a NARS1 protein into a plant cell; and (b) growing the plant cell into a plant.

[0037] The term “plant cell” refers to any cell of a plant. A plant cell is the basic structural and functional unit of plants. A plant cell comprises a protoplast and a cell wall. A plant cell can be in the form of an isolated single cell, part of an aggregate of cells, or part of a higher order structure or plant. The plant cells used with the present invention are part of or are derived from plants that lack root nodules.

[0038] As used herein, “introducing” describes a process by which exogenous polynucleotides are introduced into a recipient cell. Suitable introduction methods include, without limitation, *Agrobacterium*-mediated transformation, transposon-based plant transformation, the floral dip method, bacteriophage or viral infection, electroporation, heat shock, lipofection, microinjection, vacuum-infiltration, and particle bombardment.

[0039] In the Examples, the inventor utilized *Agrobacterium*-mediated transformation to introduce NARS1-encoding polynucleotides into plants. Thus, in preferred embodiments, the polynucleotides are introduced via *Agrobacterium*-mediated transformation. In this method, the NARS1-encoding polynucleotide is delivered into plant cells as part of a binary *Agrobacterium* vector, in which it is flanked by two transfer DNA (T-DNA) border repeat sequences. Prior to transformation into plant cells, this binary vector is co-transformed into *Agrobacterium tumefaciens* along with a second vector that is referred to as a vir helper plasmid. The vir helper plasmid encodes components necessary for integration of the region flanked by the T-DNA border repeat sequences into the genome of plant cells. Thus, when the binary vector and the vir helper plasmid are both present in the same *Agrobacterium* cell, proteins encoded by the vir helper plasmid act in trans on the T-DNA border repeat sequences to mediate processing, secretion, and host genome integration of the intervening transgene. Genome insertion occurs without any significant bias with respect to insertion site sequence.

[0040] In the present methods, NARS1-encoding constructs are introduced into a plant cell. Suitable constructs and parts thereof (i.e., heterologous promoters and NARS1-encoding polynucleotides) for use in these methods are described above, in the section titled “Plants.”

[0041] As used herein, “growing” describes a process in which suitable conditions (i.e., light, soil, water, nutrients, temperature) for plant growth are established and maintained.

[0042] Any type of plant that lacks root nodules may be generated using the present methods. In some embodiments, the plant is selected from the group consisting of maize, tobacco, hemp, rice, canola, potato, wheat, cotton, and sugar beet. In some embodiments, the plant is *Arabidopsis thaliana* or maize.

Methods of Growing Plants that Overexpress NARS1:

[0043] In a fourth aspect, the present invention provides methods of growing a plant that lacks root nodules and overexpresses NARS1. The methods comprise (a) planting a seed described herein; and (b) growing the seed into a plant. As used herein, “planting” describes a process in which a seed is placed in soil or another suitable growth medium (e.g., peat moss, coconut coir, vermiculite, perlite, sand, polymer-based gels).

[0044] In the Examples, the inventor demonstrates that transgenic *Arabidopsis thaliana* and maize plants that overexpress NARS1 grow larger and faster than wild-type control plants. Thus, in some embodiments, the plant grown using the present methods has an increased growth rate and/or yield as compared to a control plant.

[0045] The term “growth rate” describes the rate at which the size of a plant increases. Growth rate can be assessed, for example, by measuring the leaf area, leaf width, leaf length, leaf number, stem length, rosette diameter, root length, seed number, seed weight, height, biomass, or volume of a plant over time.

[0046] The term “yield” describes the amount of harvestable (i.e., useful) material produced by a plant. Examples of harvestable plant materials include, without limitation, flowers, pollen, seedlings, tubers, leaves, stems, fruit, seeds, and roots.

[0047] The present disclosure is not limited to the specific details of construction, arrangement of components, or method steps set forth herein. The compositions and methods disclosed herein are capable of being made, practiced, used, carried out and/or formed in various ways that will be apparent to one of skill in the art in light of the disclosure that follows. The phraseology and terminology used herein is for the purpose of description only and should not be regarded as limiting to the scope of the claims. Ordinal indicators, such as first, second, and third, as used in the description and the claims to refer to various structures or method steps, are not meant to be construed to indicate any specific structures or steps, or any particular order or configuration to such structures or steps. All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context. The use of any and all examples, or exemplary language (e.g., “such as”) provided herein, is intended merely to facilitate the disclosure and does not imply any limitation on the scope of the disclosure unless otherwise claimed. No language in the specification, and no structures shown in the drawings, should be construed as indicating that any non-claimed element is essential to the practice of the disclosed subject matter. The use herein of the terms “including,” “comprising,” or “having,” and variations thereof, is meant to encompass the elements listed thereafter and equivalents thereof, as well as additional elements. Embodiments recited as “including,” “comprising,” or “hav-

ing” certain elements are also contemplated as “consisting essentially of” and “consisting of” those certain elements.

[0048] Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein. For example, if a concentration range is stated as 1% to 50%, it is intended that values such as 2% to 40%, 10% to 30%, or 1% to 3%, etc., are expressly enumerated in this specification. These are only examples of what is specifically intended, and all possible combinations of numerical values between and including the lowest value and the highest value enumerated are to be considered to be expressly stated in this disclosure. Use of the word “about” to describe a particular recited amount or range of amounts is meant to indicate that values very near to the recited amount are included in that amount, such as values that could or naturally would be accounted for due to manufacturing tolerances, instrument and human error in forming measurements, and the like. All percentages referring to amounts are by weight unless indicated otherwise.

[0049] No admission is made that any reference, including any non-patent or patent document cited in this specification, constitutes prior art. In particular, it will be understood that, unless otherwise stated, reference to any document herein does not constitute an admission that any of these documents form part of the common general knowledge in the art in the United States or in any other country. Any discussion of the references states what their authors assert, and the applicant reserves the right to challenge the accuracy and pertinence of any of the documents cited herein. All references cited herein are fully incorporated by reference, unless explicitly indicated otherwise. The present disclosure shall control in the event there are any disparities between any definitions and/or description found in the cited references.

[0050] The following examples are meant only to be illustrative and are not meant as limitations on the scope of the invention or of the appended claims.

EXAMPLES

Example 1

[0051] In the following example, the inventor demonstrates that transgenic *Arabidopsis thaliana* plants that over-express NARS1 grow larger and faster than wild-type controls.

BACKGROUND

[0052] Target of rapamycin (TOR) is a highly conserved serine/threonine protein kinase that coordinates growth and development with nutritional status in eukaryotes. TOR regulates key pathways such as nucleotide biosynthesis, ribosome biogenesis, and leaf initiation. TOR is well-studied in humans because it is dysregulated in many human diseases, including diabetes and cancer. In animals and fungi, many of the upstream mechanisms that activate TOR are known, but almost nothing is known about how TOR activity is regulated in plants.

[0053] Asparaginyl-tRNA synthetase 1 (NARS1; official names: EMB2755 and SYNC1; gene ID: At5g56680) was identified as a potential regulator of TOR in plants in a

forward genetic screen for defective embryonic cell-cell (plasmodesmatal) trafficking that repeatedly identified mutants defective in TOR signaling (Brunkard et al., Proc. Natl. Acad. Sci. U.S.A 117 (9): 5049-5058, 2020; Kim et al., Development 129 (5): 1261-1272, 2002). NARS1 mutants have stunted growth and defective TOR signaling. RNA sequencing revealed that NARS1 mutants exhibit several signatures of TOR inactivation, including significant repression of ribosomal protein genes and induction of proteolytic and catabolism-related genes (Busche et al., Plant Cell 33 (5): 1615-1632, 2021; Xiong et al., Nature 496 (7444): 181-186, 2013). Further, NARS1 mutants and knockdowns also show significantly less phosphorylation of S6K-T449 (orthologous to human S6K-T389), a residue that is uniquely phosphorylated by TOR kinase in plants. Thus, NARS1 activates TOR in plants and represents the first proposed amino acid sensor in plants.

Materials and Methods:

[0054] RNA was extracted from *Arabidopsis thaliana* Col-0 and used as a template for RT-PCR with SuperScript III and Phusion, following the manufacturer’s instructions. The NARS1 coding sequence was amplified from this RNA using the forward primer caccATGGCTGATGAGATTGTG (SEQ ID NO: 20) and the reverse primer AAGATCAGCTTTTCCAGGATAGCG (SEQ ID NO: 21), which were synthesized by IDT DNA. The forward primer included a CACC overhang for directional cloning into D-TOPO pENTR vectors. The reverse primer was designed to exclude the final stop codon from the NARS1 coding sequence to allow translational readthrough to C-terminal epitope tags. The PCR product size was verified by agarose gel electrophoresis and was purified using a gel extraction kit (NEB Monarch kit).

[0055] The product was subcloned into pENTR using the D-TOPO pENTR kit (Invitrogen) and used to transform chemically competent *E. coli* cells (genotype DH10B) using kanamycin selection. Resistant colonies were screened for by colony PCR and positive transformants were used in minipreps to isolate plasmid (NEB Monarch kit). Purified plasmid was sequenced using Sanger sequencing. This plasmid was digested with EcoRV and recombined with pEarleyGate 103 using Gateway LR Clonase (for full plasmid information, see abrc.osu.edu/stocks/number/CD3-685 [abrc.osu.edu]). pEarleyGate 103 includes a C-terminal GFP tag and a CaMV 35S promoter. Insertion into pEarleyGate 103 was validated using Sanger sequencing. The resulting plasmid is referred to herein as 35S: PRO: NRS1: pEG103.

[0056] 35S: PRO: NRS1: pEG103 was introduced into *Agrobacterium tumefaciens* (genotype GV3101) and used to transform *Arabidopsis thaliana* inflorescences following the standard floral dip protocol. Positive transformants were screened for herbicide resistance, NARS1 insertions were validated by PCR, and NARS1 overexpression was validated by fluorescence microscopy. Dwarf transgenic lines showed an insertion but no GFP fluorescence, indicating gene silencing, whereas large transgenic lines showed strong GFP fluorescence in leaves.

Results:

[0057] To test the effects of constitutively overexpressing NARS1 in plants, the *Arabidopsis thaliana* NARS1 coding sequence (SEQ ID NO: 1, which encodes the NARS1

protein of SEQ ID NO: 2) was cloned into pEarleyGate 103 and was used to transform *Arabidopsis thaliana* ecotype Col-O using an *Agrobacterium* vector (GV3101) and the floral dip method. Several stable transgenic *Arabidopsis thaliana* lines that overexpress NARS1 fused to a GFP reporter were generated. Consistently, NARS1 transgenic plants with visible GFP fluorescence (which indicates successful overexpression of the NARS1 protein) were found to grow larger and faster than wild-type controls (FIG. 1A). On average, NARS1 overexpression significantly increased the leaf width and length compared to wild-type controls (Col-0) (FIG. 2 and FIG. 3). This phenotype is similar to TOR overexpression, which causes increased organ and cell size. Yet, overexpressing another annotated asparaginyl tRNA synthetase protein, i.e., NARS2, had no effect on plant growth or TOR activity (FIG. 1B). This indicates that this effect is specific to NARS1 and not a general property of asparaginyl tRNA synthetases. These results suggest that NARS1 overexpression could be used in agriculture to improve plant growth and yield.

Example 2

[0058] In the following example, the inventor demonstrates that transgenic maize plants that overexpress NARS1 grow larger and faster than wild-type controls.

Materials and Methods:

Cloning ZmUbi1_{PRO}:NARS1

[0059] The *Arabidopsis thaliana* NARS1 coding sequence was subcloned into a plasmid such that it was flanked by the promoter and terminator from the *Zea mays* ubiquitin 1 (UBQ1) gene to drive high, consistent levels of NARS1 expression throughout maize development. Specifically, an open reading frame (ORF) encoding the *Arabidopsis thaliana* NARS1 protein was synthesized de novo using GenSmart to optimize codons for expression in maize. Adaptors were added to the 5' and 3' ends of the ORF to facilitate GoldenGate cloning. The 5' adaptor sequence is GGTCTCTA and the 3' adaptor sequence is GCTTTGAGACC (SEQ ID NO: 22). Both adaptor sequences include Bsal recognition sites. The sequence of the resulting construct is provided as SEQ ID NO: 23.

[0060] This construct was subcloned, using a GoldenGate cloning strategy, into the T-DNA of a binary vector. The resulting vector includes the *Zea mays* Ubiquitin 1 (ZmUbi1) promoter (SEQ ID NO: 24), which includes a 5' leader sequence with an intron, upstream of the NARS1

coding sequence. The resulting vector also includes the ZmUbi1 terminator (SEQ ID NO: 25), which includes a 3' untranslated region, downstream of the NARS1 coding sequence. The resulting binary vector is referred to herein as ZmUbi1_{PRO}:NARS1.

Maize Transformation

[0061] Maize cells of inbred genotype LH244 were transformed using *Agrobacterium tumefaciens* carrying the ZmUbi1_{PRO}:NARS1 binary vector, which also includes a gene that confers resistance to the herbicide glufosinate. Transformants were selected for glufosinate resistance and cultured under conditions to induce plant regeneration. Mature transformants were self-pollinated to generate T1 seeds for further analysis.

Growth Assays

[0062] T1 seeds from two independently transformed parents were sown on wet potting soil and grown in a greenhouse in Madison, WI with ~16 h daylength for two weeks. The transgenic plants were grown alongside (i.e., in the same flat) untransformed LH244 sibling plants. All seeds germinated at similar times. Two weeks after germination, plants were photographed and analyzed for differences in growth and vigor.

Image Analysis

[0063] All images were analyzed using ImageJ.

Results:

[0064] T1 ZmUbi1_{PRO}:NARS1 plants from two independent parents (family A and family B) were compared to wild-type sibling controls two weeks after germination in the greenhouse on potting soil mix (FIG. 5). Under these conditions, the average height of wild-type LH244 plants was 6.8 cm (n=7, standard deviation 3.4 cm), the average height of family A ZmUbi1_{PRO}:NARS1 plants was 8.9 cm (n=9, standard deviation 2.1 cm), and the average height of family B ZmUbi1_{PRO}:NARS1 plants was 8.6 cm (n=5, standard deviation 2.1 cm). Therefore, the ZmUbi1_{PRO}:NARS1 plants from family A were 30.6% taller than wild-type plants, and the ZmUbi1_{PRO}:NARS1 plants from family B were 26.6% taller than wild-type plants, on average. Transgenic ZmUbi1_{PRO}:NARS1 plants also exhibited less variability in height, with standard deviations that were 61.5% lower than the standard deviations observed for wild-type plants. Transgenic ZmUbi1_{PRO}:NARS1 plants from both families also qualitatively appeared to have larger, greener, and more vigorous shoots than the wild-type plants.

SEQUENCE LISTING

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Sequence total quantity: 27
SEQ ID NO: 1          moltype = DNA length = 1716
FEATURE              Location/Qualifiers
source                1..1716
                     mol_type = genomic DNA
                     organism = Arabidopsis thaliana

SEQUENCE: 1
atggctgatg agattgtgcc gccggcgact caattagccg cegtatcett agaaaaacgat 60
ggatctacgg tgcaaagagc tcaattctct aaccgggttt tgattcgaac gattttggat 120
cgacctgacg gcggagctgg actcggcggg caaacggttc gaatcggtgg ttgggtaaaa 180
tccggaagag atcaagggaa aagaacgttt tctttccttg cggttaacga tggatcttgt 240
ccagcgaatc ttcaggttat ggtggatccg tctctctatg atgtctctaa cttggttgct 300
acgggaactt gtgtgactgt tgatggagtt ttgaaggttc caccgaaagg taaaggtacg 360
    
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cagcagcaga ttgagcttaa cgttgtgaag gtgattgatg ttggaacggt ggatgcgctc 420
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cgttctagaa ctaactcgat ctcagctggt gcaagaattc gtaatgctct tgcttttgcg 540
actcacagtt tctttcaaga gcatagtttc ctttacattc acactcctat catcacgaca 600
agtgattgtg aaggtgctgg tgaaatgttc caagcaacaa ctttgatcaa ttacactgag 660
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aggctcattg tcattgagag aggtaatggt gtagcggaac tgaagctgc caaagcaagc 780
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tctcacactt caaggcatct tgctgaattc tggatggttg agcctgagat tgcttttgca 1080
gatctagagg accacatgaa ctgtgcagag gcgatgtga aatacatgtg caactggttg 1140
cttgagaaat gttacgctga ctgggaactt atggctaaga atttcgactc aggatgcatt 1200
gacaggctaa aattgggtgc ctctactccg tttggcgcta taacatacac taaagcaata 1260
gagctacttg aggaagctgt ggctaaagga aaggaatttg ataacaatgt ggagtgggga 1320
atcgacttag cctcggagca tgaaagatac ttgacagagg ttctgtttca aaagccactt 1380
attgtctata actaccgaa aggaatcaaa gctttttaca tgagacttaa cgatgatgag 1440
aagaacgttg ctgcoatgga tgcctcgtt ccaaaggttg gagaactcat tggtggaagc 1500
caaaggggaa aacgatatga tgtcatcaaa aagaggattg aggagatggg tcttccaata 1560
gagccatatg agtggtagct agactgccc cgttatggaa cagtgaagca ttgtgggttt 1620
ggacttgctc tcgaacgtat gattctctc gctactggac ttgataacat cagagacggt 1680
attcctttcc ctgctatcc tggaaaagct gatctt 1716

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SEQ ID NO: 2          moltype = AA length = 572
FEATURE              Location/Qualifiers
source                1..572
                     mol_type = protein
                     organism = Arabidopsis thaliana

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SEQUENCE: 2
MADEIVPPAT QLAAVSLEND GSTVQRAQFS NRVLIRTILD RPDGGAGLAG QTVRIGGWVK 60
SGRDQGKRTF SFLAVNDGSC PANLQVMVDP SLYDVSNLVA TGTCVTVDGV LKVPKKGKT 120
QQQIELNVVK VIDVGTVDAS KYPLPKTKLT LETLRDVLHL RSRTNSISAV ARIRNALAFA 180
THSFFQEHFS LYIHTPIITT SDCEGAGEMF QATTLINYTE RLEQDLIDNP PPTBADVEEA 240
RLIVIERGNV VAEKAAKAS KEAITAAVAE LKIAKETFAH IDERSRLRPG LPKKGNDIDY 300
SKDFFGRQAF LTVSGQLQVE TYACALSINVY TFGPTFRAEN SHTSRHLAEF WMVEPEIAFA 360
DLEDDMNC AE AYVKYMCNWL LEKCYADMEL MAKNFDSGCI DRLKLVASTP FGRITYTKAI 420
ELLEBAVAKG KEFDMNVEWG IDLASEHERY LTEVLFQKPL IVYNYPKGIK AFYMLRNDD E 480
KTVAAMDVLV PKVGELIGGS QREERYDVIK KRIEEMGLPI EPYEWYLDLR RYGTVKHCGF 540
GLGFERMILF ATGLDNIRDV IPPPRYPGKA DL 572

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SEQ ID NO: 3          moltype = AA length = 572
FEATURE              Location/Qualifiers
source                1..572
                     mol_type = protein
                     organism = Brassica rapa

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SEQUENCE: 3
MGDIIPPVD QLAAVSLTSD SSTVQKARFS DRVRIQSILG RPDGGAGLAG QKVRISGWVK 60
TGREQKGAF AFLVNDGSC PANLQVMVDA SVSLSKLLIA TGTCVTVDGC LKLPPEGKGT 120
KQKIELSVVE VVDVGTVDPG TYPKPKTKLT LERLREFLHL RARTNSISAI ARIRHALAIA 180
THTFFDEEFG LYVQTPIITT SDCEGAGEMF QVTTLISTTE KLERELIENP PPTBADVEEA 240
RVVVKERGEA VAEKAAKAS KEAILASVAE LNDAKANLAA TEARSRLKPG LPKVDGKIDY 300
SQDFFGRQAF LTVSGQLQVE TYACGLSDVY TFGPTFRAEN SHTSRHLAEF WMVEPELAF A 360
DLEDDMNC AE AYVKYMCNWL LEKRYDDMEL MAKNFDKGCI DRLKLVASTP FGRLTYTEAI 420
KILEBAVAKG KKFDMNVEWG IDLASEHERY LTEVVFQKPL IVYNYPKGIK AFYMLRNDD G 480
KTVAAMDVLV PKVGELIGGS QREERIDVIM ERLEEIGLPV EPYEWYLDLR RYGTAKHSGF 540
GLGFERMVLF ATGMNDIRDV IPPPRYPGRA DL 572

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SEQ ID NO: 4          moltype = AA length = 565
FEATURE              Location/Qualifiers
source                1..565
                     mol_type = protein
                     organism = Gossypium hirsutum

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SEQUENCE: 4
DQALPPSDQL AAMNLTDTVE KHAFSDRVLI RSIVGRPDGG AGLAGQVRVA GGWVKTGREQ 60
GKGTFAFLEL NDGSCPANLQ VIVDAGLAVL SKLVATGTCV VVDGILKVPV EGTRQKIELR 120
VEKVVSVEVE DPAKYPIPKT KLTLEFLRDH LHLRARTNTI AAIARIRNAL AFATHSFFQE 180
HNFLVHTPI LTTSDCEGAG EMPQVTLTIS ESEKLEKELI KNPPPSEVDI EAARQVVSER 240
GEAVKQLKAA KASKSEITAS VAEKAAKAKEN LSKLEERSKL KPGIPKKGK IDYTQDFAR 300
QAFLTVSGQL QVETYACAVS NVYTFGPTFR AEHSHTSRHL AEFWMVEPEI AFADLQDDMN 360
CAEAIVKYMC NWLLDKCLDD MLFMAKSYDK GCIDRLRMVA SVPPVRSIVT EAVEELLEAV 420
RGGKKFENEV KWGIDLASEH ERYLTEVKFQ KPLIVYNYPK GIKAFYMLRN DDLKTVAAAM 480
VLVPKVGELI GGSQREERYE VIRERILEVG LPLEPEYEWY LDRRYGTVKH CGFGLGFERM 540
ILFATGIEMI RDVIPPRYP GRADL 565

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SEQ ID NO: 5          moltype = AA length = 570

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FEATURE Location/Qualifiers
source 1..570
mol_type = protein
organism = Vitis vinifera

SEQUENCE: 5
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TGREQKGSF AFLELNDGSC PANLQVIUDA AVAPLQQLVQ TGTCTVHVEGL LKVPPEGTKQ 120
RVELRVEKVH HVGPVDPAPY PLPKTRLTLE FLRDFVHFRP RTNTISAVAR IRNALAYATH 180
TFFQNHGFLY IHTPIITSD CEGAGEMFQV TTLISDAEKV EKELIKNPPP SEADIEAACA 240
LVKEKGEAVA QLKSAKASKG EITASVAELN KAKENLSRLE ERSKLKPGIP QKDGKIDYSQ 300
DFFARQAFLT VSGQLQVETY ACAVSSVYTF GPTFRAEHSR TSRHLAEFWM VEPEIAFADL 360
KDDMNCAEAY VKFLCQWLLD NCIDDMEFMA KNFDKGCIDR LRMVASTPFE RISYTEAIKL 420
LEEAVKDKKK FENKVEWGID LASEHERFLT EVLFKPKVIV HDYKPKIKAF YMLNDDMKT 480
VAAMDVLVVK VEGELIGGSQR EERYEVIEKR ILEMGLPLEP YEWYDLRRY GTVKHCGFGL 540
GFERMILFAT GIDNIRDVIP FPRYPGRADL 570

SEQ ID NO: 6 moltype = AA length = 569
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source 1..569
mol_type = protein
organism = Malus domestica

SEQUENCE: 6
MADDSTPPSA QLAAVTSLND ASVLKAEFSD RVLIRSIISR PDGGGSLAGQ KIRIGGWVKT 60
GRKADKDAFA FLELNDGSCA SNLQVIVKAD KYDLGQLVQT GTCLSDVDGVL KLPPDGKKQK 120
VELDVEKVIH VGPVDPAPY LPKTRLTLEF LREHVHLRSR TNTISAVTRI RNALAPATHS 180
FFHKHGFLYV QTPPIITSDC EGAGEMFQVT TLFSEAEKLE KDLIENPPPS EADIEAAKLI 240
VKERGGDDVAQ LKSAKASKQE IGAAVAELKK AKDNVLLKEE RSKLKPPIPR KDGKVDYTDQ 300
FFARQAFLTV SGQLQVESYA CALSSVYTFG PTFRAENSHT SRHLAEFWMV EPEIAFAELE 360
DDMNCAEAYV KFLCQWLLDD CREDMEFMAD KIDKTCIDRL TMVAKTPPKR ITYTEAVDLL 420
IDAVKKGKFF ENHVEWGIDL ASEHERYLTE VLFQKPVIVY NYPKGIKAFY MRLNDDLKTV 480
AAMDVLVVKV GELIGGSQRE ERYDIIMSRI EEMGLPAEYP EWYDLRRRFG TVKHCGFGLG 540
FERMVLFATG IDNIRDVIPF PRYPGRADL 569

SEQ ID NO: 7 moltype = AA length = 567
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source 1..567
mol_type = protein
organism = Citrus sinensis

SEQUENCE: 7
MADNELPVDQ VATMDLNDDA VQRHQFSDRV LIKSILTRPD GGAGLAGRQV RVGGWVKTGR 60
EQGKGSFAFL EVNDGSCPAN LQVIVDKDVA DLGQLVPTGT CVYVEGMLKN PPEGTKQKIE 120
LRVQKVVVDG MVDPAKYPIP KTKLTLLEFLR DRIPFRPRNT TIAAVARIRN ALAYATHPTL 180
QKQGLYIHT PIIITSDCEG AGEMFQVTTL ISDADKLEKE LIKNPPPSEA DIEAAKLVIK 240
EKGEAVAKLK SDKAGREAIS ASVTELTAK ENLAKLEERS KLKPGIPQKD GKIDYTDQDF 300
ARQAFLTVSG QLQVETACVA VSNVYTFGPT FRAEHSHTSR HLAEFWMVEP EMAFSDLKDD 360
MNCAEAYVKF MCDWLLDCHF DMEFMKAKNY DKSCINRLRM VASTPFRIT YTEAVELLE 420
AVKEGKHFEN KVEWGIDLAS EHERYLTEVK FQKPVIVYNY PKGIKAFYMR LNDDLKTVA 480
MDVLVPKVG LIGGSQREER YDVIKSRIED MGLPLEPEYEW YDLRRRFGTV KHSFGFLGFE 540
RMILFATGID NIRDVIPPR YPGRADL 567

SEQ ID NO: 8 moltype = AA length = 566
FEATURE Location/Qualifiers
source 1..566
mol_type = protein
organism = Glycine max

SEQUENCE: 8
MADASSPPTD QLAATLDEV PKANFSDRVP IRSIISRTDG GSGLAGRKAR VGGWVKTGRK 60
ADKDAFAFLE INDGSCAGNL QVIVEAALGE LGQLVPTGTC VVVDGHLKLP PAGTRQKIEL 120
RADKVLHVGP VDPAPYPLPK MRLTLEFLRD FVHLRSRTNT ISAVARIRNA LAYATHTFN 180
KEGFLYVHTP IVTSDCEGA GEMFQVTTL SEAERLEKEL LQNPSPSEAD VEAARVVVQE 240
KGEVVSQKKA AKASKQEIGA AVDQLKKAKE SLAKVEEWSK LKPGIPKIDG KVDYKDFFA 300
RQAFLTVSGQ LQVESYACAL SSVYTFGPTF RAENSHTSRH LAEFWMVEPE IAFELKDDM 360
NCAEAYVKFM CQWLLDNCLE DMEFMADKFD KGCIDRLKLV ASTPFIRVY TEAVEILED 420
VKNGKGFENE VKGWIDLASE HERYLVEVKF QKPVIVYNY KDIKAFYMLR NDDLKTVAAM 480
DVLVPKVGEL IGGSQREERY DVIQTRIKEM GLPVEPEYEW YDLRRYGTVK HAGFGLGFER 540
MILFATGLEN IRDVIPPRY PGRADL 566

SEQ ID NO: 9 moltype = AA length = 561
FEATURE Location/Qualifiers
source 1..561
mol_type = protein
organism = Arachis hypogaea

SEQUENCE: 9
AEQLAATTIN DDGIVPKAEF SDRVPINSII SRADGGGSLA GKARVGGWV KTGRKADKDA 60
FAPLELNDGS CPGNLQVIVE AGLVELSQVV PTGTCVLDG ELKLPPEGAK QKVELRAEKV 120
VHVGPVDPAP YPLPKMRLTL EFLRDFVHLR SRTNTISAVA RIRNALAYAT HTPFNKHGFL 180

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YVHTPIITTS	DCEGAGEMFQ	VTTLLSEAE	LEKELIHNPP	PTEAEVEAAR	VVVKKEGEIV	240
SQLKSAKASK	KEIGAAVDEL	KKSKDTLSKL	BERSKLPKI	PKKDGNDVYA	KDFFARQAPL	300
TVSGQLQVES	YACALSSVYT	FGPTFRAENS	HTSRHLAEFW	MVEPELAFAE	LKDDMNCAEA	360
YVKFLCQWLL	DNCLEDKMF	ADKFDKGCID	RLKMVASTPF	VRVSYTEAVE	ILEEAVKNGK	420
KFDNEVKWGI	DLASEHERYL	TEVKFQKPI	VYNYPKDIKA	FYMRLNDDSK	TVAAMDVLVP	480
KVGELIGGSQ	REERYDIVQ	RIKEMGLPLE	PYEWYDLRR	YGTVKHSGFG	LGFERMILFA	540
TGLENIRDVI	PPFRYPGRAD	L				561

SEQ ID NO: 10 moltype = AA length = 565
 FEATURE Location/Qualifiers
 source 1..565
 mol_type = protein
 organism = Prunus dulcis

SEQUENCE: 10

MADDLTAQLA	TATLDNNASV	QRAQFSNRVP	IKSIILRPDG	GSGLAGKLAR	VGGWVKTRK	60
ADKDAFAPLE	LNDGSCSGNL	QVIVEADKGD	LGQLVLTGTS	LVVDGVLKLP	PDGKRQKVEL	120
RVEKVIHVGL	VDPSKYPLPK	TRLPLEFLRD	YVHLRSRTNT	ISAVLRIRDA	LAYATHFFPH	180
DHGFRYVQTP	IITSDCEGA	GEMFQVTTLI	NEAEKLEKDL	IENPPSEADI	EAAKLIAKER	240
GIDVAQLKSA	KASKQEIIGAA	VVELQKAKDN	LVKLEERSKL	QPGIPRKDGK	IDYTQDFFR	300
QAFLLTVSGQL	QVESYACLSL	TVYTFGPTFR	AENSHTSRHL	AEFWMVEPEI	AFAELEDDMN	360
CAEAYVKYLC	QSLLDNCRE	MEFMADKIDK	SCIDRLTMVA	KTPFVRI TYT	EAVELEL TEAV	420
KNGKFKENHV	EWGIDLASEH	ERYLTVRFQ	KPVIVYNYPK	GIKAFYMRLN	DDSKTVAAMD	480
VLVVKVGEI	GGSQREERYD	VIVSRIKEMG	LPLEPEYEWL	DLRRYGTVKH	CGFGLGFERM	540
VLFATGIDNI	RDVIPFRYP	GRADL				565

SEQ ID NO: 11 moltype = AA length = 566
 FEATURE Location/Qualifiers
 source 1..566
 mol_type = protein
 organism = Lactuca sativa

SEQUENCE: 11

MADNGVLPVD	KLTISETVKE	AEFSQRVLR	SILGRPDGGA	GLAGQTLKVG	GWVKKGREQG	60
KGSFAPLELN	DGSCTANLQL	IIYSDVAPLG	QPTPTGTSLH	VEGLVQMPPA	DKQKQSIEL	120
KVSKVLDVGA	ADPAKYPLPK	TRLTLEFLRD	YVHLRPRNT	ISAIARIRNA	LAYATHFFQ	180
KHGFLYVHTP	IITSDCEGA	GEMFQVTTLI	NDSEKLEKEL	LKNPPPSQED	VDAARAAVKE	240
KGGIVAKLKS	LKADKSAITV	AVAELEKAKE	TLKIEEERFN	QKPGIPKIDG	KVDYSQDFFA	300
RQAFLLTVSGQ	LQVETFACAL	SSVYTFGPTF	RAEHSHTSRH	LAEFWMVEPE	IAFADIEDDM	360
KCAEAYVRFM	CQWLLDNCLD	DMEFIAEKFD	EHAIRLKMV	ASTNFVRLTY	TEAVTILEEA	420
VSKGHQFENK	VEWGIDLASE	HEKYL TETKF	ESPVIVYNYP	KGIKAFYMKV	NPDNKTVAM	480
DVLVVKVGEI	IGGSQREENY	EVIKERILEM	GLPLEPEYEW	LDLRRYGTVK	HSGFGLGFER	540
MILFATGIDN	IRDVIPFRPF	PGRADL				566

SEQ ID NO: 12 moltype = AA length = 569
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 source 1..569
 mol_type = protein
 organism = Fragaria vesca

SEQUENCE: 12

MADDAQLGSG	QLAETALNES	VSDLKAEFSD	RVPIKSIISR	PDGGSAPAGQ	KVRVGGVVK	60
GRKADKDAFA	FLELNDGSCP	GNLQVIVQAD	KGDLGQLVPT	GTCVVVEGEL	KLPPAGVKQK	120
VELRVEEVHL	LGPVDPLKYP	LHKGKISLER	LRDVVHLRPR	TNTISAVARI	RDDLAYATH	180
FFRKHGFRV	HTPIITSDC	EGAGEMFQVT	TLISEGERLE	RELIKNPPPS	EADVEANLI	240
VTEKGDVAVS	LKSAKASKDE	IGAAVAELKR	AKENVLKLQE	RAKLQPGIPK	KDGKVDYTQD	300
FFARQAFLLTV	SGQLQVESYA	CAISSVYTFG	PTFRAENSHT	SRHLAEFWMV	EPELAPAELE	360
DDMNCAEAYV	KFLCKWLLDN	CYDDMEFFSR	QYDKTCIERL	RMVASTPFER	ITYTEAVELL	420
IDAVKNGKKF	ENPVEWGIDL	ASEHERFLTE	VKFPQKPIVY	NYPKGIKAFY	MRLNDDNKTV	480
AAMDVLVVKV	GELIGGSQRE	ERYDVIHSRI	AEMDLPIEPY	EWYLDLRRFG	TAKHSGFGLG	540
FERMVLFATG	MDNIRDVIPF	PRFPGRADL				569

SEQ ID NO: 13 moltype = AA length = 563
 FEATURE Location/Qualifiers
 source 1..563
 mol_type = protein
 organism = Solanum tuberosum

SEQUENCE: 13

MSVDSAPPVE	KLTLSDAVEE	AKFSQRVPIR	SIVGRPDGGA	GLAGNVVKIG	GWVKTGREQG	60
KGSFAPLEVN	DGSCPGNLQV	IVDASVHKL	DLVLTGTCVH	VEGELKVPPE	GAQKQVELRV	120
QKVISVGTVD	AAKYPLPKTK	LTLLEFLRDV	HLRSRTNTIS	AVARIRNALA	YATHFFQNN	180
GFLYIHTPII	TSDCEGAGE	MFQVTSLSL	AEKLEKELKE	NPAPSESDIQ	AAEQLAKEKG	240
EVVAQLKASK	ASNKISAAV	AELQRAKENL	LKLQERSRLS	AGIPKKGDKI	DYSEDFFRQ	300
AFLTVSGQLQ	VETYACALSS	VYTFGPTFRA	EQSHTTRHLA	EFWMVEPEIA	FADIQDDMNC	360
AEAYVKFLCQ	WLLDHCLDDM	EFMTKFDVKD	ALSRLRMVAS	SNCHRITYTE	AIDILEVAAK	420
VKKFENKVEW	GIDLASEHER	YLTEEHFKAP	VIVFNYPKGI	KAFYMKVNE	NKTVAAMDLL	480
VPKVGEIIGG	SQREENYEV	RSRILDMGLP	LEPYEWYLDL	RRYGTVKHSG	FGLGFERMIL	540
FATGIENIRD	VIPFRYPGR	ADL				563

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SEQ ID NO: 14 moltype = AA length = 563
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 source 1..563
 mol_type = protein
 organism = Solanum lycopersicum

SEQUENCE: 14

MSVDSAPPVE	KLTLTDVVEE	ARFSQRVPIC	SIVGRTDGGG	GLAGNVVKIG	GWVKTGREQQ	60
KGSFAPLEVN	DGSCPGNLQV	IVDASVHKLK	DLVLTGTGVH	VEGELKVPPE	GAQKQVELRV	120
QKVISVGTVD	AAKYPLPKTK	LTLEFLRDVV	HLRSRTNTIS	AVARIRNALA	YATHTPFQNN	180
GFLYIHTPII	TTSDCEGAGE	MFQVTLISE	AEKLEKELKE	NPAPSESDTQ	AAEKLAKEKG	240
EVVAQLKASK	ASNKKISAAV	AELQRAKENL	LKLQERSRLS	AGIPKKGDKI	DYSEDFFRGQ	300
AFLTSGGQLQ	VETYACALSS	VYTFGPTFRA	EQSHTTRHLA	EFWMVEPEIA	FADIQDDMNC	360
AEAYVKFLCQ	WLLDHCLDDM	EFMTKFDVKD	ALSRLRMVAS	SNCHRITYTE	AIDILEVAAK	420
VKKFENKVEV	GIDLASEHER	YLTEEHFKAP	VIVFNYPKGI	KAFYMKVNEE	NKTVAAMDLL	480
VPKVGELIGG	SQREENEYVL	RSRILDGLPL	LEPYEWYLDL	RRYGTVKHSG	FGLGFERMIL	540
FATGIENIRD	VIPFPYPGR	ADL				563

SEQ ID NO: 15 moltype = AA length = 536
 FEATURE Location/Qualifiers
 source 1..536
 mol_type = protein
 organism = Oryza sativa

SEQUENCE: 15

TDRTIRISIL	ADGAARAGER	VVVGWVKTG	REQKGTGTFP	LELNDGSCAS	NLQVLVDAAV	60
HPLAPLTATG	TSVLVEGELK	KPPEGAKQRV	ELRVDRVIEV	GEVDPAAVPL	PKTKLTLENL	120
RDVHLRSRT	NTIGAVARIR	HQLACATHRF	FDENGFLYVH	TPIITTSDC	GAGEMFQVTT	180
LFSHAEKVEK	ELKENPAPSE	SDIEAARVVV	KEKGDVAQAL	KAASKAQEI	TAAVAELNKA	240
KENVSRLEER	SKLKPGLPRD	DGTVAYENDF	FKRQAFLTVS	GQLQVETVAC	ALSSVYTFGP	300
TFRAENSHTS	RHLAEFWMVE	PEIAFANLQD	DMNCAERYVQ	YLCKWLLHEC	REDMEPMVKN	360
YDKTAIERLE	LVSSTPPQRI	SYTKAVELLK	NVTDKKFENK	VEWIGDLASE	HERYLTEVIF	420
KKPVIVYNYV	KEIKAFYMLR	DDQKTVAAM	DVLVPKVGEL	VGGSQREERL	DLKTRIQDA	480
GLPLEPYEWY	LDLRRFGSVK	HSGFGLGFER	MILFATGLEN	IRDVIPFPYP	PGRADL	536

SEQ ID NO: 16 moltype = AA length = 553
 FEATURE Location/Qualifiers
 source 1..553
 mol_type = protein
 organism = Ananas comosus

SEQUENCE: 16

GDESAPPTSE	MAEPSLQRTL	IKSIVSSSAA	AAALVGRVVV	VVGWVKTGRE	QKGTGTFAPLE	60
LNDGSCLANL	QVIVDAKVHP	LSELVPTGTC	VLVEGELKKP	PEGTKQRIEL	RVDKVLALGP	120
VDSKYPLPKT	RLTLENLREV	VHLRARTNTI	SAVARIRDEL	AYGHTTFFRK	NGFRYVHTPI	180
ITTSCEGAG	EMFQVTLFS	LAEKTEKELK	QNPPSESEV	EAAKVLVKEK	GEAVAQLKSS	240
KASKEEISAS	VAELTKAKEN	LSRLEERFKL	KPGIPRKDGG	IAPENDFFKR	QAFLTVSGQL	300
QVETVACALG	NVYTFGPTR	AENSHTSRHL	AEFWMVEPEM	AFANLQDDMN	YAESYVKFLC	360
QWLLDHCLD	MEFMVKNYDK	TAIERLKLVA	SVPFEHISYT	KAVQLLKNVT	DKEFENKVEW	420
GIDLASEHER	YLTEVIFKPK	VIVYNYPKGI	KAFYMLRND	QKTVAAMDVL	VPKVGELIGG	480
SQREERIDVL	KSRILESLPL	LEPYEWYLDL	RRYGTVKHCG	FGLGFERMIM	FATGLDNIRD	540
VIPFPYPGK	ADL					553

SEQ ID NO: 17 moltype = AA length = 535
 FEATURE Location/Qualifiers
 source 1..535
 mol_type = protein
 organism = Triticum aestivum

SEQUENCE: 17

TSRTRIRAIL	DAGDAMAGER	VVVGWVKTG	RQQKGEFAP	LEVNDGSCQG	NLQVMVDKDV	60
HPLASLTHTG	TSVLVEGVLK	KPPAEAKQRI	ELKVERVIEL	GEVDAAYPL	PKTKITLETL	120
RDFVHLRART	NTIGAVARIR	HQLAYATHSF	FDENGFLYIH	TPIITTSDC	GAGEMFQVTA	180
LFSQAQKVEK	ELKENPAPSE	ADVEAAKLVV	KEKGDVAQAL	KAASKAQEI	TAAVSVLTKA	240
KENVLRVEER	SKLKPGLPKD	DGKIAPENDF	FKRQAFLTVS	GQLQVETVAC	ALSNVYTFGP	300
TFRAENSHTS	RHLAEFWMVE	PEIAFANLQD	DMNCAERYVQ	YLCKWLLKHC	REDMEPMVKH	360
VDKTAIERLE	LVSSTPPQRI	SYTKAVEILE	GVDDKVFENK	EWGIDLASEH	ERYLTEVIFK	420
KPVIVYNYPK	GKAFYMLRLN	DDQKTVAAM	VLVPKVGELI	GGSQREERLD	ILKQRIILDAD	480
LPLEPYEWYL	DLRRFGSVKH	SGFGLGFERM	ILFATGLENI	RDVIPFPYP	GRADL	535

SEQ ID NO: 18 moltype = AA length = 516
 FEATURE Location/Qualifiers
 source 1..516
 mol_type = protein
 organism = Zea mays

SEQUENCE: 18

VVVGWVVRTG	REQKGSFAP	LELSDGSCAA	TLQAIVDAAV	HPLARLTATG	TSVLVEGVK	60
EPPEGTKQNV	ELKVSRLVEV	GEVDAAVYPL	PKVKTLEKL	RDVHLRSRT	NTIGAVARIR	120
HQLAYASHRF	FDENGFLYVH	TPIITTSDC	GAGEMFQVTT	LFSQAQKTEK	ELRENPKPSD	180
SEIEAAKVLV	KEKGDVAQAL	KAASKAQEI	STAVDELNRA	KEIVSKLEER	FKLKPPIPRK	240

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DGSIAFENDF PKRQAFLTVS GQLQVETYAC ALSSVYTFGP TFRAENSHTS RHLAEFWMVE 300
 PEIAPANLQD DMNCAEKYVQ YLCKWLLDHC QEDMEFMVKN YDKSAIERLE LVSSTPFVRI 360
 SYTKAVELLK NVTDKKFDNK VEWGIDLASE HERYLTEDIF KKPVIYVYNP KGIKAPYMRL 420
 NDDDKTVAAM DVLVPKVGEL IGGSQREERL DVLKQRILDA GLPLEPYEWY LDLRRFGSVK 480
 HSGFGLGFER MILFATGMEN IRDVIPPFRY PGRADL 516

SEQ ID NO: 19 moltype = AA length = 544
 FEATURE Location/Qualifiers
 source 1..544
 mol_type = protein
 organism = Beta vulgaris

SEQUENCE: 19
 APFSKRIPIR SILNRSDDGA SLAGQRVTVR GWVRTGRKQN KGALGFIELN DGSCCANLQV 60
 IVESSVCELS QLLSPGSCVS LSGELKSPPO SGNDKQKIEL FADHVIEWVP VDPTEYPLPK 120
 TRIITEYLRD FVHLRPRSS FAAVARIRSE LAYATHTPFR DNGFINVHTP IITIDCEGA 180
 GETFQVTTLI KDAEKLEQEL LENPAPSEAD VEAAKLVVEE KGEVLKLLKA AEASKEEIK 240
 AVGVLMKAKG DLAKLEERAK LQPGIPKKG KVDYAHDFFA RQAFLTVSGQ LQAESFACSL 300
 GSVYTFGPTF RAEHSHTSRH LAEFWMVEPE IAFADLEDDM NCAEAYIKFM CQWLLDRCLD 360
 DMVVIKLFDF NTAIDRLKMW ASTAFVRITY TEAVTLLEQV KDKKFENKVE WGSDLASEHE 420
 RYLTEVIFKK PVIVYNYPKG IKAFYMLRND DGKTVAAAMDV LVPKVGELVG GSQREERYDV 480
 IVKRIVEMGL PLEHYKWYLD LRRYGTAIHA GFGLGFERML LFATGIDNIR DVIPFRYPG 540
 RADL 544

SEQ ID NO: 20 moltype = DNA length = 22
 FEATURE Location/Qualifiers
 source 1..22
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 20
 caccatggct gatgagattg tg 22

SEQ ID NO: 21 moltype = DNA length = 24
 FEATURE Location/Qualifiers
 source 1..24
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 21
 aagatcagct tttccaggat agcg 24

SEQ ID NO: 22 moltype = DNA length = 11
 FEATURE Location/Qualifiers
 source 1..11
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 22
 gctttgagac c 11

SEQ ID NO: 23 moltype = DNA length = 2014
 FEATURE Location/Qualifiers
 source 1..2014
 mol_type = other DNA
 organism = synthetic construct

misc_feature 1..8
 note = 5' adaptor

misc_feature 2004..2014
 note = 3' adaptor

SEQUENCE: 23
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 tgccagacta tgcggcgagg gcatatccat atgatgtacc cgactacgca ggtggtggat 120
 ccggtggggg gtccgggggt tccgctgtga gccaccgcga gtttgaaaaa ggtggcgagg 180
 gcggagggcg cagcggcggg tccgctgtgt cacatcctca gttcgagaaa tcaggctctg 240
 agacaccccg cacttcagaa tcagccactc cagagtccgg gtctatggct gacgagattg 300
 tccgccagc tactcaactc gcggcagtga gcctcgaaaa tgacgggtct acagtacagc 360
 gcgctcaatt cagtaacaga gtgctgatcc gtaccathtt agatcggcca gatggtggcg 420
 cgggccttgc tgggcagact gtccgattg gaggatgggt caaatccggg cgggaccaag 480
 gcaagaggac attttcttct ctggcgggta atgacggaag ttgtcctgct aatctgcagg 540
 tgatggttga tccatcgtc tatgacgtga gcaatttggg agctacgggc acctgtgtca 600
 ccgtcgatgg tgtgcttaag gttccgccta aaggcaaggg gacccaacag cagatcgagc 660
 ttaatgttgt caaagtgatt gatgttggaa cgggtggagc ctcaaaagtat cctctgccta 720
 agaccaagct gacgttagag actctgaggg acgtgtcca ctaaggagc aggactaata 780
 gcatttctgc tgttgcaagg atacgtaatg ctctcgcctt cgcactactc agcttcttcc 840
 aggagcacag tttctgttac attcatacgc cgateatcac cacctcggac tgcgagggtg 900
 cgggtgagat gtttcaagcg accactctta tcaactacac cgagcggctg gagcaagatc 960
 ttatagacaa cccccgccg actgaagcag acgtggaggg agcagagactg atcgctattg 1020
 agagagaaaa cgtggtggct gaattgaaaag cagcaaaagg gagcaaggag gctatcacgg 1080
 cggcagtagc tgagctgaag atcgccaaag aaaccttcgc gcacattgat gaacgctcac 1140

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gattgagggc tggactacc aagaaggacg gcaacataga ttactccaag gattttttcg 1200
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tctccaatgt gtacactttc ggcccagcgt tccggcgcaga aaattcccat acatcgcggc 1320
acctggccga attctggagt gtcgagcctg aaattgcctt tgctgatttg gaagatgaca 1380
tgaactgcgc ggaggcttac gtgaagtaca tgtgcaactg gttgctagag aagtgtctatg 1440
cagatatgga gttgatggcg aagaacttcg acagtggctg catcgacagg cttaaagttag 1500
tagcttcaac cccctttggc cgcataacat atacaaggc catcgagctt cttgaggaag 1560
ccgtcgccaa ggggaaaagag tttgataaca acggtgaatg ggggattgac ctccgctctg 1620
agcatgaaag atatttgaca gaagtgtctt tccaaaagcc actcatcgtg tacaactatc 1680
ccaaaggcat caaggcgctt tacatgcgcc tgaatgatga tgagaaaaca gtcgcccgca 1740
tggacgtcct cgtcccaaaa gttggcgaac tcataggtgg aagccagaga gaggagcgtc 1800
atgatgtcat caaaaagaga attgaagaga tgggctacc catagaacca tacgagtggt 1860
acctggatct caggaggtta ggaacagtca agcactgcgg ctttggattg ggctttgagc 1920
gcatgatttt gtttgcacag ggctcgcaca acatacggga tgmtatccca ttcccgcggc 1980
accctggcaa agctgacctg tgagctttga gacc 2014

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SEQ ID NO: 24      moltype = DNA length = 2014
FEATURE           Location/Qualifiers
source            1..2014
                  mol_type = genomic DNA
                  organism = Zea mays

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SEQUENCE: 24
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ttgcatgtct aagttataaa aaattaccac atattttttt tgtcacactt gtttgaagtg 120
cagtttatct atctttatca atatatttaa actttactct acgaataata taatctatag 180
tactacaata atctcagtgt tttagagaat catataaatg aacagttaga catggcttaa 240
aggacaattg agtatttttg caacaggact ctacagtttt atctttttag tgtgcatgtg 300
ttctcctttt tttttgcaaa tagcttcacc tatataatac ttcacccatt ttattgtac 360
atccatttag ggtttagggg taatggtttt tatagactaa tttttttagt acatctattt 420
tattctattt tagcctctaa attaagaaaa ctaaaactct attttagttt ttttatttaa 480
taatttagat ataaaataga ataaaataaa gtgactaaaa ataaacaaa taccctttaa 540
gaaattaaaa aaactaagga aacatttttc ttgtttcgag tagataatgc cagcctgtta 600
aacgcccgtc acgagctctaa cggacaccaa ccagcgaacc agcagcgtcg cgtcggggca 660
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tccaccgttg gacttgctcc gctgtcggca tccagaaaat gcgtggcgga gcggcagacg 780
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ggaatcctgg gatggctcta gccgttccgc agacgggatc gatttcatga ttttttttgt 1260
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attcatatgc tctaaccttg agttacctat tattataata aacaagtatg ttttataatt 1860
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gccctgcctt catacctat ttatttgctt ggtactgttt cttttgtcga tgcaccacct 1980
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SEQ ID NO: 25      moltype = DNA length = 932
FEATURE           Location/Qualifiers
source            1..932
                  mol_type = genomic DNA
                  organism = Zea mays

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SEQUENCE: 25
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tttaataatg gaccgggtgt gttgtgtgtg cgtactaccg agaactatga caaatcatga 180
ataagtttga tgtttgaaat taaagcctgt gctcattatg ttctgtcttt cagttgtctc 240
ctaattttg cctgcaggta ctggctatct accgtttctt acttaggagg tgtttgaaatg 300
cactaaaact aatagtttagt ggctaaaatt agttaaaca tccaaacacc atagctaata 360
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gctaattcaa ctaacaattt ttagccaact aacaattagt ttcagtgcac tcaaacacc 480
ccttaattgtt aacgtggttc tatctaccga ctccataat atggttgatt gttcggtttg 540
ttgctatgct attgggttct gattgctgct agttcttctg gaatccagaa gttctcgtag 600
tatagctcag attcatatta tttatttgag tgataagtga tccaggttat tactatgtta 660
gctagggttt ttttacaagg ataaattatc tgtgatcata attcttatga aagccttatg 720
ttctcggag gcagtgccat gcaatgcgat acagcaactt gatcacacca gctgaggtag 780

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atacggtaac aaggttctta aatctgttca ccaaatcatt ggagaacaca catacacatt 840
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cttttgcca tgagtcgta ccgcttgaga cc 932
```

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SEQ ID NO: 26      moltype = DNA length = 1341
FEATURE           Location/Qualifiers
source            1..1341
                  mol_type = genomic DNA
                  organism = Cauliflower mosaic virus
```

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SEQUENCE: 26
atctcagaat tccaatccca caaaaatctg agcttaacag cacagttgct cctctcagag 60
cagaatcggg tattcaacac cctcatatca actactacgt tgtgtataac ggtccacatg 120
cgggtatata cgatgactgg ggttgtacaa aggcggcaac aaacggcgtt cccggagttg 180
cacacaagaa atttgccact attacagagg caagagcagc agctgacgcg tacacaacaa 240
gtcagcaaac agacaggttg aacttcatcc ccaaaggaga agctcaactc aagcccaaga 300
gctttgctaa ggccttaaca agcccacca agcaaaaagc ccactggctc acgctaggaa 360
ccaaaagccc cagcagtgat ccagcccca aagagatctc ctttgccccc gagattacaa 420
tggacgattt cctctatctt tacgatctag gaaggaagtt cgaaggtgaa ggtgacgaca 480
ctatgttcac cactgataat gagaaggtta gcctcttcaa tttcagaaa aatctgacc 540
cacagatggt tagagaggcc tacgcagcag gtctcatcaa gacgatctac ccgagtaaca 600
atctccagga gatcaaatc cttcccaaga aggttaaaga tgcagtcaaa agattcagga 660
ctaattgcat caagaacaca gaaagaagca tatttctcaa gatcagaagt actattccag 720
tatggacgat tcaaggcttg cttcataaac caaggcaagt aatagagatt ggagtctcta 780
aaaaggtagt tcctactgaa tctaaggcca tgcattggag ctaagatcca aatcagggat 840
ctaacagAAC tgcgcgtgaa gactggcgaa cagttcctac agagtctttt acgactcaat 900
gacaagaaga aaatctctgt caacatgggtg gagcagcaca ctctggtcta ctccaaaat 960
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gaaaaggaag gtggctccta caaatgccat cattgcgata aaggaaaagg tatcattcaa 1140
gatctctctg ccgacagtggt tcccaaatg ggacccccac ccacgaggag catcgtggaa 1200
aaagaagacg ttccaaccac gtcttcaaa caagtggatt gatgacat ctccactgac 1260
gtaagggatg acgcacaatc ccactatcct tegcaagacc cttctctctat ataaggaagt 1320
tcatttcatt tggagaggac a 1341
```

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SEQ ID NO: 27      moltype = DNA length = 346
FEATURE           Location/Qualifiers
source            1..346
                  mol_type = genomic DNA
                  organism = Cauliflower mosaic virus
```

```
SEQUENCE: 27
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cgataaagga aaggctatca ttcaagatct ctctgccgac agtggtccca aagatggacc 180
cccaccacag aggagcatcg tggaaaaaga agacgttcca accacgtctt caaagcaagt 240
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agacccttcc tctatataag gaagttcatt tcatttggag aggaca 346
```

What is claimed:

1. A plant that lacks root nodules and is engineered to overexpress an asparaginyl-tRNA synthetase 1 (NARS1) protein as compared to a control plant.
2. The plant of claim 1, wherein the plant comprises a construct comprising a heterologous promoter operably linked to a polynucleotide encoding the NARS1 protein.
3. The plant of claim 2, wherein the heterologous promoter is a constitutive promoter.
4. The plant of claim 3, wherein the constitutive promoter is a cauliflower mosaic virus (CaMV) 35S promoter or a ubiquitin promoter.
5. The plant of claim 1, wherein the plant is a non-leguminous plant.
6. The plant of claim 1, wherein the plant is selected from the group consisting of maize, tobacco, hemp, rice, canola, potato, wheat, cotton, and sugar beet.
7. The plant of claim 1, wherein the plant is *Arabidopsis thaliana* or maize.
8. The plant of claim 1, wherein the NARS1 protein is an endogenous NARS1 protein.

9. The plant of claim 8, wherein expression of an endogenous gene encoding the NARS1 protein is upregulated in the plant.

10. The plant of claim 1, wherein the NARS1 protein is a heterologous NARS1 protein.

11. The plant of claim 1, wherein the NARS1 protein is at least 90% identical to an amino acid sequence selected from SEQ ID NOs: 2-19.

12. The plant of claim 1, wherein the NARS1 protein is overexpressed in at least one tissue selected from roots, leaves, fruit, or seeds of the plant.

13. A seed produced by the plant of claim 1.

14. A method of generating a plant that overexpresses a NARS1 protein, the method comprising:

- a) introducing a construct comprising a heterologous promoter operably linked to a polynucleotide encoding the NARS1 protein into a plant cell from a plant that lacks root nodules; and
- b) growing the plant cell into the plant that overexpresses the NARS1 protein.

15. The method of claim **14**, wherein the plant is selected from the group consisting of *Arabidopsis thaliana*, maize, tobacco, hemp, rice, canola, potato, wheat, cotton, and sugar beet.

16. The method of claim **14**, wherein the NARS1 protein is at least 90% identical to an amino acid sequence selected from SEQ ID NOs: 2-19.

17. A plant or seed produced by the method of claim **14**.

18. A method of growing a plant that overexpresses NARS1, the method comprising:

- a) planting the seed of claim **13**; and
- b) growing the seed into a plant that overexpresses NARS1.

19. The method of claim **18**, wherein the plant has an increased growth rate as compared to a control plant.

20. The method of claim **18**, wherein the plant has an increased yield as compared to a control plant.

* * * * *